

## **PhD Thesis**

Per Jensen

# Translocator Protein Imaging with <sup>123</sup>I-CLINDE SPECT

- Method Development and Clinical Research

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**Title:** Translocator Protein Imaging with <sup>123</sup>I-CLINDE SPECT – Method Development and Clinical Research

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### Introduction

Molecular imaging of the translocator protein (TSPO) in the brain provides a unique window into diseases of the brain. It visualises reactive microglia, astrocytes as well as macrophages crossing the blood brain barrier from the blood into the brain parenchyma, which together constitute an immune response.

Since the discovery of TSPO in the late 1970's, it has been linked to a broad range of neurological, psychiatric and oncological diseases. However, TSPO imaging has yet to be implemented clinically. There are several reasons: methodological issues, the cost of molecular imaging, and the clinical relevance for TSPO imaging has not been demonstrated in diagnostics, disease progression or treatment response.

Since 2010, at the Neurobiology Research Unit, TSPO imaging has been conducted using <sup>123</sup>I-CLINDE, a second-generation single photon emission computed tomography (SPECT) TSPO radiotracer.

The major aim of the current work is to advance <sup>123</sup>I-CLINDE SPECT imaging into clinical research of neurological diseases. To reach this aim this thesis presents four studies, arranged like a classical three-stage rocket.

The first part consists of two 'proof of concept' studies demonstrating the feasibility of <sup>123</sup>I-CLINDE SPECT in diverse and challenging clinical settings: the fatal cancer disease glioblastoma and the recently discovered autoimmune disease anti-NMDA receptor encephalitis. Each study points forward to future larger clinical studies.

The middle part focuses on further development and validation of the <sup>123</sup>I-CLINDE SPECT scanning method. This study was performed in collaboration with postdoc Ling Feng. New and importans discoveries about TSPO tracer stability were made and implemented in the final study. The tip of the rocket is a clinical investigative study describing the longitudinal development of the TSPO expression in recovering patients 2-18 weeks after stroke. Here I also had the opportunity to develop experience as a neurologist in training.

I think that the rocket has reached orbit, although much work is still to be done.

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### List of publications

### Study 1

**Jensen, P.**, Kondziella, D., Thomsen, G., Dyssegaard, A., Svarer, C., & Pinborg, L. H. (2015) Anti-NMDAR encephalitis: demonstration of neuroinflammation and the effect of immunotherapy. *Neurology* 

#### Study 2

**Jensen, P.**, Feng, L., Law, I., Svarer, C., Knudsen, G. M., Mikkelsen, J. D., ... Pinborg, L. H. (2015). TSPO imaging in glioblastoma multiforme: A direct comparison between 123ICLINDE-SPECT, 18F-FET PET and gadolinium-enhanced MRI. *Journal of Nuclear Medicine*. https://doi.org/10.2967/jnumed.115.158998

#### Study 3

Feng, L\*., **Jensen, P.\***, Thomsen, G., Dyssegaard, A., Svarer, C., Knudsen, L. V., ... Pinborg, L. H. (2016). The variability of translocator protein signal in brain and blood of genotyped healthy humans using in vivo 123I-CLINDE SPECT imaging – a test-retest study. *Journal of Nuclear Medicine*, jnumed.116.183202.

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#### Study 4

**Jensen P.**, Feng L., Meden P., Thomsen G., Knudsen L. V., Steglich-Arnholm H., Dyssegaard A., Møller K., Thomsen C., Svarer C., Beliveau V., Ozenne B., Mikkelsen J. D., Knudsen G. M., and Pinborg L. H.. Expression, evolution and prognostic value of Translocator protein in middle cerebral artery stroke patients - A longitudinal <sup>123</sup>I-CLINDE-SPECT imaging study. *Un-submitted manuscript* 

### **Abbreviations**

**2TCM** - Two-tissue compartment model

CLINDE - 6-Chloro-2-(4'-123I-Iodophenyl)-3-(N,N-Diethyl)-Imidazo[1,2-a]Pyridine-3-

Acetamide

**COV** - Coefficient of variation

**FET** – Flouro-ethyl-thyrosine

CE-CT - Contrast enhanced Computed tomography

**DAMPs** - Damage associated molecular patterns

GAMs - Glioma associated microglia/macrophages

**GBM** - Glioblastoma multiforme

**Gd-MRI** - Gadolinium enhanced magnetic resonance imaging

**GLS** - General least squares

**HPLC** - High-performance liquid chromatography

ICC - Intraclass correlation coefficient

kDa - Kilo Dalton

LG1 - Leucine-rich glioma inactivated-1

NMDAr - N-methyl-D-aspartate receptor

MRI - Magnetic resonance imaging

**PAMPS** - Pathogen associated molecular patterns

**PBR** - Peripheral benzodiazepine receptor

**PD** - Percentage difference

**ROI** - Region of interest

SD - Standard deviation

**SNR** - Signal to noise ratio

**SQ** - Sørensen-Dice coefficient

SUV - Standard uptake value

**TAC** - Time activity curve

TSPO - Translocator protein

V<sub>T</sub> - Distribution volume

**VDAC** - Voltage dependent anion channel

### **Summary in English**

The translocator protein (TSPO) is upregulated in reactive glial cells and expression is increased in acute and chronic neuroinflammatory conditions. This thesis focuses on the expression of TSPO imaged by <sup>123</sup>I-CLINDE single photon emission computed tomography (SPECT) in four settings: anti-NMDAr encephalitis (study 1), glioma (study 2), healthy volunteers (study 3) and ischemic stroke (study 4).

In study 1, the binding of <sup>123</sup>I-CLINDE to TSPO was investigated in a case of anti-NMDA receptor encephalitis. High binding was demonstrated at the initiation of immunotherapy in cortical and subcortical brain regions similar to the distribution of the NMDA receptor. After 7 weeks of treatments, symptoms had subsided and a second scan revealed almost normalized <sup>123</sup>I-CLINDE binding compared to a healthy control.

In study 2, <sup>123</sup>I-CLINDE-SPECT was investigated in relation to conventional <sup>18</sup> F-Flouro-ethyl-tyrosine positron emission tomography (<sup>18</sup>F-FET PET) and gadolinium-enhanced magnetic resonance imaging (gd-MRI) in three glioma patients. Results were compared to tumor progression on contrast-enhanced structural imaging at follow-up. Results suggested that TSPO imaging may be a predictor of active tumor cell proliferation and tumor progression.

Study 3 is a methodological test-retest study in the variability of <sup>123</sup>I-CLINDE SPECT imaging in healthy volunteers. 18 patients were scanned twice. Results revealed that the <sup>123</sup>I-CLINDE SPECT scanning modality showed better test-retest variability than the first generation PET tracer <sup>11</sup>C-PK11195 and comparable variability to previously tested second generation PET tracers. Furthermore, the study demonstrated the importance of immediate centrifugation of the blood-samples, as <sup>123</sup>I-CLINDE is distributed into blood cells if left in the vial, which affects the test-retest variance considerably.

Finally in study 4, a cohort of 12 patients was studied longitudinally at three time-points after ischemic stroke in the middle cerebral artery territory with <sup>123</sup>I-CLINDE SPECT, structural MRI and clinical rating. TSPO expression in stroke was furthermore compared to a cohort of 10 healthy volunteers. Results revealed an incredibly dynamic TSPO expression that could not be correlated to recovery after stroke, however the overlap between the lesion on structural MRI and the region of high <sup>123</sup>I-CLINDE binding was negatively correlated to the volume of lesion necrosis. Interestingly, regions un-related to the lesion were demonstrated to have lower <sup>123</sup>I-

CLINDE binding to TSPO in patients compared to similar regions in healthy volunteers initially and increased to comparable values at later time-points.

In conclusion, the studies of this thesis demonstrate the feasibility of performing research using <sup>123</sup>I-CLINDE SPECT imaging in a variety of settings including both healthy volunteers and neurological patients.

### Dansk resumé

Translocator proteinet (TSPO) er opreguleret i reaktive gliaceller under akutte samt kroniske neuroinflammatoriske tilstande. Denne afhandling fokuserer på ekspressionen af TSPO målt med den molekylære billeddannende modalitet <sup>123</sup>I-CLINDE SPECT i fire forskellige studier: anti-NMDA receptor (NMDAr) encephalitis (studie 1), glioblastoma multiforme (studie 2), raske forsøgspersoner (studie 3) samt apopleksia cerebri (studie 4).

Det første studie undersøger TSPO ekspression i en patient med anti-NMDAr encephalitis skannet med <sup>123</sup>I-CLINDE SPECT to gange: ved behandlingsstart og efter 7 uger. Den første scanning viste forhøjet binding af <sup>123</sup>I-CLINDE til TSPO i de samme kortikale og subkortikale hjerneregioner, som udtrykker NMDA receptorer. Ved skanning nummer to var patienten bedret betydeligt og <sup>123</sup>I-CLINDE bindingen faldet til næsten samme værdier som hos en rask forsøgsperson.

I studie nummer to blev tre patienter med gliablastoma multiforme undersøgt. Studiet sammenligner <sup>123</sup>I-CLINDE SPECT med to konventionelle billeddannende modaliteter ved denne sygdom: <sup>18</sup>F-Flouro-Ethyl-Tyrosine positron emission tomography samt magnetisk resonans billeddannelse med kontrast (gd-MRI). Derudover bliver resultaterne sammenlignet med tumortilvækst på opfølgende skanning senere i forløbet. Studiet viste, at der er områder i og omkring tumor med høj <sup>123</sup>I-CLINDE binding, og at disse områder udviser potentiale til at prædiktere tumor proliferation og tilvækst.

Studie nummer tre er et metodestudie, der undersøger test-retest variansen af <sup>123</sup>I-CLINDE SPECT hos 16 raske forsøgspersoner. Studiet viste en bedre test-retest varians for <sup>123</sup>I-CLINDE SPECT end førstegenerations TSPO radioliganden PK-11195 og sammenlignelig varians i forhold til tidligere studier af 2. Generations radioliganderne <sup>11</sup>C-PBR28 PET og <sup>11</sup>C-DPA-713 PET. Desuden vises der i dette studie, at det har en betydning for aktiviteten i blodplasma i de blodprøver der bliver brugt til at beregne <sup>123</sup>I-CLINDE bindingen til TSPO, hvis man venter med at centrifuge blodprøverne.

Det sidste og fjerde studie er en longitudinel undersøgelse af 12 patienter skannet tre gange efter blodprop efter iskæmisk apopleksi i arteria cerebri media territoriet. Data bliver sammenholdt med rehabilitering målt gennem forløbet samt TSPO ekspressionen hos 10 raske forsøgspersoner. Her finder vi at TSPO ekspressionen efter blodprop er meget heterogen og dynamisk. Det bliver undersøgt, om TSPO ekspressionen er en markør for rehabiliteringen men

ikke fundet sammenhæng. Derudover ses der initialt lavere <sup>123</sup>I-CLINDE binding i hjerneregioner, som umiddelbart ikke er berørt af, eller anatomisk forbundet med blodproppen når man sammenligner med raske forsøgspersoner. Den afsluttende konklusion på denne afhandling er at <sup>123</sup>I-CLINDE SPECT er en god og brugbar metode til at foretage kliniske forskningsprojekter af TSPO ekspression hos både raske og syge.

### 1. Background

### The Translocator Protein - History, localization, structure and function

The link between the translocator protein (TSPO) and the consequences of its expression is complex, poorly understood, and hence still a subject of research and debate. The following text outlines the central history, structure and cellular functions of TSPO with emphasis on the TSPO expressing cells of the brain.

The first discovery of TSPO was made in the late 1970's by Danish biochemist Claus Bræstrup and colleagues. It was originally named the peripheral benzodiazepine receptor (PBR) because of its discovery in non-brain tissues and affinity to benzodiazepines. Later on, the molecule was also identified in brain tissue (Braestrup & Squires, 1977) and almost thirty years later, the PBR nomenclature was changed to TSPO based on discoveries in structure and molecular function (Papadopoulos et al., 2006).

The tertiary molecular structure of TSPO comprises 5 trans-membrane domains that environ the benzodiazepine binding site (Scarf & Kassiou, 2011). The 18 kDa TSPO molecule is primarily located in the outer mitochondrial membrane at contact sites between inner and outer mitochondrial membranes (Papadopoulos et al., 2006; Scarf & Kassiou, 2011). During active cell proliferation and steroid synthesis TSPO has the ability to form oligomers (Delavoie et al., 2003; Lacapère & Papadopoulos, 2003). It has been demonstrated that TSPO is associated with the mitochondrial membrane located voltage dependent anion channel (VDAC) and several other mitochondrial membrane proteins including peripheral benzodiazepine receptor-associated protein 1 (PRAX-1, steroidogenic acute regulatory protein (StAR), peripheral benzodiazepine receptor-associated protein (PAP7) and diazepam binding inhibitor (DBI) (G.-J. Liu et al., 2014), and that the association with proximal proteins can affect the binding of ligand to TSPO towards both higher and lower affinity (Midzak et al., 2015).

TSPO is an evolutionary ancient protein and expressed throughout all tissues but at the highest concentration in steroid synthesising tissues in body and brain. When performing a <sup>11</sup>C-PBR28 PET scan of the torso and head, it is apparent that TSPO is present in high concentrations in the

heart and lungs (Kreisl et al., 2010). TSPO is also abundant in monocytes and granulocytes (Canat et al., 1993). The name TSPO was changed from PBR because of its involvement in intracellular movement of protoporphyrins and cholesterol (Papadopoulos et al., 2006). However, more recently this function of mammalian TSPO has been taken up for renewed discussion (Zhao et al., 2016). TSPO has been demonstrated to be involved in numerous cellular processes including steroidogenesis, cell growth and proliferation, bile acid synthesis, calcium flow, heme synthesis, mitochondrial respiration and apoptosis (Chen & Guilarte, 2008).

#### TSPO and neuroinflammation

In the healthy brain, TSPO is expressed at low levels (Chen & Guilarte, 2008), but upregulation occurs after damage to brain tissue or invasion of pathogens (Martín et al., 2010). TSPO-upregulation has been paralleled with the concept of neuroinflammation, and used as a molecular target for studying human neuroinflammation in vivo by molecular imaging. In early TSPO imaging literature, emphasis was often exaggerated on TSPO upregulation in microglia, but it is now recognised that astrocytes and invading macrophages also make up a substantial part of the TSPO-signal (Martín et al., 2010). Since TSPO upregulation is not specific to types of immunological cells or their active properties, a change in TSPO expression has to be interpreted in the context of the condition that is studied (G.-J. Liu et al., 2014). A description of the TSPO expressing cells of the brain is given below.

### Microglia

Microglia are often referred to as the resident macrophages of the brain. However, microglia show very unique properties and do not originate from haematopoetic cells in the bone marrow but rather the yolk sac and enter the brain during early development of the brain (Ginhoux et al., 2010).

Under healthy conditions, microglia are distributed throughout the brain tissue in a ramified phenotype, chemically surveying the local extracellular environment with motile processes emitting from the cellular body (Prinz & Priller, 2014). In this state TSPO is only expressed to a small degree (Chen & Guilarte, 2008).

As a response to neuronal damage or pathogens in the extracellular environment, microglia transform into a "reactive" state and move to the lesion site. Microglia show very diverse properties in this reactive state and play a pivotal role orchestrating astrocyte activation and recruitment of peripheral macrophages (Tang, & Feng, 2011). TSPO is highly upregulated in reactive microglia (Albrecht, et al., 2016).

Historically, microglia activation has been paralleled to macrophage polarization with activation into M1 and M2 phenotypes. This paradigm suggests that microglia can activate in a classical pathway in response to pathogens or neuronal damage leading to a M1 pro-inflammatory phenotype producing proinflammatory cytokines and reactive oxygen species. Alternatively, in response to chemokines, microglia activate into a M2 neuroprotective phenotype that promotes tissue remodelling and produces anti-inflammatory cytokines. (Nakagawa & Chiba, 2015; Prinz & Priller, 2014). It is generally accepted that microglia retain the above-mentioned properties, however, the polarization paradigm is disputed both for macrophages and microglia (Ransohoff, 2016).

### **Astrocytes**

Astrocytes are the most diverse resident cells of the brain and play many structural and functional roles under healthy and pathological conditions (Cekanaviciute & Buckwalter, 2016). Like microglia, astrocytes can transition into a high TSPO expressing reactive state in response to neuronal damage and pro-inflammatory cytokines (Cekanaviciute & Buckwalter, 2016). The upregulation of TSPO in reactive astrocytes usually shows a temporally slower and more prolonged upregulation pattern compared to microglia (Boutin & Pinborg, 2015). TSPO upregulation in astrocytes has been linked to the synthesis of the two neusteroids pregnenolone and progesterone (Le Goascogne et al., 2000; Schumacher et al., 2000). Activated astrocytes like microglia regulate the local immune response by releasing cytokines and chemokines in the affected tissue. Furthermore, astrocytes acts as a physical barrier by forming astrocytic scars to wall of sites of neuronal injury (Cekanaviciute & Buckwalter, 2016).

#### **Macrophages**

Peripheral macrophages have the ability to cross the BBB and infiltrate the brain parenchyma in response to injury or infection (Minogue, 2017) and express TSPO (Canat et al., 1993). The degree of macrophage invasion varies greatly depending on the disease and is generally assumed to be larger in diseases with disruption of the BBB (Albrecht et al., 2016), but the role of infiltrating macrophages in neuroinflammation is unclear and confounded by their similarity to activated microglia in both morphology and phenotype (Minogue, 2017).

#### **TSPO** radiotracers

The first generation TSPO radiotracers for molecular imaging were developed in the late 1980's (G.-J. Liu et al., 2014) and since then, more than 50 TSPO tracers have been developed (Ching et al., 2012). One of the first-generation tracers is <sup>11</sup>C-PK11195, which is still widely used for PET imaging today. For SPECT imaging, iodinated first-generation analogues exist such as <sup>123</sup>I-RO5-4864 and <sup>123</sup>I-PK11195 (Gildersleeve et al., 1989; Stevenson et al., 2010). Because of relatively low affinity to TSPO for the first-generation radiotracers, second generation TSPO radiotracers have since been developed. <sup>123</sup>I-CLINDE is a second generation radiotracer for SPECT imaging (Arlicot et al., 2008; Mattner et al., 2008).

Currently, the most widespread second generation PET radiotracers are <sup>11</sup>C-PBR28 and <sup>18</sup>F-DPA714. However, the search for a high affinity radiotracer that is unaffected by the rs6971 polymorphism is still on-going and tracer development is now entering the third generation.

### The rs6971 polymorphism

After the development of second generation radiotracers with higher signal to noise ratio, substantial indefinable differences in binding of ligand to TSPO were measured that could not be reproduced with <sup>11</sup>C-PK11195 especially for the radiotracer <sup>11</sup>C-PBR28 (Kreisl et al., 2010). The cause of the difference remained a mystery until Owen et al. described the effect of the rs6971 polymorphism on the affinity (Owen et al., 2012). The rs6971 single nucleotide polymorphism is an amino acid substitution in the trans-membrane pocket of TSPO at position 147 of Threonine (T) by Alanine (A). This results in a division of subjects into high affinity binders (HABs, TT), mixed affinity binders (MABs, AT) and low affinity binders (LABs, AA). There is large variance in the polymorphism throughout different ethnic populations. In previous studies on white Caucasian populations the incidence of HABs is 65%, MABs 30%, and LABs around 5% (Mizrahi et al., 2012). The rs6971 polymorphism is usually only attributed to differences in the affinity to second generation TSPO tracers, however, an association of the single-nucleotide polymorphism has been made to Bipolar disorder (Colasanti et al., 2013). <sup>123</sup>I-CLINDE is a second generation TSPO tracer and thus sensitive to the rs6971 polymorphism (Feng et al., 2014). The rs6971 polymorphism provides a limitation for quantification in TSPO studies using tracers whose affinity to TSPO are affected, and should be accounted for in the study design and analysis (Owen et al., 2015).

### TSPO imaging studies in diseases of the human brain

Since the link between TSPO and neuroinflammation was drawn in the early 1980's, in vivo TSPO imaging has been performed in a number of the diseases of the brain where inflammation or other causes for TSPO upregulation (e.g. glioma) are known or hypothesised to play a role. Furthermore, TSPO has been studied in a variety of brain diseases with acute inflammation (stroke, encephalitis, multiple sclerosis and tumor) or chronic inflammation (dementia, Alzheimer's disease, schizophrenia). This thesis includes a more in-depth description on TSPO expression in the studied diseases in the 'description of research projects' section.

### 2. Objectives

At the initiation of the current PhD project, eleven subjects had been scanned at our site, and the first methodological paper on the quantification of <sup>123</sup>I-CLINDE SPECT was in the workings (Feng et al., 2014). The main objective of the current thesis was to further advance <sup>123</sup>I-CLINDE SPECT imaging in clinical research. This was done in three ways:

- 1) To perform two proof-of-concept studies with <sup>123</sup>I-CLINDE SPECT imaging in challenging clinical settings:
  - a. To investigate the longitudinal dynamics of the TSPO expression at the initiation of treatment and after clinical recovery in an Anti-N-Methyl-D-Aspartate receptor (Anti-NMDAr) encephalitis patient (Study 1).
  - b. In Glioblastoma multiforme (GBM) patients, to compare baseline <sup>123</sup>I-CLINDE SPECT with gadolinium-enhanced MRI imaging, <sup>18</sup>F-FET PET, and follow-up structural imaging (Study 2).
- 2) To further the validation and development of the <sup>123</sup>I-CLINDE SPECT scanning method by performing a test-retest study in healthy volunteers and testing the effect of blood sample handling (Study 3).
- 3) Finally, to perform a clinical study with longitudinal <sup>123</sup>I-CLINDE SPECT imaging in stroke patients (Study 4) and investigate the temporal evolution of TSPO expression after stoke, compare results to rehabilitation measures and TSPO expression in healthy volunteers. Here the following hypotheses were tested for the stroke patients:
  - a. Increased perilesional binding of <sup>123</sup>I-CLINDE to TSPO at 1-2 weeks after stroke is a biomarker of poor recovery at 25-26 weeks.
  - b. Increased binding of <sup>123</sup>I-CLINDE to TSPO in the ipsilesional thalamus and pons at 1-2 weeks after stroke is a biomarker of poor recovery at 25-26 weeks.
  - c. Increased binding of <sup>123</sup>I-CLINDE to TSPO in the contralesional thalamus at 5-6 weeks after stroke is a biomarker of good recovery at 25-26 weeks.

### 3. Description of the research projects

### Study 1 – Proof of concept: 123 I-CLINDE-SPECT in anti-NMDAr encephalitis

#### **Background**

The autoimmune encephalitis (AIE) subtype Anti-NMDAr encephalitis is a relatively newly discovered, but prevalent antibody-mediated disease with neurologic and psychiatric symptoms associated with the limbic regions of the brain (e.g. amnesia, confusion and epileptic seizures) and a mortality of 14 % (Blaabjerg et al., 2015). The two most prevalent autoimmune encephalitis types anti-NMDAr encephalitis and anti-Leucine-rich glioma-inactivated 1-encephalitis (anti-LG1 encephalitis) comprise 80% of all diagnosed AIE's in Denmark (Blaabjerg et al., 2015). Anti-NMDAr encephalitis is associated with production of antibodies towards the glutamate receptor subtype N-methyl-D-aspartate receptor. An underlying cancer diagnosis is made in 40-50 % of anti-NMDAr encephalitis cases (Titulaer et al., 2013). The first description of Anti-NMDAr encephalitis symptoms was in 2005 and the first correlation between symptoms and antibodies was made in 2007. AIE is generally under-diagnosed which greatly influences treatment response and prognosis. Twenty-four months mortality is 9.4% after anti-NMDAr encephalitis onset and 20% have severe loss of function (Blaabjerg et al., 2015).

The current first line treatment for anti-NMDAr encephalitis is high-dose methylprednisolone (1g a day for five days) in combination with plasmapheresis (5-7 treatments during 10-14 days) or intravenous immunoglobulin (2 g pr. Kg bodyweight distributed over 5 days). (Blaabjerg et al., 2015). Around 50% respond to treatment (Titulaer et al., 2013). Furthermore, symptoms and/or underlying cancer diagnosis are addressed separately.

Previously, TSPO has been demonstrated to be upregulated in two human case of Rasmussen's encephalitis using <sup>11</sup>C-PK11195 PET, demonstrating both focal and diffuse binding increase in the affected hemisphere (Banati et al., 1999), but no previous human TSPO studies on AIE have been published.

#### **Case description**

The patient was a 35-year-old male, originally admitted to the department of psychiatry because of sudden onset of severe psychosis. At the transfer to the department of neurology, the patient suffered from psychosis, amnesia, orofacial dyskenesia, catatonia and autonomic instability. Antibodies for the NMDA receptor were detected in the cerebrospinal fluid (CSF), and the diagnosis of anti-NMDAr encephalitis was confirmed (Dalmau et al., 2011). The patient was treated with plasmapheresis and methylprednisolone and had a good recovery. Seven weeks after treatment, the patient (Titulaer et al., 2013) was back at work part time, despite mild cognitive symptoms. At 6 months follow-up the patient was back to work full time.

### Study design and materials

<sup>123</sup>I-CLINDE SPECT, T1 weighted MRI, genotyping, post processing and kinetic modelling were performed as described in the general methods section. The patient was scanned twice with <sup>123</sup>I-CLINDE SPECT: at 2 days and 7 weeks after initiation of immunotherapy. Results were compared to an age- gender- and genotyped matched (MAB) healthy volunteer.

### Study 2 – Proof of concept: 123 I-CLINDE SPECT in Glioma

### **Background**

Glioblastoma multiforme remains the most frequent and malignant primary CNS tumor with a median overall survival of 15 months from the time of diagnosis in spite of conventional treatment (Stupp et al., 2005). To visualise and quantify tumor progression and treatment response, GBM patients undergo structural imaging with contrast enhanced MRI (gd-MRI) and molecular imaging with 0-(2-<sup>12</sup>F-fluoroethyl]-<sub>L</sub>-tyrosine PET (<sup>18</sup>F-FET PET) (Dunet et al., 2012; Pauleit, 2005). However, it is known that <sup>18</sup>F-FET PET is not entirely glioma-specific and that increased <sup>18</sup>F-FET uptake is also present in astrogliosis, secondary to infection, haematoma, ischemia and as a cause of radiation injury (Floeth et al., 2006; Pichler et al., 2010; Salber et al., 2007; Spaeth et al., 2004).

TSPO has been demonstrated to be expressed in glioma cell lines and the density of TSPO has been positively correlated with WHO malignancy grade (Louis et al., 2016), cell proliferation index and mortality (Miettinen et al., 1995; Vlodavsky & Soustiel, 2007; Winkeler et al., 2012).

In astrocytic brain tumor cell lines, TSPO is expressed primarily by the neoplastic cells and to a small degree in glioma associated microglia/macrophages (GAMs) (Awde et al., 2013; Winkeler et al., 2012). <sup>123</sup>I-CLINDE SPECT has been validated for investigation of TSPO expression in a GL26 mouse glioma model (Tsartsalis et al., 2015) and investigated in glioma patients (Feng et al., 2014). Here, no significant changes in <sup>123</sup>I-CLINDE binding were found in areas of gadolinium leak on gd-MRI leading to the assumption that <sup>123</sup>I-CLINDE binding parameters are not affected by an impaired BBB. The aim of this study was to test the hypothesis that at baseline <sup>123</sup>I-CLINDE SPECT is a predictor of tumor progression at follow-up structural imaging.

#### Study design and demographics

**Patients:** The study included three patients in advanced state of GBM (Table 3.1). All patients had undergone debulking tumor surgery and received radio- and chemotherapy with temozolomide, bevacizumab and irinotecan before inclusion. At baseline, patients were scanned with <sup>123</sup>I-CLINDE SPECT, <sup>18</sup>F-FET PET and gd-MRI within two days. At follow-up, patients 1 and 2 were re-scanned with gd-MRI and patient 3 with contrast-enhanced CT (CE-CT). The current study joined in the advanced stage of a larger study with autologous lymphoid effector cells specific against tumor cells (ALECSAT) which was received by patient 2 and 3. No further patients were included because of the termination of the ALECSAT study.

| 191 |
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| 188 |
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|     |

*Table 3.1.* Overview of glioma patients included in the study. Genotype, rescan modality, time intervals between scans and injected <sup>123</sup>I-CLINDE dose.

### Study 3- Advances in protocol and test-retest variability of 123 I-CLINDE SPECT

### **Background**

Performing a test-retest study is a way to test the reproducibility and reliability of a given modality by performing two identical tests and comparing the results. This is useful for the design of future studies and to compare with other similar methods for validation. A method with high reproducibility is good for performing longitudinal studies, since fewer subjects would be required in the study. However, one should bear in mind that a test-retest study does not give a measure on the specificity or sensitivity of the method. If a biological variance occurs that is not taken into account when measuring, a method with high sensitivity would yield a higher test-retest variance. In the current study, the tested modality was <sup>123</sup>I-CLINDE SPECT scanning in healthy volunteers. The outcome measures are percentage difference (PD), which is an estimate of test-retest reproducibility and tracer reliability measured as intra-class correlation coefficient (ICC) and coefficient of variation (COV) (see methods for study 3 for definitions).

The test-retest variance has previously been examined in the first generation TSPO tracers <sup>11</sup>C-PK1195 (Jučaite et al., 2012) revealing moderate test-retest variability, and in the second generation tracers <sup>11</sup>C-DPA 713 (Coughlin et al., 2014) and <sup>11</sup>C-PBR28 (Collste et al., 2015; Park et al., 2015) with moderate to good test-retest variability.

#### Study design and demographics

Healthy volunteers were screened for the rs6971 genotype before inclusion in order to restrict the inclusion to MABs and HABs. All subjects had a normal neurological examination and blood screen. A total of sixteen healthy controls were included in the study (MABs=8, HABs=8, females=9). Healthy volunteers were <sup>123</sup>I-CLINDE SPECT scanned twice and scanned once with T1-weighted MRI. See table 3.2 for an overview of the patients.

Test-retest variability measures were estimated on both brain SUV and  $V_T$ 's derived from 2TCM.

In the current study, the variability of <sup>123</sup>I-CLINDE was tested independently in both brain and blood. During the study, we discovered that the blood measurements were sensitive to the time from drawing to centrifugation (see the results section for study 3). This is probably because of continuous binding of parent tracer to TSPO in blood cells where monocytes and granulocytes

are known to express abundant levels of TSPO (Canat et al., 1993). This finding made us change the protocol for handling and centrifugation the blood samples. In the old protocol, a batch of blood samples were drawn and stored on ice. They were then transported to the HPLC site and waited for some time before centrifugation. In the worst-case scenario, blood samples were resting on ice for 90 minutes from drawing to centrifugation. This protocol was applied for the first eight healthy volunteers. For the new protocol, a centrifuge was acquired to be in the near vicinity of the scanner, so that blood samples could be centrifuged as fast as possible. This immediate centrifugation was timed to be within 5 minutes from drawing the blood samples to starting the centrifuge. Immediate centrifugation was applied for the remaining eight healthy volunteers Finally, we implemented a population-based approach based on whole blood curves which theoretically should be unaffected by being kept in the vial for any amount of time.

Table 1 Demographic data and methodological information specified by subject

| Subject | Gender | Age<br>(yrs) | Body<br>weight<br>(kg) | TSPO<br>genotype | Injected<br>activity<br>(MBq) | Scan<br>interval<br>(days) | Centrifugation | Purity (%) | Plasma<br>control<br>(%) |
|---------|--------|--------------|------------------------|------------------|-------------------------------|----------------------------|----------------|------------|--------------------------|
| 1       | M      | 61           | 80                     | MAB              | 120.9                         | 21                         | delayed        | 96.1       | 95.4                     |
|         |        |              |                        |                  | 117.8                         |                            |                | 93.5       | 91.4                     |
| 2       | M      | 70           | 92                     | MAB              | 112.2                         | 42                         | delayed        | 95.4       | 95.4                     |
|         |        |              |                        |                  | 123.6                         |                            |                | 94.9       | 93.8                     |
| 3       | M      | 59           | 76.5                   | HAB              | 118.1                         | 35                         | delayed        | 96.4       | 94.7                     |
|         |        |              |                        |                  | 112.0                         |                            |                | 94.9       | 93.2                     |
| 4       | F      | 38           | 110                    | HAB              | 116.3                         | 28                         | delayed        | 96.0       | 95.0                     |
|         |        |              |                        |                  | 114.3                         |                            |                | 90.7       | 88.4                     |
| 5       | F      | 62           | 70                     | HAB              | 119.0                         | 42                         | delayed        | 96.1       | 95.1                     |
|         |        |              |                        |                  | 119.4                         |                            |                | 96.6       | 94.0                     |
| 6       | F      | 58           | 50                     | MAB              | 116.0                         | 28                         | delayed        | 96.1       | 96.2                     |
|         |        |              |                        |                  | 119.7                         |                            |                | 95.3       | 91.4                     |
| 7       | F      | 68           | 87                     | HAB              | 124.8                         | 56                         | delayed        | 92.3       | 90.1                     |
|         |        |              |                        |                  | 115.1                         |                            |                | 95.2       | 90.5                     |
| 8       | M      | 52           | 94                     | HAB              | 118.5                         | 56                         | delayed        | 98.1       | 93.8                     |
|         |        |              |                        |                  | 124.1                         |                            |                | 89.8       | 88.5                     |
| 9       | M      | 42           | 80                     | MAB              | 118.8                         | 28                         | immediate      | 95.7       | 94.6                     |
|         |        |              |                        |                  | 115.8                         |                            |                | 93.4       | 90.3                     |
| 10      | F      | 45           | 86.2                   | MAB              | 117.4                         | 56                         | immediate      | 95.4       | 94.6                     |
|         |        |              |                        |                  | 114.1                         |                            |                | 94.9       | 94.7                     |
| 11      | M      | 35           | 100                    | HAB              | 119.9                         | 56                         | immediate      | 93.0       | 93.2                     |
|         |        |              |                        |                  | 114.4                         |                            |                | 90.7       | 89.9                     |
| 12      | F      | 40           | 73                     | MAB              | 112.0                         | 7                          | immediate      | 92.8       | 91.8                     |
|         |        |              |                        |                  | 130.7                         |                            |                | 94.7       | 94.1                     |
| 13      | M      | 24           | 95.6                   | MAB              | 115.4                         | 14                         | immediate      | 94.4       | 92.2                     |
|         |        |              |                        |                  | 116.0                         |                            |                | 92.0       | 90.2                     |
| 14      | F      | 69           | 83                     | MAB              | 123.0                         | 28                         | immediate      | 94.5       | 92.7                     |
|         |        |              |                        |                  | 127.1                         |                            |                | 94.4       | 90.4                     |
| 15      | F      | 30           | 68                     | HAB              | 105.8                         | 42                         | immediate      | 91.0       | 82.9                     |
|         |        |              |                        |                  | 116.0                         |                            |                | 91.1       | 88.1                     |
| 16      | F      | 34           | 74.2                   | HAB              | 118.4                         | 28                         | immediate      | NA         | NA                       |
|         |        |              |                        |                  | 116.3                         |                            |                | NA         | NA                       |
| Mean    |        | 49±          | 82.5±                  |                  | 117.9±                        | 35±                        |                | 93.4±      | 91.4±                    |
| ±STD    |        | 15           | 14.4                   |                  | 4.9                           | 15                         |                | 1.7        | 3.2                      |

*Table 3.2:* The table displays the data for the healthy volunteers included.

### Study 4 - Clinical study: 123I-CLINDE SPECT in stroke

### **Background**

Stroke is a large burden to patients and society and the major contributor to inherited disability in developed countries ("WHO | The Atlas of Heart Disease and Stroke,"). Molecular imaging of TSPO after stroke is of particular interest because of upregulation in microglia, macrophages and astrocytes, as a response to brain injury (Chen & Guilarte, 2008). This provides a window into the acute and chronic immune-response after cerebral infarction. Since microglia, macrophages and astrocytes possess both detrimental proinflammatory and neuroprotective properties, TSPO expression after stroke has been proposed to be a biological marker of recovery, a possible therapeutic target and a marker of clinical response to future immunomodulatory therapy (Boutin & Pinborg, 2015).

Animal studies have demonstrated that TSPO is expressed both lesionally and perilesionally in strokes with reperfusion, whereas the core infarct has low TSPO expression in strokes with permanent ischemia. Here TSPO was primarily expressed in the infarct margin and perilesional region. Furthermore, investigation into the temporal profile of TSPO expressing cells has revealed that initially in rodent stroke models TSPO is expressed primarily in microglia and macrophages and slightly later on primarily by astrocytes.(Boutin & Pinborg, 2015)

Previous TSPO imaging studies in human stroke have emphasised a handful of brain regions of interest (Feng et al., 2014; A Gerhard et al., 2000; Alexander Gerhard, Schwarz, Myers, Wise, & Banati, 2005; Gulyas et al., 2012; Gulyás et al., 2012; Jensen et al., 2015; Pappata et al., 2000; Price, 2006; Radlinska et al., 2009; Ramsay et al., 1992; Ribeiro et al., 2014; Thiel et al., 2010) (Figure 3.3). Generally the regions can be divided into three subtypes. Lesion-related regions, regions anatomically connected to the lesion and regions un-related to the lesion: Lesion-related regions are the lesion area and the perilesional area. In these areas, TSPO has been shown to be expressed from 72 hours, peaking between 2-3 weeks and then decreasing gradually. TSPO expression in the perilesional area is of special interest since proinflammatory microglia and macrophages potentially have the ability to be detrimental to potentially salvageable brain tissue (Weinstein et al., 2010).

For MCA strokes, anatomically connected regions are pons and thalamus. Previous studies have shown increased TSPO expression in both the ipsilesional (Gerhard et al., 2005; Pappata et al., 2000) and contralesional thalamus (Gerhard et al., 2005). In the study by Radlinska et al. (2009) TSPO expression caudal to the stroke in the pyramidal tract was demonstrated in patients where the pyramidal tract was affected as seen on Diffusion Tensor Imaging (DTI). Later on in the same patient cohort, Gerhard et al. (2010) demonstrated a negative partial correlation between the follow-up function of the affected upper extremity as measured with the Rivermead motor function scale and TSPO upregulation in the pyramidal tract. It was proposed that TSPO upregulation in connected tissues can be a consequence of Wallerian degeneration.

TSPO expression in regions un-related to the lesion are regions supposed to be unaffected by stroke. Typically this would be the cerebellum or contralesional hemisphere and these regions have been used as reference regions in quantification of the binding of tracer to TSPO. Since we use a metabolite corrected input function for quantification of <sup>123</sup>I-CLINDE SPECT (see general methods) we found it interesting to examine these regions. There have been no previous reports on an effect of stroke in humans for these regions.

| Author                     | Year | Number<br>of<br>patients | Stroke                            | Ligand and imaging  | Time of PET                       | Results   |
|----------------------------|------|--------------------------|-----------------------------------|---|-----------------------------------|---|
| Ramsey et al.              | 1992 | 1                        | I MCA                             | [ <sup>11</sup> C]-PK11195 PET                            | 6, 13 and 20<br>days              | Increased TSPO expression in hemisphere of stroke on day 13.  |
| Gerhard et al.             | 2000 | 5                        | 4 MCA<br>1 PCA                    | [ <sup>11</sup> C]-PK11195<br>T1 MRI                      | 5-53 days (1 rescanned)           | Increased activity in the<br>stroke region as shown on<br>T1W MRI   |
| Pappata et al.             | 2000 | 7                        | мса                               | [ <sup>11</sup> C]-PK11195<br>PET +T1 MRI                 | 60-365 days                       | Increased TSPO expression in ipsilesional thalamus, periinfarct areas and capsula interna.  |
| Gerhard et al.             | 2005 | 6                        | 4 MCA<br>1 PCA<br>1 BS            | [ <sup>11</sup> C]-PK11195<br>PET +T1 MRI                 | 3-150 days<br>(2 rescanned)       | Increased TSPO expression up to 28 days. After 150 days extending to connected ipsilateral and contralesional regions.  |
| Price et al.               | 2006 | 4<br>4 C                 | MCA<br>+ control                  | [ <sup>11</sup> C]-PK11195<br>PET +T1,T2 MRI              | 2-30 days<br>(3 reskanned)        | TSPO expression within 72 hours, rising within a week before some reduction by weeks 3-4.   |
| Radlinska,<br>Thiel et al. | 2009 | 21                       | Sub-cortical                      | [ <sup>11</sup> C]-PK11195<br>PET +T1 and DTI<br>MRI (3T) | 2-20 days                         | Specific remote caudal<br>TSPO expression in<br>subjects with subcortical<br>infarct in pyramidal tract.<br>(using DTI to deliniate<br>fibertracts)   |
| Thiel et al                | 2010 | 18                       | Sub-cortical                      | [ <sup>11</sup> C]-PK11195  PET +T1 and DTI MRI (3T)      | Within 3 weeks. 6 month follow up | Correlation between PT damage on DTI (R <sub>FA</sub> ) and PK uptake in the brainstem. No correlation between PK uptake in the infarct or brainstem and motor score (RMFT) at follow-up.                                 |
| Gulyàs et al.              | 2012 | 4                        | Focal                             | [ <sup>11</sup> C]-PK11195                                | 28-55 days                        | Comparative study of [11C]-PK11195 And [11C]-PK11195 And [11C]-Inpocetine. Vinpocetine has possible nonspecific binding or different receptor profile than PK.  |
| Gulyàs et al.              | 2012 | 9                        | Cortical and subcortical          | [ <sup>11</sup> C]vinpocetine                             | 7-98 days                         | Increased TSPO expression<br>in perilesional region for<br>several weeks after stroke   |
| Feng et al                 | 2014 | 9                        | 3 MCA, 1rescan<br>2 PCA, 1 rescan | [ <sup>123</sup> I]CLINDE                                 | 28-252 days                       | Increased TSPO expression in lesional and perilesional region at 4 weeks after stroke, decreasing at later timepoints. One patient with increasing TSPO expression in the contralesional insula at 8 months after stroke. |
| Ribeiro et al              | 2014 | 9                        | МСА                               | [ <sup>18</sup> F]DPA-714                                 | 8-18 days                         | Increased TSPO expression activation in the lesional and perilesional region  |

Table 3.3 An overview of Previous TSPO studies in stroke patients

### Study design and demographics

**Patients:** The investigation of stroke patients was designed as a longitudinal study (Figure 3.1). Patients were recruited from Rigshospitalet and Bispebjerg Hospital, Copenhagen University Hospital, in the first week after stroke then and scanned longitudinally with concomitant <sup>123</sup>I-CLINDE SPECT and structural MRI three times at, i.e. 1-2, 5-6 and 15-18 weeks after stroke. Furthermore, the patients underwent a clinical rating battery of rehabilitation measures at scandays and at 25-26 week follow-up.

Patients were included according to the following in- and exclusion criteria: Inclusion criteria:

- First ever MCA stroke with cortical/subcortical involvement and affection of the contralateral upper extremity
- HAB (TT) or MAB (AT) genotype
- Age above 18

### Exclusion criteria:

- Prior relevant neurological impairment or a severe medical condition that prohibiting participation in the study
- Insufficient ability to participate in the study and its informed consent process
- Contraindications for SPECT or MRI imaging

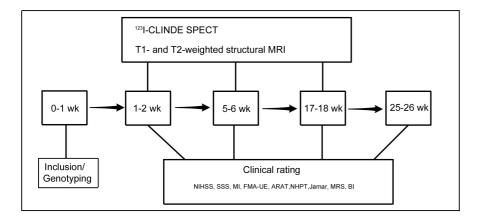


Figure 3.1: Diagram of the study design from study 4.

In total, 12 stroke patients completed the study (Table 3.4). Rehabilitation measures and MRI sequences are detailed in the methods section for study 4.

**Healthy volunteers:** Genotyping revealed that 10 out of 12 stroke-patients were HAB (Table 3.4). Thus, 10 healthy HAB volunteers were recruited and underwent <sup>123</sup>I-CLINDE SPECT and MRI once. Some of the healthy volunteers were also participating in the test-retest study (study 3) and therefore scanned twice. In these cases, the first <sup>123</sup>I-CLINDE SPECT scan was used. See table 3.3 for demographics of the healthy volunteers.

| Volunteer no. | Gender | Age at inclusion (Years) | Injected<br>activity<br>(MBq) |
|---------------|--------|--------------------------|-------------------------------|
| HV 1          | M      | 49                       | 124.9                         |
| HV 2          | M      | 36                       | 119.9                         |
| HV 3          | F      | 48                       | 104.5                         |
| HV 4          | M      | 25                       | 112.5                         |
| HV 5          | F      | 34                       | 115.3                         |
| HV 6          | F      | 44                       | 116.2                         |
| HV 7          | F      | 30                       | 105.8                         |
| HV 8          | F      | 34                       | 118.4                         |
| HV 9          | F      | 24                       | 128.2                         |
| HV 10         | F      | 21                       | 120.9                         |

Table 3.3: Demographics of the healthy volunteers in study 4

| Patient no. | Days<br>after<br>stroke | Gender | Age at inclusion (Years) | lesion<br>side /<br>Artery | Etiology                  | TSPO<br>Genotype | Injected activity (MBq) | Centrifugation |
|-------------|-------------------------|--------|--------------------------|----------------------------|---------------------------|------------------|-------------------------|----------------|
| Pt. 1       | 10                      |        |                          | 1.0                        | TCA II                    |                  |                         | Delayed        |
|             | 38                      | F      | 52                       | Left<br>MCA                | ICA dissection (Trauma)   | HAB              | 113.4                   | Delayed        |
|             | 101                     |        |                          | 1,1011                     | (1144114)                 |                  | 103.1                   | Delayed        |
| Pt. 2       | 10                      |        |                          | T . 0                      | ICA linearitan            |                  | 112.3                   | Delayed        |
|             | 46                      | M      | 63                       | Left<br>MCA                | ICA dissection - (HT)     | HAB              | 102.5                   | Delayed        |
|             | 157                     |        |                          |                            | (111)                     |                  | 117.5                   | Immediate      |
| Pt. 3       | 12                      |        |                          | 1.0                        | Th                        |                  | 135.7                   | Immediate      |
|             | 40                      | M      | 57                       | Left<br>MCA                | Thrombo-<br>embolism (HC) | HAB              | 114.8                   | Immediate      |
|             | 130                     |        |                          | 1,1011                     | emoonsm (rre)             |                  | 126.8                   | Immediate      |
| Pt. 4       | 15                      |        |                          | T . 0                      | Thrombo-                  |                  | 110.9                   | Immediate      |
|             | 43                      | M      | 60                       | Left<br>MCA                | embolism                  | HAB              | 115.9                   | Immediate      |
|             | 139                     |        |                          | 1,1011                     | (HT+HC)                   |                  | 128.3                   | Immediate      |
| Pt. 5       | 11                      |        |                          | D: 14                      | Thrombo-                  |                  | 127.3                   | Immediate      |
|             | 39                      | F      | 42                       | Right<br>MCA               | embolism                  | HAB              | 114.7                   | Immediate      |
|             | 131                     |        |                          | 1,1011                     | (Contraception)           |                  | 108.9                   | Immediate      |
| Pt. 6       | 9                       |        |                          | D: 14                      | TOA 1:                    |                  | 121.5                   | Immediate      |
|             | 35                      | M      | 68                       | Right<br>MCA               | ICA dissection<br>- (HC)  | MAB              | 123.2                   | Immediate      |
|             | 135                     |        |                          | 1,1011                     | (110)                     |                  | 113.0                   | Immediate      |
| Pt. 7       | 7                       |        |                          | D: 14                      | Thrombo-                  |                  | 118.9                   | Immediate      |
|             | 41                      | F      | 72                       | Right<br>MCA               | embolism                  | HAB              | 117.7                   | Immediate      |
|             | 126                     |        |                          | Wich                       | (AF)                      |                  | 108.3                   | Immediate      |
| Pt. 8       | 13                      |        |                          | D: 1.                      | Thrombo-                  |                  | 117.8                   | Immediate      |
|             | 41                      | F      | 68                       | Right<br>MCA               | embolism -                | HAB              | 122.1                   | Immediate      |
|             | 125                     |        |                          | Wich                       | (HC+HT)                   |                  | 103.2                   | Immediate      |
| Pt. 9       | 10                      | F      | 59                       | T 0                        | m 1                       |                  | 137.0                   | Immediate      |
|             | 37                      |        |                          | Left<br>MCA                | Thrombo-<br>embolism (HC) | MAB              | 117.4                   | Immediate      |
|             | 121                     |        |                          | Wich                       | emoonsm (rre)             |                  | 122.1                   | Immediate      |
| Pt. 10      | 13                      |        |                          | D: 1.                      | 10.4 1:                   |                  | 123.7                   | Immediate      |
|             | 41                      | F      | 55                       | Right<br>MCA               | ICA dissection<br>- (HT)  | HAB              | 130.5                   | Immediate      |
|             | 132                     |        |                          | Wich                       | (111)                     |                  | 111.3                   | Immediate      |
| Pt. 11      | 10                      |        |                          | τ Δ                        | Thrombo-                  |                  | 113.6                   | Immediate      |
|             | 38                      | M      | 61                       | Left<br>MCA                | embolism                  | HAB              | 112.8                   | Immediate      |
|             | 122                     |        |                          | 1,1011                     | (HT)                      |                  | 114.4                   | Immediate      |
| Pt. 12      | 10                      | М      |                          |                            | Thrombo-                  |                  | 117.6                   | Immediate      |
|             | 44                      |        | 62                       | Right<br>MCA               | embolism                  | HAB              | 124.9                   | Immediate      |
|             | 122                     |        |                          | 111011                     | (AF)                      |                  | 109.9                   | Immediate      |

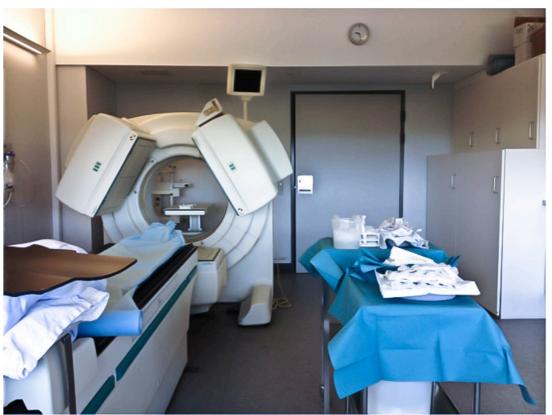
 Table 3.4: Demographics of stroke patients in study 4.

### 4. Methods

### Molecular imaging with SPECT

Single-photon emission computed tomography (SPECT) is a molecular imaging modality using photon-emitting isotopes (e.g. <sup>123</sup>I, <sup>131</sup>I, <sup>99m</sup>Tc, <sup>111</sup>In) injected or inhaled in vivo and measuring the photons from the outside with circulating gamma cameras.

During the acquisition of a SPECT scan, Collimators in front of each of the gamma cameras are selecting projections perpendicular to the head of the gamma camera. After measurement, the tomographic projections are reconstructed into 3-dimensional volumes. Dynamic scans can be obtained by acquiring several volumes consecutively and provide a 4-dimensional image with time as the last dimension. The final post processing part in the quantification of the SPECT signal involves Region of interest (ROI) delineation and quantification of binding using e.g. kinetic modelling. SPECT, albeit lower spatial resolution compared to PET, provides large clinical impact because of the geographically prevalent availability of SPECT tracers since the hospital or clinic is not limited by the need of having a cyclotron available.



**Figure 4.1** A photo of the triple head IRIX camera used at the department of neurology at Rigshospitalet for <sup>123</sup>I-CLINDE SPECT scanning. To the right in the picture the syringes for blood sampling are displayed.

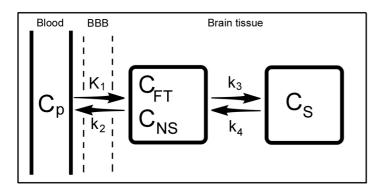
#### **Radiotracers**

A large quantity of radiotracers has been developed to measure a broad range of physiological and biological processes (i.e. blood flow, metabolism, neuroreceptors and transporters). The radioisotope is usually attached to a ligand (e.g. CLINDE), and this ligand has specific affinity to the molecular target of interest. Radioisotopes are administered in tracer doses, meaning that the in vivo concentration occupies a maximum of 10 % of the target sites (Innis et al., 2007). This is enough to measure, but not enough to cause biological effects or saturate the binding sites. Several conditions need to apply for a radiotracer to be optimal for quantification of molecular targets in the central nervous system (CNS):

- organs, the BBB provides an extra barrier for examining the CNS. Generally the ability to passively cross the BBB depends on the lipophilicity of the radiotracer but too high lipophilicity can increase non-specific binding in brain tissue (Waterhouse, 2003). Radiotracers should cross the BBB by passive diffusion, and there should be no active BBB transport of radiotracer. Finally, radiometabolites and radioactive impurities arising from the tracer production should not cross the BBB.
- 2) High affinity and high selectivity to the molecular target as well as low non-specific binding in tissue are required to get optimal signal to noise ratio.
- 3) The tracer must provide suitable pharmacokinetics for quantification of the signal with kinetic modelling.

### 2-tissue compartment modelling

The optimal model for quantification of <sup>123</sup>I-CLINDE SPECT has been demonstrated to be 2-tissue compartment modelling (2TCM) with metabolite corrected arterial input (Feng et al., 2014). This model is often generally interpreted as gold standard for molecular imaging quantification (Gunn et al., 2001). The 2TCM actually consists of three compartments. (Figure 4.2):



**Figure 4.2:** A diagram of the 2-tissue compartment model. BBB=blood brain barrier,  $C_p$  = plasma concentration,  $C_{FT}$  = concentration of free tracer in tissue,  $C_{NS}$  = concentration of non-specifically bound tracer in tissue.  $C_s$  concentration of tracer bound to target.  $K_1$ ,  $k_2$ ,  $k_3$  and  $k_4$  are rate constants of diffusion between compartments.

 $C_p$  is the concentration of tracer in blood plasma. From the blood, tracer is diffusing into the tissue compartment  $(C_1)$ , which consists of both free tracer  $(C_{FT})$  and non-specifically bound  $(C_{NS})$ . The rate constants describing the diffusion of tracer to and from the plasma are called  $K_1$  and  $k_2$ . Once inside the tissue the tracer can also bind specifically to the target of interest, which is interpreted as an individual compartment  $(C_S)$ , and the rate constant describing the binding to and from target are denoted  $(k_3)$  and  $(k_4)$  (Innis et al., 2007).

This system can be described by the following differential equations:

$$\frac{dC_{1}(t)}{dt} = K_{1}C_{p}(t) - (k_{2} + k_{3})C_{1}(t) + k_{4}C_{S}(t)$$
 and 
$$\frac{dC_{2}(t)}{dt} = K_{3}C_{1}(t) - k_{4}C_{S}(t)$$
 where  $C_{1} = C_{FT} + C_{NS}$ 

The metabolite corrected input function ( $C_p(t)$ ) is measured by performing blood sampling and  $C_p+C_{ns}+C_{FT}+C_s$  is measured by the SPECT scanner. The rate constants ( $K_1$ ,  $k_2$ , $k_3$  and  $k_4$ ) are estimated by non-linear fitting. The outcome measure from 2TCM used in <sup>123</sup>I-CLINDE SPECT is the distribution volume ( $V_T$ ), which can be interpreted as the volume fraction of the same amount of tracer in the two tissue compartments divided by the blood compartment. The unit for  $V_T$  is cm<sup>3</sup>/mL and estimated as:

$$V_T = \frac{K_1}{k_2} + \frac{K_1 \times k_2}{k_2 \times k_4}$$

# The <sup>123</sup>I-CLINDE SPECT scanning protocol

All studies are performed in accordance with the Helsinki Declaration, and have been approved by the ethical committee of Copenhagen (H-2-2010-086, amendments 35394, 39319, 42262, 51278, 51900 and 55783) and registered to the Danish Data Protection Agency (j.nr. 2012-58-0004 local j.nr. 30-0739). Prior to beginning the experiment, a signed informed consent form was acquired from the subject to be examined.

#### **SPECT** scanning

<sup>123</sup>I-CLINDE is produced by MAP Medical Technologies (Tikkakoski, Finland) calibrated to 185 MBq at 13:00 hours the next day. <sup>123</sup>I-CLINDE is transported by flight to Copenhagen. The next morning, the tracer is collected at the airport and transported to Rigshospitalet by a professional courier.

A venous catheter is applied and used to draw venous blood samples to be analysed for standard blood parameters at the Department of Biochemistry, Rigshospitalet. For the analysis of radiometabolites in blood, arterial blood samples are needed, and an arterial catheter is therefore placed in the radial artery by an experienced anaesthesiologist.

A standard blood panel is to be analysed by the Department of Biochemistry at Rigshospitalet. Furthermore, blood is drawn at this time in order to perform plasma standard HPLC, and a blood sample is drawn to keep at -80 degrees Celsius for future cytokine evaluation.

Prior to imaging, subjects receive an intravenous injection of 200 mg potassium perchlorate to block thyroidal uptake of free radioiodine.

The SPECT scanner is a triple head IRIX camera (Phillips Medical, Cleveland USA) fitted with low-energy, general all-purpose, parallel-holed (LEGAP) collimators and a 9.5 mm sodium iodine scintillation crystal (Feng et al., 2014) (Figure 4.1).

The subject is positioned between the scanner heads and a headband is applied to fixate the head during the scan. Between 110-130 MBq <sup>123</sup>I-CLINDE is injected into the brachial vein simultaneously with scanner start. The scanner acquires images in two stages, the first stage is 20 minutes consisting of ten 2-minute frames and the second 70-minute stage consists of seven 10-minute frames. In total, a 90 minute dynamic scan is acquired. The first scans we performed (used in study 2) were acquired with a longer second stage of 130 minutes and consequently a total scan-time of 150 minutes. While scanning, 21 blood samples are drawn from the radial artery catheter to count the activity in whole blood and plasma, and eight samples are drawn to perform metabolite analysis using high-performance liquid chromatography (HPLC) by a trained pharmacist.

# Post processing of <sup>123</sup>I-CLINDE SPECT images and blood measures.

The post processing of <sup>123</sup>I-CLINDE SPECT data has changed as the protocol has been evolved and refined during the studies.

To reconstruct the SPECT images and quantify the TSPO signal in different brain regions, the measured raw data undergo a series of post processing steps:

- SPECT images are reconstructed using filtered back projection method for each scan stage and afterwards combined to a single dynamic 90 minute scan.
- Co-registration of the concentration weighted 3-dimensional SPECT image and the T1
  MRIs are performed manually using Interactive Image Overlay (IIO) (Willendrup et al.,
  2004).
- Regions of Interest (ROIs) are delineated from the coregistered structural MRI and SPECT images depending on the specific study. Automatic delineation of regions is performed using the PVE-LAB processing pipeline (Svarer et al., 2005). Time activity curves (TACs) are then extracted from the ROIs.
- Blood TACs are prepared from plasma and whole-blood counts.
- 2-TCM is performed using PMOD 3.0 (PMOD Technologies Inc., Switzerland).

- Firstly, a Hill function (study 1) or an exponential (study 3 and 4) or is fitted to
  the slope of the metabolite measurements. This estimated function is used to
  model the metabolite corrected input function from the plasma activity.
- $\circ$  Secondly, the 2-TCM model is fitted to each brain TAC data using non-linear fitting. Rate constants (K<sub>1</sub>, k<sub>2</sub>, k<sub>3</sub> and k<sub>4</sub>) are estimated for each brain region and V<sub>T</sub> measures are calculated.

# Magnetic resonance imaging

For safety reasons, a standard MRI safety questionnaire is filled out for each subject prior to scanning. Structural MRI was performed on all subjects using either a Siemens Verio 1.5 Tesla scanner or a Siemens Prisma 3 Tesla scanner (Siemens, Erlangen). All subjects received T1 and T2 weighted images and gadolinium-enhanced T1 weighted images was also performed in study 1 and 2.

# Methods used in study 1

All methods used in this study are described in the "general methods section". See the study design for study 1 in the 'Descriptions of research projects' section.

# Methods used in study 2

## Acquisition and reconstruction of <sup>18</sup>F-FET PET:

All patients fasted for at least six hours before <sup>18</sup>F-FET injection. A single-frame static PET acquisition was performed on a 64-slice-CT Biograph TruePoint PET/CT scanner (Siemens, Erlangen, Germany) 20-40 minutes after intravenous injection of 200 MBq <sup>18</sup>F-FET and attenuation-corrected using a low dose CT acquired immediately before the PET scan. Subsequently, PET images were corrected for scatter and dead time and reconstructed with an ordered subset maximization three-dimensional algorithm (6 iterations, 16 subsets) and a 5 mm Gaussian filter.

#### Regions of interest:

**SPECT:** To standardize glioma-related high <sup>123</sup>I-CLINDE binding ROIs between MAB and HAB, a cerebellar reference approach was applied by firstly defining three adjacent cerebellar regions manually on the coregistered structural MRI image and then estimating the mean

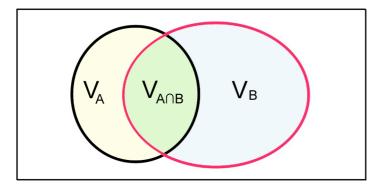
cerebellar count on the SPECT image. Secondly, glioma-related high <sup>123</sup>I-CLINDE binding regions were selected by automatically drawing an iso-contour around voxels above 1.5 times the mean cerebellar count.

**PET:** For [<sup>18</sup>F]FET-PET tumor region delineation, a 3D crescent shaped background ROI encompassing the activity above 70 % of maximum was delineated in healthy appearing grey and white matter of four contiguous brain slices above the insula in the contralateral hemisphere to the tumor. The high uptake [<sup>18</sup>F]FET-PET ROI was auto-contoured in 3D defining tumor tissue at a threshold of above 1.6 of mean SUV (standardized uptake value) in the background ROI. The ROI definition was performed using the Syngo-TrueD software tool (Siemens). This region delineation method is adapted from Pauleit et al. (2005).

**Structural Images:** Structural tumor ROIs were delineated by outlining the contrast-enhanced areas on gd-MRI or CE-CT. Tumor progression ROIs were defined as the additional contrast-enhanced area from baseline to follow-up scan.

#### The Sørensen-Dice index

The Sørensen-Dice index (SQ) is a measure of similarity between the overlap of two volumes (or ROIs) in space. See Figure 4.3



**Figure 4.3:** Individual and overlapping volumes  $V_A$  and  $V_B$  is the total volumes of each region and  $V_{A \cap B}$  is the common or "overlapping" volume of  $V_A$  and  $V_B$ .

The SQ is defined as.

$$SQ = \frac{2V_{A \cap B}}{V_A + V_B}$$

with  $\cap$  being the symbol for common volume.

A SQ of 1 depicts total similarity and a SQ of 0 depicts no similarity between ROIs.

#### Methods used in study 3

Several measures of test-retest variability were applied in this study. The study also implements a population based approach to rescue the datasets with delayed centrifugation of blood samples. The population-based data-rescue method is only addressed shortly in the current thesis, as its concentration and data-analysis primarily were done by Ling Feng.

#### Regions

In this study, mSUV, mSUV $_{60-90}$  and  $V_T$  were investigated in the following ROIs: Midbrain, Thalamus, Pons, Pallidostriatum, Hippocampus, Cerebellum and Neocortex. ROIs were chosen for comparison with previous test-retest studies in TSPO tracers.

#### Brain uptake

Brain uptake was estimated separately as the area under the curve of the standard uptake value (mSUV) derived from the brain TAC of the investigated ROIs. mSUV's were estimated for the full 90 minutes and from 60-90 minute mSUV<sub>60-90</sub>.

#### **Blood uptake**

To test the effect of delayed versus immediate centrifugation and HAB versus MAB in blood, MSUV's were calculated for the plasma to whole blood ratio and parent fraction.

#### Percentage difference

Percentage difference (PD) is a measure of reproducibility and is calculated for each subject as the test-retest difference divided by the average between test and retest. In study 3, calculations of PD were performed on both brain and blood mSUVs and  $V_T$ 's. Furthermore the absolute PD was calculated for brain mSUVs and  $V_T$ 's, to compare results with previous studies.

#### Intraclass correlation coefficient

The intraclass correlation coefficient (ICC) is a reliability measure, as it estimates the withinsubject variability relative to the between subject reliability. In a test-retest situation the ICC can be interpreted as whether the variation of measures are derived from the measured group of subjects or from the variation of the measures in individual subject. To calculate this measure, firstly the deviation of measures from the mean is estimated the as the mean sum of squares between subjects (MSS<sub>btw</sub>) and within subjects (MSS<sub>within</sub>):

$$MSS_{btw} = \frac{1}{N-1} \sum_{i=1}^{N} (\overline{V}_{Ti} - \overline{V}_{T})^{2}$$

$$MSS_{within} = \frac{1}{N} \sum_{i=1}^{N} \sum_{j=1}^{k} (V_{Tij} - \overline{V}_{Ti})^2$$

 $\overline{V}_{Ti}$ : Mean of test and retest measures of subject i,  $\overline{V}_{T}$ : mean of all observations.  $V_{Tij}$ : one of the measures of subject i. N: the number of subjects.

From this this ICC can be calculated as follows:

$$ICC = \frac{MSS_{btw} - MSS_{within}}{MSS_{btw} + (k-1)MSS_{within}}$$

k: number of repeated measures (k=2).

If  $MSS_{btw} = 0$  there is no variation between subjects, thus all variance is described by the within subject variation, there is no reliability, and ICC=-1.

If  $MSS_{within}$ =0 there is no variation within subjects (test and retest values are identical), thus all variance is described by between subject variation, there is maximum reliability, and ICC=1.

#### Coefficient of variation

The coefficient of variation (COV) is the ratio between the standard deviation and the mean.

$$COV = \frac{\sqrt{\frac{1}{n \times t} \times \sum_{n,t} (x_{n,t} - m)^2}}{m}$$

where the mean

$$m = \frac{1}{n \times t} x \sum_{n,t} x_{n,t}$$

n is the number of subjects, t is time point and  $x_{n,t}$  is measure per patient and point of time. COV is calculated on all observations and does not account for the fact that some of the observations are test and retest of the same subjects.

The test-retest variation measures PD, ICC and COV are affected by many factors. The quantification of a SPECT scan relies on several independent measures: Activity in blood and plasma samples, HPLC analysis and the SPECT measure. Furthermore a biological intrasubject variation often occurs (e.g. time of day) and finally, for ICC and COV the intersubject variation also affects the measure.

Previous studies testing the reliability of second generation TSPO tracers (Collste et al., 2015; Coughlin et al., 2014; Park et al., 2015) did not report to have genotyped subjects for the rs6971 polymorphism and thus calculated the ICC value for LABs, MABs and HABs separately. Since binder-status is known to affect the affinity to TSPO for second generation TSPO tracers (David R Owen et al., 2012), bulking all genotypes into one group will increase the *MSS*<sub>btw</sub> relative to the *MSS*<sub>within</sub> and consequently artificially increase the ICC value. In a clinical study it is not possible to compare HABs and MABs without taking the difference in binding of tracer to TSPO into account. Thus, in study 3 we calculated ICC values for the whole group to compare with previous studies, and separately for HABs and MABs to give a more valid ICC estimate of the tracer.

#### General least squares model

A general least squares (GLS) model is similar to a standard linear correlation model, but can be modified to account for complex covariance structures (e.g. repeated measures). In study 3, GLS model was applied to test the effect of the following:

- Test versus retest
- Immediate centrifugation versus delayed centrifugation
- HAB versus MAB binder-status

#### Methods used in study 4

#### Structural MRI sequences

Stroke patients underwent T1 and T2 weighted structural MRI at all scans using a 3 Tesla Siemens Prisma MRI scanner (Siemens, Erlangen).

#### **Delineation of ROIs**

The following ROIs were delineated for the stroke patients:

- Stroke related ROIs: Structural T2-weighted lesion, <sup>123</sup>I-CLINDE lesion, <sup>123</sup>I-CLINDE perilesional.
- Connected anatomical ROIs: Ipsi- and perilesional thalamus and Pons.
- Anatomical ROIs unrelated to the stroke: Ipsilesional cerebellum and contralesional occipital cortex.

For the healthy volunteers, anatomical ROIs were delineated for occipital cortex and cerebellum. The structural lesion ROI was delineated on the first t2-weighted MRI (scan 1).

The <sup>123</sup>I-CLINDE lesion ROI was delineated on the first SPECT scan (scan 1) by automatically selecting voxels in the vicinity of the stroke with a value above 1.5 times the mean cerebellar <sup>123</sup>I-CLINDE uptake.

The Perilesional SPECT region was delineated by subtracting the T2-weighted lesion ROI from the <sup>123</sup>I-CLINDE lesion ROI.

Automatic delineation of anatomical ROIs is described in the general methods section.

#### Rehabilitation measures

The rehabilitation measures in this study can be divided into three categories:

- Neurological impairment rating scales
  - National Institute of Health Stroke Scale (NIHSS)
  - Scandinavian Stroke Scale (SSS)
  - o Fugl-Meyer Assessment of the Upper Extremity (FMA-UE)
  - Motricity Index (MI)
- Functional rating scales of the upper extremity
  - o Action Research Arm Test (ARAT)
  - o Nine-Hole Peg Test (NHPT)
  - o Grip strength (GS)

- Disability rating scales
  - o Barthel Index (BI)
  - Modified Ranking scale (MRS)

Neurological impairment rating scales are clinically used tests that estimate the grade of functional impairment from a neurological deficit perspective (e.g. paresis, sensory loss, aphasia). Functional test is a test of impairment by making the patient perform different tasks and rating or measuring the performance. Disability rating scales are an estimate of the loss of function in daily life.

#### Statistical analysis

Testing the TSPO expression on absolute values is limited for second-generation TSPO tracers by the rs6971 polymorphism's effect on affinity of tracer to TSPO. Having  $V_T$ 's as outcome measure for the TSPO expression, there is no exact workaround for this issue (Owen et al., 2015). When studying longitudinal development one can calculate the relative differences as PD between scans. However, since  $V_T$  is a measure of the activity difference between total tissue  $(C_{FT}+C_{NS}+C_s)$  and blood  $(C_P)$ , comparing PD measures in pooled populations of genotypes can be biased by free tracer and non-specific binding  $(C_{FT}+C_{NS})$  which are not as directly affected by the affinity to the tracer as the specifically bound  $(C_S)$  part.

Comparison with healthy volunteers:

In this study, ten of the stroke patients were HABs and two were MABs. Therefore, we decided to do the comparison between stroke and healthy volunteers by comparing the ten stroke HABs at scan 1,2 and 3 with ten healthy volunteer HABs were compared to the  $V_T$ 's of cerebellum and occipital cortex in healthy volunteers using Mann-Whitney U tests.

Longitudinal evolution of the TSPO signal in stroke patients:

This was done in two ways. In the ten HAB patients by performing repeated measures ANOVA on all three scans. In all twelve patients, by calculating the PD between scans 1 and 3 and performing a one-sample two-sided t-test.

Correlations between the TSPO signal and rehabilitation measures and lesion necrosis were estimated by performing the following Pearson correlations:

- Initial V<sub>T</sub> was correlated with PD of rehabilitation measures between scan 1 and 6 months follow-up.
- Overlap percentage of the T2-weighted lesion ROI by the CLINDE-Lesion ROI was correlated with the volume of lesion necrosis at scan 3 (15-18 weeks after stroke).

PD of rehabilitation measures were calculated as:

Repeated measure ANOVA, t-test, Pearson correlation, and Mann-Whitney U tests were performed using Prism 6.0c (Graphpad Software Inc.). Percentage difference was calculated using Matlab (Mathworks Inc.).

# 5. Results

## Results - study 1

At the beginning of immunotherapy, the patient had high binding of  $^{123}$ I-CLINDE to TSPO in the neocortex ( $V_T = 6.3 \text{ mL/cm}^3$ ) and after 7 weeks the binding of  $^{123}$ I-CLINDE to TSPO had decreased and almost normalised ( $V_T = 4.0 \text{ mL/cm}^3$ ) compared to the healthy volunteer MAB ( $V_T = 3.0 \text{ mL/cm}^3$ ).

# Results - study 2

Results are shown in table 5.1.

#### **Baseline**

VOIs of increased <sup>123</sup>I-CLINDE binding and <sup>18</sup>F-FET uptake showed varying degrees of overlap (12-42 %). In all cases, baseline gadolinium-MRI VOIs showed greater similarity to <sup>18</sup>F-FET VOIs (SQ 0.54, 0.69 and 0.40) compared to <sup>123</sup>I-CLINDE VOIs (SQ 0.05, 0.36 and 0.18). The percentage overlap of gadolinium-MRI baseline VOIs was greater for <sup>18</sup>F-FET VOIs (79-93%) than for <sup>123</sup>I-CLINDE VOIs (15-30%).

## Follow-up

In the two patients where follow-up tumor progression volumes were estimated, the volume with tumor progression at follow-up compared to baseline overlapped to a greater extent with <sup>123</sup>I-CLINDE VOIs than with <sup>18</sup>F-FET VOIs (21% vs. 8 % and 72% vs. 55%).

|   | CLINDE-<br>SPECT  |  | MRI-  | MRI/CT - contrast  |  |  |
|---|---|--|---|--|--|--|
| FET-PET                                 |   | FET-PET∩ CLINDE-SPECT  |   | (follow up -<br>baseline)  |  |  |
|   |   |  | (baseline)  |  |  |  |
| 17.3                                    | 36.6  | 2.0 (12%)  | 7.2   | 97.5   |  |  |
| 196.9                                   | 97.9  | 62.6 (32%)   | 150.5   | NA   |  |  |
| 180.0                                   | 140.7   | 75.0 (42%)   | 60.7  | 44.8   |  |  |
|   |   |  |   |  |  |  |
| FET-PET ∩ MRI-contrast<br>baseline (ml) |   | $\textbf{CLINDE-SPECT} \cap \textbf{MRI-contrast}$   | 90  | SQ <sub>SPECT+MRI</sub>  |  |  |
|   |   | baseline (ml)  | SQ PET+MRI  |  |  |  |
| 6.7 (93%)                               |   | 1.1 (15%)  | 0.54  | 0.05   |  |  |
| 120.1 (80%)                             |   | 44.4 (30%)   | 0.69  | 0.36   |  |  |
| 48.0(79%)                               |   | 18.3 (30%)   | 0.4   | 0.18   |  |  |
|   |   |  |   |  |  |  |
| FET-PET ∩ MRI/CT-                       |   | CLINDE-SPECT $\cap$ MRI/CT-  |   |  |  |  |
| contrast (follow up - baseline)         |   | contrast (follow up - baseline)  |   |  |  |  |
| 7.7 (                                   | (8%)  | 20.9 (21%)   | <del></del>   |  |  |  |
| 24.64                                   | (550/)  | 22.2 (720/)  |   |  |  |  |
|   | 196.9 180.0  FET-PET () baselin  6.7 ( 120.1 48.0(  FET-PET ( contrast (follow) | FET-PET SPECT  17.3 36.6 196.9 97.9 180.0 140.7  FET-PET ∩ MRI-contrast baseline (ml)  6.7 (93%) 120.1 (80%) 48.0(79%)  FET-PET ∩ MRI/CT-contrast (follow up - baseline)  7.7 (8%) | FET-PET         SPECT         FET-PET ∩ CLINDE-SPECT           17.3         36.6         2.0 (12%)           196.9         97.9         62.6 (32%)           180.0         140.7         75.0 (42%)           FET-PET ∩ MRI-contrast baseline (ml)           6.7 (93%)         1.1 (15%)           120.1 (80%)         44.4 (30%)           48.0(79%)         18.3 (30%)           FET-PET ∩ MRI/CT-contrast (follow up - baseline) | FET-PET         CLINDE-SPECT SPECT         FET-PET ∩ CLINDE-SPECT (baseline)         contrast (baseline)           17.3         36.6         2.0 (12%)         7.2           196.9         97.9         62.6 (32%)         150.5           180.0         140.7         75.0 (42%)         60.7           FET-PET ∩ MRI-contrast baseline (ml)         CLINDE-SPECT ∩ MRI-contrast (ml)         SQ PET+MRI           6.7 (93%)         1.1 (15%)         0.54           120.1 (80%)         44.4 (30%)         0.69           48.0(79%)         18.3 (30%)         0.4           FET-PET ∩ MRI/CT- CLINDE-SPECT ∩ MRI/CT- contrast (follow up - baseline)           7.7 (8%)         20.9 (21%) |  |  |

**Table 5.1.** (A): volume sizes for ROIs of high  $^{18}F$ -FET uptake  $^{123}I$ -CLINDE binding and tumor ROIs on contrast-enhanced structural imaging at baseline and follow-up. Furthermore, common volumes between  $^{18}F$ -FET PET and  $^{123}I$ -CLINDE SPECT (FET PET  $\cap$  CLINDE SPECT) and their percentage overlap with  $^{18}F$ -FET ROIs as reference is calculated.

- (B): Baseline common volumes of FET PET  $\cap$  gd-MRI and CLINDE SPECT  $\cap$  gd-MRI common along with Sørensen-Dice coefficients (SQ).
- (C): Follow-up common volumes of FET PET  $\cap$  gd-MRI and CLINDE SPECT  $\cap$  gd-MRI common volumes are given along with percentage overlap (first mentioned modality as reference).

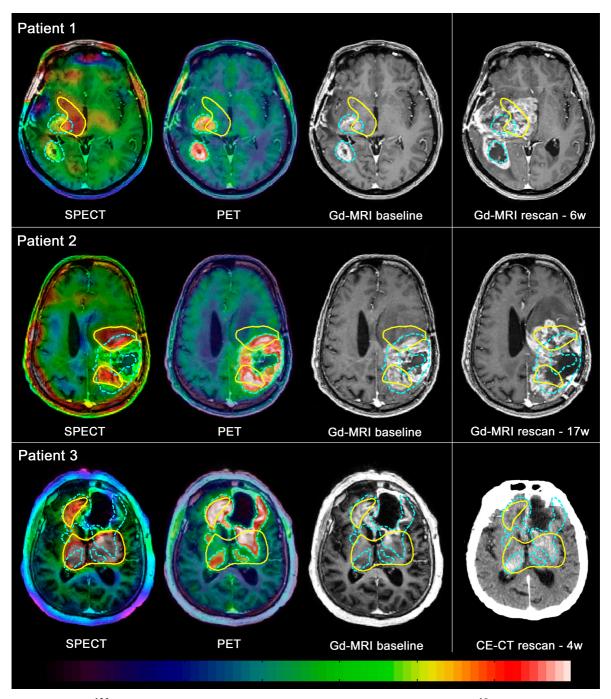


Figure 5.1. <sup>123</sup>I-CLINDE SPECT weighted mean images (first column), <sup>18</sup>F-FET PET summed scans (second column) and gadolinium-enhanced T1 weighted MRI scans at baseline (third column) for three glioma patients. The last column displays gadolinium enhanced T1 weighted MRI for patient 1 and 2 at follow-up, and follow-up contrast enhanced structural CT for patient 3. Yellow ROIs delineate high <sup>23</sup>I-CLINDE binding areas and blue ROIs delineates high <sup>18</sup>F-FET uptake. Comparing baseline scans with follow-up, visually reveals that the glioma predominantly expands in high <sup>123</sup>I-CLINDE binding areas compared to high <sup>18</sup>F-FET uptake areas.

# Results - study 3

# Test-retest variability of brain uptake and distribution volumes and the effect of genotype on variability measures

Addressing the brain uptake (mSUV), we found high correlation between test and retest mSUV for the same subjects, with correlation coefficients of  $0.86\pm0.07$  (p<0.05, N=16).

On average (GLS model, age and gender adjusted), the PD was -3.6±8.1% for MABs and -7.0±7.8% for HABs. No significant PD difference in mSUV between MABs and HABs was found. ICC values for mSUV of MABs and HABs are given in table 5.2

COV was 20.0±2.3% in MABs and 9.2±2.0% in HABs.

A significant difference between MABs and HABs was found both in ICC (p= 2.29e-05) and COV (p=1.38e-05) with a MAB:HAB ratio of 1.8 for ICC and 2.2 for COV.

Test-retest variability measures for the  $V_T$  estimates were calculated both separately and combined with regard to the eight immediately centrifuged and the eight delayed centrifuged population adjusted datasets. (Table 5.2)

On average Test-retest PD of  $V_T$ 's for all subjects across 7 ROIs was -12.1 $\pm$ 8.3% for HABs and -11.7 $\pm$ 8.6% for MABs when evaluated as a repeated measure in an age and gender adjusted GLS.

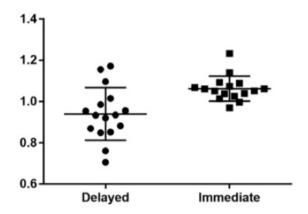
The average COV of  $V_T$ 's was -12.1 $\pm$ 8.3% for HABs and -11.7 $\pm$ 8.6% for MABs. ICC values are presented for each region in table 5.2.

|                 | V <sub>T</sub> using population-adjusted<br>input function of delayed<br>centrifugation N=8 |                    | $V_T$ of immediate centrifugation N=8 |                 | V <sub>⊤</sub> of all subjects N=16 |      |                 |                    | Brain uptake N=16 |                |            |                 |                    |               |               |               |
|-----------------|---|--------------------|---------------------------------------|-----------------|-------------------------------------|------|-----------------|--------------------|-------------------|----------------|------------|-----------------|--------------------|---------------|---------------|---------------|
| Regions         | PD (%)  | Absolute<br>PD (%) | ICC                                   | PD (%)          | Absolute<br>PD (%)                  | ICC  | PD (%)          | Absolute<br>PD (%) | ICC of<br>MABs    | ICC of<br>HABs | ICC of all | PD (%)          | Absolute<br>PD (%) | ICC of<br>MAB | ICC of<br>HAB | ICC<br>of all |
| Midbrain        | $2.8 \pm 14.5$  | 12.4               | 0.96                                  | -1.5 ± 15.6     | 11.8                                | 0.86 | $0.6 \pm 14.8$  | 12.1               | 0.41              | 0.91           | 0.93       | -2.5 ± 11.5     | 10.1               | 0.72          | 0.39          | 0.71          |
| Thalamus        | $11.0 \pm 10.5$   | 12.8               | 0.94                                  | -12.3 ± 13.3    | 12.7                                | 0.86 | -0.7 ± 16.7     | 12.8               | 0.68              | 0.84           | 0.90       | -4.7 ± 14.6     | 11.6               | 0.74          | 0.53          | 0.71          |
| Pons            | -2.3 ± 14.2   | 9.6                | 0.90                                  | $-8.2 \pm 23.8$ | 18.9                                | 0.72 | -5.3 ± 19.2     | 14.2               | 0.54              | 0.60           | 0.82       | -2.7 ± 12.4     | 11.0               | 0.81          | 0.52          | 0.77          |
| Pallidostriatum | $5.8 \pm 21.9$  | 17.6               | 0.85                                  | $-7.0 \pm 19.0$ | 14.1                                | 0.87 | $-0.6 \pm 20.9$ | 15.8               | 0.22              | 0.79           | 0.85       | $-2.2 \pm 13.7$ | 11.1               | 0.79          | 0.50          | 0.74          |
| Hippocampus     | $14.9 \pm 17.3$   | 15.7               | 0.75                                  | $-5.9 \pm 11.8$ | 9.7                                 | 0.91 | 4.5 ± 17.9      | 12.7               | 0.58              | 0.78           | 0.83       | -2.0 ± 15.2     | 12.3               | 0.57          | 0.23          | 0.56          |
| Cerebellum      | $9.6 \pm 9.4$   | 10.2               | 0.94                                  | $-4.9 \pm 10.0$ | 8.2                                 | 0.96 | $2.3 \pm 12.0$  | 9.2                | 0.76              | 0.88           | 0.95       | $-1.5 \pm 9.4$  | 7.8                | 0.90          | 0.45          | 0.87          |
| Neocortex       | $5.8 \pm 12.6$  | 10.2               | 0.92                                  | $-3.2 \pm 12.8$ | 10.5                                | 0.92 | $1.3 \pm 13.2$  | 10.4               | 0.88              | 0.79           | 0.92       | $-2.8 \pm 9.6$  | 8.5                | 0.91          | 0.57          | 0.85          |

**Table 5.2:** Results study 3 - Test-retest variability measures of  $V_T$ s and mSUV

#### Effect of centrifugation on plasma to whole-blood ratio

Applying a GLS model adjusted for genotype on the mSUV of the plasma to whole-blood ratio (PoB) showed an increase from 0.95 to 1.07 (p=0.0003) when changing from delayed to immediate centrifugation. This results in a 12.4% decrease in PoB when delaying the time to centrifugation (Figure 5.2). This shift may be caused by radioactivity leaving the plasma-phase as the sample is waiting in the vial, and will consequently cause the input function to be underestimated and the  $V_T$  to be over-estimated.



*Figure 5.2 Effect of time to centrifugation on the plasma to whole blood ratio.* 

## Effect of genotype on the parent fraction, brain uptake and distribution volumes

Parent fraction: As mentioned in the study-design, this study only included MAB and HAB genotypes. The testing of genotype differences in mSUV of the parent fraction was performed on the immediately centrifuged group applying a GLS model adjusted for age and gender.

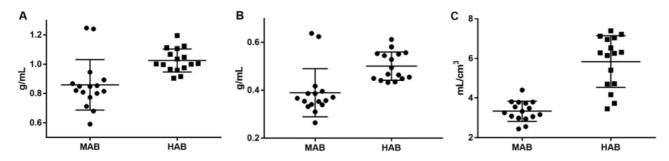
The mSUV of parent fraction was significantly different between MAB and HAB both before (p=0.0006) and after (p=0.0002) parent fraction adjustment.

Parent fraction mSUV was 0.37±0.07, mean ±SE in MAB and 0.22±0.05, mean ±SE in HAB, with a MAB:HAB ratio of 1.65.

The parent fraction time curves between HABs and MABs were also different. After adjustment of parent fraction, the mean starting point for MABs was 94% intact <sup>123</sup>I-CLINDE and 77% for HABs. Mean intact <sup>123</sup>I-CLINDE for MABs versus HABs at 20 minutes post injection was 53%(MAB) and 36% (HAB). At 90 minutes they were 38%(MAB) versus 27%(HAB).

mSUV: When applying a GLS with different residual variances on all scans and 7 ROIs, the mSUV of a 49 year old male HABs was 16.8% higher than a MAB, and for mSUV $_{60-90min}$  a HAB was 27.6% higher than an MAB. The binder-status difference between MAB and HAB was borderline for mSUV (p=0.0535) and significant for mSUV60-90min (p=0.0105).

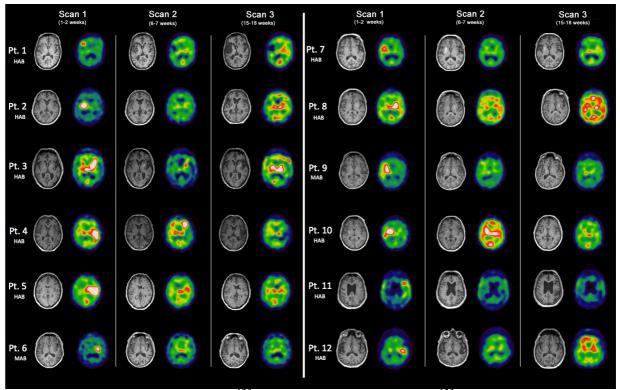
Distribution volumes of HABs were 65% higher on average than MABs when applying a GLS model on  $V_T$ s of 7 ROIs from scans with both immediately and population-based adjusted, delayed centrifuged blood samples (32 scans). The average  $V_T$  of a 49 year old male HAB was  $7.5\pm1.4$  mL/cm<sup>3</sup> (mean  $\pm$ SE), and  $4.6\pm1.4$  mL/cm<sup>3</sup> (mean  $\pm$ SE) for a MAB (Figure 5.3).



**Figure 5.3:** The effect of genotype in mSUV, mSUV<sub>60-90</sub> and  $V_T$ 's

## Results - study 4

This study generally showed that the TSPO expression after MCA stroke is remarkably dynamical. Lesional and perilesional size and intensity of the CLINDE uptake was very different for the patients at scan 1 and the expression varied substantially in the following scans (Figure 5.4).



**Figure 5.4:** T1-Weighted MRI and <sup>123</sup>I-CLINDE SPECT in stroke. <sup>123</sup>I-CLINDE SPECT is normalised by weight and injected <sup>123</sup>I-CLINDE dose for each patient

#### TSPO expression in lesion-related ROIs

For the ten HAB patients, when performing repeated measures ANOVA,  $V_T$ 's decreased lesionally over the three scans (p=0.004) with a relative decrease between scans 1 and 3 of 34.7%. A trend towards a perilesional decrease (p=0.07) was found (Figure 5.5). Performing a one-sided two-tailed t-test PD of  $V_T$  on all 12 patients lesionally and perilesionally revealed a significant mean decrease of 22.6 % lesionally (p=0.04) and 21.3 % perilesionally (p=0.001). See table 5.3 for descriptive data.

#### Correlations between TSPO expression, clinical outcome measures and necrosis

We found no significant correlation between the PD of rehabilitation measures between scan 1 and the 25-26 week follow-up and any of the ROIs. (Figure 5.5). Furthermore, we also did not find a correlation between the  $V_T$  of the contralesional thalamus measured at 5-6 weeks and the PD of rehabilitation measures between 5-6 weeks and 25-26 weeks. However, the percentage overlap between lesional-SPECT and the T2-weighted structural lesion at scan 1 correlated negatively with volume of lesion necrosis at scan 3 (p=0.001).

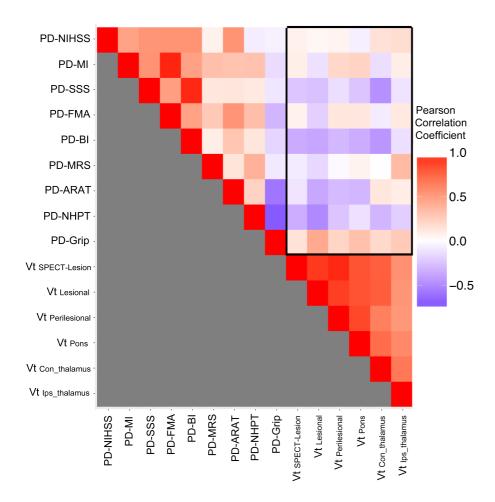


Figure 5.5: Plot of  $V_T$ 's at 1-2 weeks after stroke correlated to the PD of rehabilitation measures tested in the study using Pearson correlation. Abbreviations: Percentage difference (PD), National Institutes of Health Stroke Scale (NIHSS), Scandinavian Stroke Scale (SSS), Fugl-Meyer Assessment of the Upper Extremity (FMA-UE), Motricity Index (MI), Action Research Arm Test (ARAT), Nine-Hole Peg Test (NHPT), Grip strength (Grip), Modified Ranking Scale (MRS) and Barthel Index (BI).

#### TSPO expression in ROIs unrelated to the lesion

For the HAB patients, mean  $V_T$  in the ipsilesional cerebellum and the perilesional occipital cortex increased by 72.3% (p=0.01) and 51.4% (p=0.002). Comparing HABs of healthy volunteers with stroke patients (ten in each group) by Mann-Whitney U test, revealed a significantly lower binding of  $^{123}$ I-CLINDE to TSPO for stroke patients at scan 1 (p=0.001 and p=0.002). No significant differences at scan 2 (p=0.11 and p=0.52) and 3 (p=0.45 and p=0.31) were found. See figure 5.6 for examples of low initial TSPO expression compared to healthy volunteers, figure 5.7 for graphical representation of data, and table 5.3 for descriptive data.

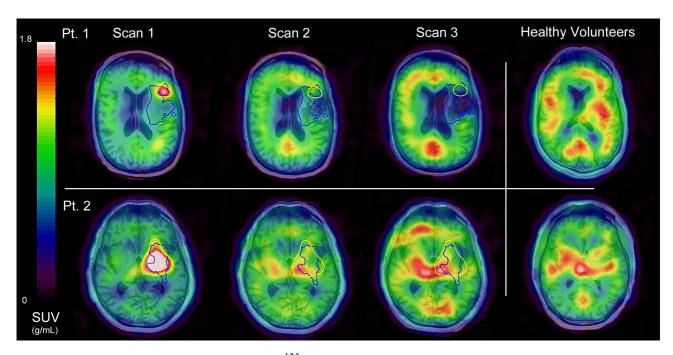


Figure 5.6 T1-weighted MRI overlaid by <sup>123</sup>I-CLINDE normalised by bodyweight and injected <sup>123</sup>I-CLINDE dose. Yellow ROIs depict the lesional SPECT upregulation and blue ROIs depict the structural lesion delineated on T2-weighted MRI (not shown) on scan 1. The initially low TSPO expression in regions not related to the lesion is apparent when compared to two healthy HAB controls. Pt.1 developed a large area of lesion necrosis and had a small initial TSPO expression ROI whereas Pt.2 had a large area of TSPO expression initially, and only a minor part of the lesion necrotized.

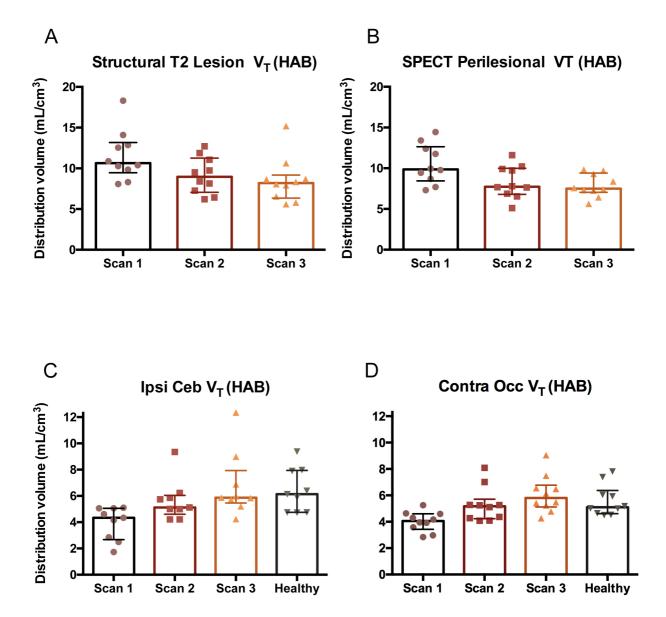


Figure 5.7:  $V_T$ 's in ten HAB patients (A-D) and ten HAB healthy volunteers (C-D). Columns represent median values and brackets span the interquartile range. Lesional  $V_T$ 's decreased (A) as well as a trend towards a decrease in the perilesional SPECT ROI (B). The ipsilesional cerebellum (C) and contralesional occipital cortex (D) ROIs were initially lower than similar regions in healthy volunteers at scan 1 and increased towards normal values at scan 3.

| Descriptive data                                     | Unit               | Median | Quartiles<br>[1st;3rd] | Range       |
|--|--------------------|--------|------------------------|-------------|
| Patient age  | Years              | 60.7   | [55.9;66.7]            | [41.7;71.7] |
| Healthy volunteer age                                | Years              | 33.6   | [24.3;45.5]            | [21.4;49.0] |
| NIHSS at stroke onset                                | Points             | 17.5   | [13.3;19.8]            | [8;22]      |
| NIHSS at scan 1                                      | Points             | 6.5    | [4.3;10.0]             | [1;12]      |
| NIHSS at follow-up                                   | Points             | 1.5    | [1.0;4.5]              | [0;5]       |
| T2 lesion volume at scan 1                           | mL                 | 40.5   | [25.1;63.8]            | [17.5;76.1] |
| T1 necrosis volume at scan 3                         | mL                 | 4.7    | [3.1;15.9]             | [2.0;50.8]  |
| Overlap: SPECT Lesion/ T2 Lesion                     | %                  | 68.8   | [51.6;80.0]            | [6.4;90.4]  |
| V <sub>T</sub> lesional scan 1 (HAB)                 | mL/cm <sup>3</sup> | 10.2   | [8.6;12.6]             | [7.5;15.0]  |
| V <sub>T</sub> lesional scan 2 (HAB)                 | mL/cm <sup>3</sup> | 8.0    | [6.3;10.5]             | [5.4;11.0]  |
| V <sub>T</sub> Lesional scan 3 (HAB)                 | mL/cm <sup>3</sup> | 7.1    | [6.2;7.8]              | [4.6;8.5]   |
| V <sub>T</sub> Con Occ cortex scan 1 (HAB)           | mL/cm <sup>3</sup> | 4.1    | [3.4;4.6]              | [2.8;5.3]   |
| V <sub>T</sub> Con Occ cortex scan 2 (HAB)           | mL/cm <sup>3</sup> | 5.2    | [4.2;5.7]              | [4.1;8.1]   |
| V <sub>T</sub> Con Occ cortex scan 3 (HAB)           | mL/cm <sup>3</sup> | 5.8    | [5.1;6.8]              | [4.3;9.1]   |
| V <sub>T</sub> Occ cortex (healthy volunteers) (HAB) | mL/cm <sup>3</sup> | 5.1    | [4.6;6.4]              | [4.5;7.8]   |

Table 5.3 Descriptive data for the patients

# 6. Discussion and perspectives

## Study 1

To our knowledge, this is the first reported human TSPO imaging study of anti-NMDAr encephalitis. The binding of <sup>123</sup>I-CLINDE to TSPO expression was initially increased. The patient responded well to treatment, and demonstrated almost normal values at the follow-up scan after 7 weeks. This implies that TSPO imaging in anti-NMDAr encephalitis has the potential to monitor treatment response in anti-NMDAr encephalitis. This could be a valuable tool since it only about 50% of anti-NMDAr encephalitis patients responds to initial treatment (Titulaer et al., 2013). If treatment non-responders could be identified and earlier given earlier secondary treatment, potentially resulting in a better prognosis.

The current study furthermore proves the feasibility of performing <sup>123</sup>I-CLINDE SPECT in anti-NMDAr encephalitis patients. The study is limited to a single patient, and further investigation into the diagnostic and monitoring role of TSPO imaging in AIE in general is warranted.

# Study 2

This is the first study in GBM patients to address the clinical value of TSPO imaging with <sup>123</sup>I-CLINDE SPECT in combination with <sup>18</sup>F-FET PET and contrast-enhanced structural imaging. The study by Pauleit (2005) demonstrated the added diagnostic value of <sup>18</sup>F-FET PET to gd-MRI and this has subsequently been standard in therapy planning and monitoring of treatment response for glioma patients in many countries including Denmark. An aim for the present study was therefore to investigate if <sup>123</sup>I-CLINDE SPECT provides new and possible valuable information when compared to <sup>18</sup>F-FET PET and gd-MRI. Due to the limited number of subjects (i.e. three) in this study, testing the prognostic value of TSPO imaging was not reasonable. Comparing <sup>123</sup>I-CLINDE and <sup>18</sup>F-FET ROIs, we found only limited overlap, indicating that TSPO binding and amino-acid uptake reflect different aspect of gliomas. We hypothesize that <sup>18</sup>F-FET positive, but <sup>123</sup>I-CLINDE negative regions primarily reflect reactive astrocytosis rather than glioma tumor cells since <sup>18</sup>F-FET is known not to be entirely glioma-specific (Floeth et al., 2006; Pichler et al., 2010; Salber et al., 2007; Spaeth et al., 2004). A marker of GBM proliferation and spatial progression would be a very valuable clinical tool in the planning of

debulking surgery and radiotherapy. This further makes TSPO a possible therapeutical target. Further research into TSPO as a marker of tumor proliferation and progression is warranted, preferably correlated with analysis of tumor biopsies.

A limitation of the study is that the patients in this study consisted of one LAB and two MABs which does not make a direct comparison of CLINDE binding to TSPO between patients possible with  $V_T$ 's as an outcome measure (Owen et al., 2015).

To compare the different scan modalities, the method of quantification was carried out as a ROI analysis. <sup>123</sup>I-CLINDE ROIs were standardized as tumor-related regions at a value above 1.5 times the mean cerebellar value. This "cerebellar threshold method" for quantification has limitations when comparing results across genotypes.

#### Study 3

# The test-retest variance of V<sub>T</sub> in <sup>123</sup>I-CLINDE SPECT

The  $V_T$  is the primary outcome measure for the binding of  $^{123}$ I-CLINDE to TSPO. The ICC values for  $^{123}$ I-CLINDE SPECT were higher than previously reported for  $^{11}$ C-PK11195 PET (Jučaite et al., 2012) and comparable to second generation TSPO tracers  $^{11}$ C-DPA-713 PET (Coughlin et al., 2014) and  $^{11}$ C-PBR28 PET (Collste et al., 2015). The compared ICC values are based upon pooled groups of HABs and MABs. This artificially increases the estimated ICC values as described in the methods section for study 4. ICC values are considerably lower when estimated separately for HABs and MABs. We were not able to compare these more factual ICC values to other second-generation TSPO tracers, as they have not been reported. The first-generation TSPO tracer  $^{11}$ C-PK11195 is not susceptible to the rs6971 genotype, which could explain part of the lower ICC values for this tracer.

In our study, ICC and COV were slightly higher for HABs than for MABs. This may be because of better signal-to-background conditions in HABs. The PD and absolute PD of  $V_{TS}$  for  $^{123}I_{TS}$  CLINDE SPECT were slightly smaller than previously reported for  $^{11}C_{TS}$  PBR28 PET (Collste et al., 2015).

#### Estimating brain uptake with mSUV and mSUV<sub>60-90</sub>

Investigating the test-retest variance and genotype difference of brain uptake by estimating mSUV and mSUV $_{60-90}$  was also done in this study. When estimating mSUV arterial blood sampling is avoided which would reduce the complexity of the scanning protocol. When comparing the mean difference in HABs and MABs, it became apparent that the  $V_T$  estimates

were more sensitive to genotype effect than mSUV and mSUV $_{60-90}$ . Thus, in situations where a change in binding of  $^{123}$ I-CLINDE to TSPO is pursued, the  $V_T$  measure is the superior measure. Other ways to avoid blood sampling is by population-based input functions, this may be a subject for a future study.

#### Effect of blood centrifugation

This study shows that PoB is decreasing if blood samples are not centrifuged immediately. Our hypothesis for this phenomenon is that parent <sup>123</sup>I-CLINDE continuously binds to TSPO in blood cells after the blood sample has been drawn, as TSPO is known to be expressed abundantly in granulocytes and monocytes, and to a lesser degree in the remaining blood cells (Canat et al., 1993). Tracer metabolites are not assumed to have this time dependent effect, as they are usually confined to the plasma phase because of lower lipophilicity and thus inability to cross cell membranes.

A lower measured plasma radioactivity because of delayed centrifugation will lead to an underestimation of the plasma input function. When performing kinetic analysis with 2TCM using a metabolite corrected arterial input function, a false low input function will result in an overestimation of  $V_T$ . In a test-retest setting an uncontrolled time to centrifugation will greatly affect the test-retest variability measures providing higher PD and lower ICC. In our protocol, the fastest possible time to centrifugation of the blood samples was within 5 minutes. It is suspected that these 5 minutes will still have an effect on the estimate of plasma and HPLC measures, especially in the first minutes after tracer injection. An important concern is therefore, firstly to be as fast as possible with the centrifugation of blood-samples and secondly, to be as consistent as possible in the time to centrifugation.

This finding will probably affect all high-affinity TSPO tracers, but it is unknown to what amount and could be a subject for further investigation.

#### Study 4

#### TSPO expression and stroke recovery

The hypothesis of the current study with regard to TSPO being a biomarker of recovery after stroke was falsified. The main reason for this negative finding may be that the TSPO signal is a combination of different reactive cell types (microglia, macrophages and astrocytes) with both pro- and anti-inflammatory properties.

On a group level, the expression of TSPO was initially high lesionally and perilesionally at 1-2 weeks and then gradually decreased perilesionally at 5-6 and 15-18 weeks. This is corresponds to other studies examining the TSPO expression between 1-4 weeks after stroke (Boutin & Pinborg, 2015).

However, on an individual level the longitudinal TSPO expression was found to be exceedingly dynamic and heterogeneous. It is a known fact that microglia cells react immediately upon damage to the brain tissue (Davalos et al., 2005). However, TSPO upregulation is first seen after 72 hours by in vivo imaging in human stroke with <sup>11</sup>C-PK11195 (Price, 2006). This time discrepancy between microglial activation and TSPO upregulation infers that there is no direct link between microglial function and TSPO expression in stroke. Another well-known limitation is that rehabilitation measures often show floor and ceiling effects which can impede the estimation of recovery especially in mild to moderate stroke (Jensen et al., 2015; Lindemann et al., 2012).

#### TSPO expression and lesion necrosis

A negative correlation was made between the initial percentage of T2-weighted lesion delineated overlapped by the <sup>123</sup>I-CLINDE ROI and the volume of lesion necrosis. This result infers that less TSPO in the lesion results in necrosis. Low TSPO expression in the core of the infarct further implies that the area is too ischemic for the TSPO expressing cells to survive and / or that the lack of perfusion hinders the arrival of tracer to target. We did not perform blood perfusion imaging (e.g. perfusion weighted MRI) to quantify the perfusion of the lesioned area. This could be of interest to investigate further, especially the overlap between perilesional TSPO expression and the ischemic penumbra. Since no lesional blood perfusion estimate was made one cannot be sure about the perfusion status of the lesion, however large necrosis of a lesion suggests permanent ischemia. Thus the results from this study seem to be in agreement with clinical studies in stroke with permanent ischemia. In these studies TSPO is primarily expressed in the

infarct margin and perilesionally initially after stroke, while the infarct core show low TSPO expression (Boutin & Pinborg, 2015).

#### TSPO expression in stroke compared to healthy volunteers

Ipsilesional cerebellum and contralateral occipital cortex had significantly lower binding of <sup>123</sup>I-CLINDE to TSPO when compared to cerebellar and occipital TSPO expression in healthy volunteers. This finding suggests a global TSPO decrease in regions not related to the lesion. Several biological processes occurring after stroke may cause this initial down regulation:

- The brain tissue could be globally affected by anti-inflammatory cytokines (e.g. IL-10) released by the high TSPO expressing reactive glial cells in the region of the lesion (Doll, Barr, & Simpkins, 2014).
- It is well known, but poorly understood that a systemic immunomodulatory process occurs after stroke (Santos et al., 2016). This could potentially affect the TSPO everywhere in the body.
- The low TSPO can be the result of low availability of TSPO expressing glia and
  perivascular macrophages, because they have migrated to the site of the lesion. However,
  TSPO is generally understood to be expressed at low concentration in healthy brain
  (Chen & Guilarte, 2008).

Previously, low TSPO expression in platelets was linked to anxiety and major depression with anxiety disorder which is hypothesised to be because of the involvement of TSPO in the production of neusteroids (Rupprecht et al., 2010). Nearly 30 % of stroke patients develop post-stroke depression (Paolucci, 2008). It could be of interest to further study this relationship.

#### Future analyses for the current study

Several future aims for the data analysis of the current study exist: Firstly, to further investigate the hypotheses of the study using more advanced statistics (e.g. latent variable models). This could provide more information on the initial TSPO expression and recovery after stroke. Secondly, additional fMRI data using a hand activation paradigm have been collected for the stroke patients and await analysis. Other unexplored data is blood samples for peripheral cytokine analysis, which was collected and stored but have yet to be analysed.

# 7. Conclusions

#### Conclusions on the studies

#### Study 1

Study 1 shows increased binding of <sup>123</sup>I-CLINDE to TSPO in anti-NMDAr encephalitis at treatment start and a decrease after 7 weeks of immunotherapy.

# Study 2

Several conclusions were drawn from study 2:

- TSPO imaging is a sensitive and specific marker of GBM.
- TSPO imaging in glioma may be a predictor of tumor cell proliferation and tumor progression.
- 123 I-CLINDE SPECT may be less susceptible to changes in BBB permeability than 18 F-FET PET.

#### Study 3

The conclusions drawn from study 3 were the following:

- The test-retest variability of <sup>123</sup>I-CLINDE SPECT is favourable compared to the first generation TSPO PET tracer <sup>11</sup>C-PK11195 and comparable to the second generation PET tracers <sup>11</sup>C-DPA713 and <sup>11</sup>C-PBR28.
- Brain SUV estimates compared to  $V_T$  estimated from 2TCM have limitations compared to  $V_T$ .
- Immediate centrifugation of blood samples is essential for correct estimation of tracer activity.

# Study 4

The following conclusions could be drawn from the final study:

- TSPO expression after stroke is remarkably heterogeneous, both initially after stroke and longitudinally.
- No correlation between initial TSPO expression and patient recovery between 2 weeks and 6 months could be made. This may be because of the heterogeneous TSPO expression in reactive glial cells with both pro- and anti-inflammatory properties.
- A negative correlation between lesional overlap percentage of <sup>123</sup>I-CLINDE with the volume of tissue necrosis was made. This corresponds to findings from animal studies and may be the result of an ischemic environment where TSPO expressing cells cannot survive and that the tracer is unavailable because the tissue is not perfused.
- Finally, when comparing regions un-related to the stroke to similar regions in healthy volunteers, the binding of <sup>123</sup>I-CLINDE to TSPO is significantly lower in stroke patients. This may be because of anti-inflammatory cytokine release from lesion-related reactive glia or a proxy to post stroke immunomodulation.

## Conclusions on the thesis objective

The main objective of the current thesis was to further advance <sup>123</sup>I-CLINDE SPECT imaging for clinical research. The following conclusions can be drawn:

- It is feasible to investigate TSPO expression research in anti-NMDAr encephalitis and glioma using <sup>123</sup>I-CLINDE SPECT and both studies points towards larger clinical studies of tumor progression in GBM and treatment response in in anti-NMDAr encephalitis.
- <sup>123</sup>I-CLINDE SPECT is a tracer with comparable properties in test-retest variance to other second-generation TSPO PET tracers. This makes it a usable tracer for research on the same terms as other second-generation TSPO tracers.
- It is feasible to conduct larger research projects in patients with the current <sup>123</sup>I-CLINDE SPECT setup and protocol (study 4).
- The clinical study (study 4) implies that the expression of TSPO is exceedingly heterogeneous after brain damage and may represent a multitude of both anti- and proinflammatory properties of the TSPO expressing cells. This is an issue when conducting studies in functional effects of TSPO dynamics.

# 8. References

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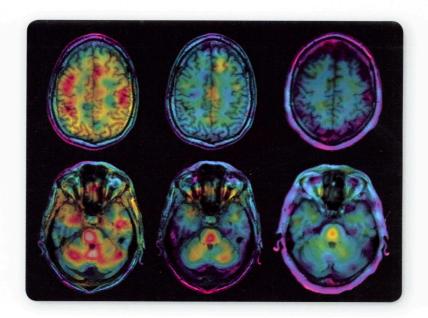
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# **Appendix**

# Study 1





Thalamus structure and function determine severity of cognitive impairment in multiple sclerosis, **p 776** 

Placebo effect of medication cost in Parkinson disease: A randomized double-blind study, **p 794**  Subclinical cardiac dysfunction increases the risk of stroke and dementia: The Rotterdam Study, **p 833** 

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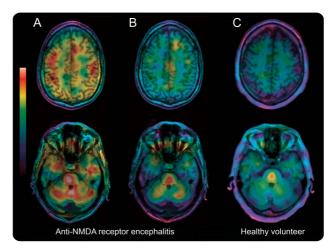
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NEUROIMAGES

# Anti-NMDAR encephalitis

Demonstration of neuroinflammation and the effect of immunotherapy

Figure [123|]CLINDE-SPECT and coregistered MRI in a patient with anti-NMDA receptor encephalitis



At the start of immunotherapy, (A) [ $^{123}$ I]CLINDE-binding to TSPO was strongly increased (neocortical distribution volume [ $V_T$ ] = 6.3 mL/cm³), but almost normalized ( $V_T$  = 4.0 mL/cm³) after 7 weeks of immunotherapy (B) compared to a healthy volunteer (C) ( $V_T$  = 3.0 mL/cm³).

A 35-year-old man presented with perceptual difficulties and delusions. At presentation, there were orofacial dyskinesias, catatonia, and autonomic instability. Anti–NMDA receptor (NMDAR) antibodies were detected in CSF.¹ Cerebral MRI was unremarkable. At the start of immunotherapy (methylprednisolone and plasmapheresis), [¹²³l]CLINDE-SPECT demonstrated a strongly increased binding to TSPO in cortical and subcortical brain regions similar to the distribution of NMDAR in the brain and different from FDG-PET changes reported in the literature (figure, A). TSPO is present on activated microglia and used as a measure of regional neuroinflammation.² After 7 weeks of immunotherapy (figure, B), TSPO binding was close to normal values (figure, C) and the patient was back to work part time as a computer scientist despite mild cognitive problems.

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Author contributions: Per Jensen: acquired and analyzed the data. Daniel Kondziella, Gerda Thomsen, and Agnete Dyssegaard: acquired data and revised the manuscript. Claus Svarer: prepared the figure, analyzed data, and revised the manuscript. Lars H. Pinborg: designed the concept, analyzed data, and wrote the clinical summary.

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# TSPO Imaging in Glioblastoma Multiforme: A Direct Comparison Between <sup>123</sup>I-CLINDE SPECT, <sup>18</sup>F-FET PET, and Gadolinium-Enhanced MR Imaging

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Here we compare translocator protein (TSPO) imaging using 6chloro-2-(4'-123|-iodophenyl)-3-(N,N-diethyl)-imidazo[1,2-a]pyridine-3-acetamide SPECT (123|-CLINDE) and amino acid transport imaging using O-(2-18F-fluoroethyl)-L-tyrosine PET (18F-FET) and investigate whether <sup>123</sup>I-CLINDE is superior to <sup>18</sup>F-FET in predicting progression of glioblastoma multiforme (GBM) at follow-up. Methods: Three patients with World Health Organization grade IV GBM were scanned with 123I-CLINDE SPECT, 18F-FET PET, and gadoliniumenhanced MR imaging. Molecular imaging data were compared with follow-up gadolinium-enhanced MR images or contrast-enhanced CT scans. **Results:** The percentage overlap between volumes of interest (VOIs) of increased <sup>18</sup>F-FET uptake and <sup>123</sup>I-CLINDE binding was variable (12%-42%). The percentage overlap of MR imaging baseline VOIs was greater for <sup>18</sup>F-FET (79%-93%) than <sup>123</sup>I-CLINDE (15%-30%). In contrast, VOIs of increased contrast enhancement at followup compared with baseline overlapped to a greater extent with baseline <sup>123</sup>I-CLINDE VOIs than <sup>18</sup>F-FET VOIs (21% vs. 8% and 72% vs. 55%). Conclusion: Our preliminary results suggest that TSPO brain imaging in GBM may be a useful tool for predicting tumor progression at followup and may be less susceptible to changes in blood-brain barrier permeability than <sup>18</sup>F-FET. Larger studies are warranted to test the clinical potential of TSPO imaging in GBM, including presurgical planning and radiotherapy.

Key Words: neurooncology; SPECT; PET; MRI

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J lioblastoma multiforme (GBM) remains the most common and aggressive primary tumor of the central nervous system. With conventional radiotherapy, chemotherapy, and debulking surgery, mean survival from diagnosis is 14.6 mo (1).

GBM lesions are strongly enhanced on contrast-enhanced structural imaging (CT or MR imaging), but populations of glioma

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cells are also present in the peritumoral environment. Thus, in clinical trials the volume for radiotherapy has been defined as the region of enhancement plus an isotropic margin of 2-3 cm (1). The combination of MR imaging and O-(2-18F-fluoroethyl)-Ltyrosine PET (18F-FET) improves the sensitivity and specificity of tumor tissue detection (2,3). However, <sup>18</sup>F-FET is not entirely glioma-specific, and increased uptake has been documented in astrogliosis secondary to infection, ischemia, radiation injury, demyelination, and hematoma (4-7). The 18-kDa translocator protein (TSPO) is a component of the mitochondrial permeability transition pore and is strongly expressed by glioma cell lines (8). In tissue resected from patients with astrocytoma, TSPO density correlates positively with malignancy and cell proliferation index and negatively with survival (9). 6-chloro-2-(4'-123I-iodophenyl)-3-(N,N-diethyl)-imidazo[1,2-a]pyridine-3-acetamide SPECT (123I-CLINDE; MAP Medical Technologies) has been validated as a second-generation TSPO tracer for use in humans (10) and recently in a GL26 mouse model of glioma (11). We present <sup>123</sup>I-CLINDE SPECT, <sup>18</sup>F-FET PET, and gadolinium-enhanced MR imaging results from 3 GBM patients at baseline compared with contrast-enhanced structural imaging at follow-up and hypothesize that 123I-CLINDE SPECT at baseline is an imaging biomarker of GBM progression.

#### **MATERIALS AND METHODS**

The study was performed in accordance with the ethical standards of the Declaration of Helsinki and approved by the ethical committee of the Copenhagen Capital Region (approval H-2-2010-086, amendment 39319). All subjects signed an informed consent form. Three patients in an advanced state of GBM (World Health Organization grade IV) were included and genotyped for the rs6971 polymorphism to determine the TSPO binder status as described previously (10). Before inclusion, all patients had undergone surgery and received radiotherapy and chemotherapy with temozolomide, bevacizumab and irinotecan. In addition, patients 2 and 3 had been treated with a cell-based immunotherapy. Patient 2 had undergone surgical resection between baseline and follow-up MR imaging. The patients were scanned with  $^{123}\mbox{I-CLINDE},~^{18}\mbox{F-FET},$  and MR imaging within 2 d. Patients 1 and 2 were rescanned with MR imaging after 6 and 17 wk, respectively, and patient 3 was rescanned with CT after 4 wk. The patients received no treatment with angiogenesis-inhibiting drugs for 6 wk before the scans and no radiochemotherapy between scans.

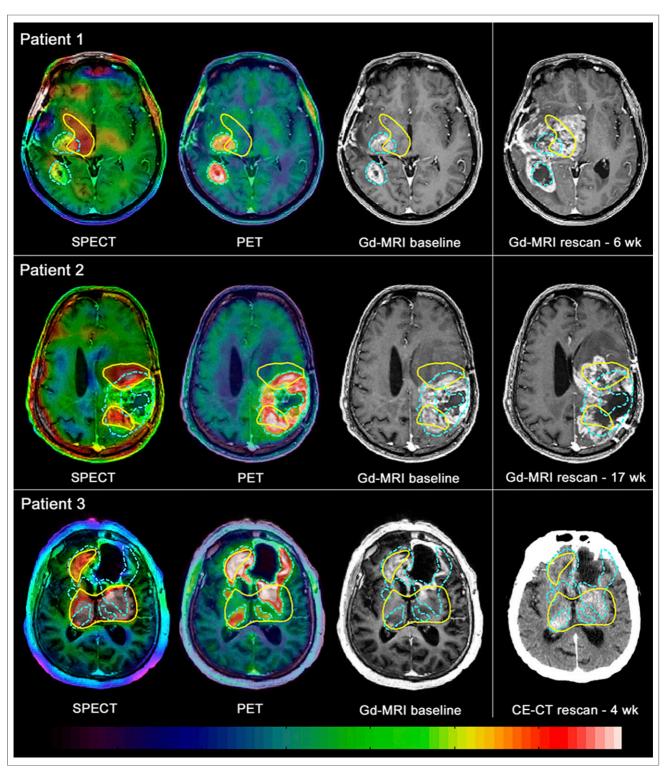
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**FIGURE 1.** Weighted mean 0- to 150-min <sup>123</sup>I-CLINDE SPECT scans, summed 20- to 40-min <sup>18</sup>F-FET PET scans, and gadolinium- or contrast-enhanced (CE) structural scans at baseline and at rescanning. Blue VOIs show high <sup>18</sup>F-FET tumor uptake, and yellow VOIs high <sup>123</sup>I-CLINDE binding. Comparison of baseline and rescanning structural scans reveals that tumor expands predominantly in areas of high <sup>123</sup>I-CLINDE binding.

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TABLE 1
Patients

| Patient | Sex | Age (y) | Binder status | Rescanning modality | Interval (wk) | <sup>123</sup> I-CLINDE injected dose (MBq) |
|---------|-----|---------|---------------|---------------------|---------------|---|
| 1       | F   | 48      | MAB           | MR imaging          | 6             | 191   |
| 2       | М   | 65      | LAB           | MR imaging          | 17            | 188   |
| 3       | М   | 64      | MAB           | СТ                  | 4             | 188   |

TABLE 2 VOIs

| Patient | <sup>18</sup> F-FET | <sup>123</sup> I-CLINDE | <sup>18</sup> F-FET ∩ <sup>123</sup> I-CLINDE | MR imaging (baseline) | MR imaging/CT (follow up - baseline) |
|---------|---------------------|-------------------------|---|-----------------------|--------------------------------------|
| 1       | 17.3                | 36.6                    | 2 (12%)                                       | 7.2                   | 97.5                                 |
| 2       | 196.9               | 97.9                    | 62.6 (32%)                                    | 150.5                 | NA                                   |
| 3       | 180                 | 140.7                   | 75 (42%)                                      | 60.7                  | 44.8                                 |

NA = not applicable. Data are in milliliters.

**TABLE 3**Volumes in Common Between Baseline Structural Imaging and <sup>123</sup>I-CLINDE or <sup>18</sup>F-FET PET

|         |                          |                              | Sørensen-Dice o                    | coefficient between                    |
|---------|--------------------------|------------------------------|------------------------------------|--|
| Patient | <sup>18</sup> F-FET (mL) | <sup>123</sup> I-CLINDE (mL) | <sup>18</sup> F-FET and MR imaging | <sup>123</sup> I-CLINDE and MR imaging |
| 1       | 6.7 (93%)                | 1.1 (15%)                    | 0.54                               | 0.05                                   |
| 2       | 120.1 (80%)              | 44.4 (30%)                   | 0.69                               | 0.36                                   |
| 3       | 48.0 (79%)               | 18.3 (30%)                   | 0.4                                | 0.18                                   |

TABLE 4
Volumes in Common Between Tumor Progression at Follow-up Structural Imaging and Baseline

18F-FET PET or 123I-CLINDE

| Patient | <sup>18</sup> F-FET (mL) | <sup>123</sup> I-CLINDE (mL) |
|---------|--------------------------|------------------------------|
| 1       | 7.7 (8%)                 | 20.9 (21%)                   |
| 3       | 24.6 (55%)               | 32.3 (72%)                   |

#### Patients

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Patient data, genotype, rescan modality, scan-rescan time intervals, and injected  $^{123}$ I-CLINDE dose are presented in Table 1.

#### **Image Acquisition**

After bolus injection of <sup>123</sup>I-CLINDE, dynamic SPECT images were acquired with a triple-head IRIX camera (Philips Medical) for 2.5 h as previously described (*10*), and weighted mean images were generated for region analysis.

A single-frame static PET acquisition was performed 20–40 min after intravenous injection of approximately 200 MBq of <sup>18</sup>F-FET on a 64-slice-CT Biograph TruePoint PET/CT scanner (Siemens). All PET scans were attenuation-corrected using low-dose CT performed immediately before the PET scan and were subsequently corrected for scatter and dead time. Images were reconstructed with an ordered-subsets expectation maximization 3-dimensional algorithm (6 itera-

tions, 16 subsets) and a 5-mm gaussian filter. All patients fasted for at least 6 h before  $^{18}\text{F-FET}$  injection.

T1-weighted MR imaging was performed on a 3-T MR Verio scanner (Siemens). Gadolinium was used in a dose of 0.1 mmol/kg of body weight (Multihance [Bracco] or Gadovist [Schering]). Patient 3 was rescanned using a CT scanner (Siemens) and a CT contrast agent containing 70 mL of iodine (350 mg/mL) (Omnipaque; GE Healthcare).

#### **Data Processing**

The weighted mean <sup>123</sup>I-CLINDE SPECT image was coregistered to the MR image of the same patient using interactive image overlay (12). <sup>18</sup>F-FET PET images and the follow-up MR images were coregistered to the first MR image of the same subject, applying the automatic rigid-body model included with Statistical Parametric Mapping, version 8 (13).

The definition of volumes of interest (VOIs) with high  $^{123}$ I-CLINDE binding was standardized for all patients by automatically selecting voxels with a value above 1.5 times the mean cerebellar count. The delineation of high-uptake  $^{18}$ F-FET volumes was standardized as previously described (2). A trained neuroradiologist supervised delineation of the MR imaging volumes. Baseline MR imaging VOIs were delineated manually as the contrast-enhanced areas of the coregistered baseline MR image. Blood vessels were omitted. Volumes for SPECT, PET, and MR imaging were determined, along with common volumes. Furthermore, the Sørensen–Dice coefficient for SPECT, PET, and baseline MR imaging was calculated as  $\frac{2V_{A\cap B}}{V_A + V_B}$ , with  $\cap$  being the symbol of common volume. Volume of tumor progression was

defined as the additional contrast-enhanced volume from baseline to follow-up. For patient 2, follow-up volume analysis was not performed because of surgical resection between the baseline and follow-up scans.

#### **RESULTS**

Table 2 shows the sizes of the <sup>18</sup>F-FET and <sup>123</sup>I-CLINDE VOIs as well as the volumes in common for high-binding <sup>123</sup>I-CLINDE VOIs and high-uptake <sup>18</sup>F-FET VOIs. The VOIs of increased <sup>123</sup>I-CLINDE binding and <sup>18</sup>F-FET uptake showed varying degrees of overlap (12%–42%). Binder status does not appear to determine the volume of <sup>123</sup>I-CLINDE VOIs.

Table 3 shows the volumes in common between baseline structural imaging and <sup>123</sup>I-CLINDE or <sup>18</sup>F-FET PET. In all cases, baseline MR imaging VOIs showed greater similarity to <sup>18</sup>F-FET VOIs (Sørensen–Dice coefficients, 0.54, 0.69, and 0.40) than to <sup>123</sup>I-CLINDE VOIs (Sørensen–Dice coefficients, 0.05, 0.36, and 0.18). The percentage overlap of MR imaging baseline VOIs was greater for <sup>18</sup>F-FET VOIs (79%–93%) than for <sup>123</sup>I-CLINDE VOIs (15%–30%).

Table 4 shows the volumes in common between tumor progression at follow-up structural imaging and baseline <sup>18</sup>F-FET PET or <sup>123</sup>I-CLINDE. In the 2 patients for whom follow-up tumor progression volumes were estimated, the volume with de novo contrast enhancement at follow-up overlapped to a greater extent with <sup>123</sup>I-CLINDE VOIs than with <sup>18</sup>F-FET VOIs (21% vs. 8% and 72% vs. 55%).

#### DISCUSSION

This study follows 4 previous studies demonstrating TSPO in 1–3 patients with GBM using the TSPO PET tracer <sup>11</sup>C-PK11195 PET (*14–17*). To our knowledge, this is the first study to address the clinical value of TSPO imaging in GBM patients by comparing the second-generation TSPO SPECT tracer <sup>123</sup>I-CLINDE with <sup>18</sup>F-FET PET and contrast-enhanced structural imaging.

The first aim of the present study was to evaluate the similarity of <sup>123</sup>I-CLINDE and <sup>18</sup>F-FET VOIs. We found only limited VOI overlap, indicating that imaging of amino acid uptake and TSPO binding reflects different aspects of GBM pathology. Increased <sup>18</sup>F-FET uptake has been documented experimentally and clinically in reactive astrogliosis secondary to infection, ischemia, radiation injury, demyelination, and hematoma, indicating that <sup>18</sup>F-FET is not entirely glioma-specific (4-7). In contrast, no TSPO expression was found in reactive astrogliosis in a recent study in untreated human glioma patients (17). In addition to binding to glioma cells, TSPO tracers bind to myeloid cell lines, including glioma-associated microglia and macrophages (18). In contrast, uptake of <sup>18</sup>F-FET is not increased in areas of infiltrating activated macrophages and activated microglia in experimental abscesses (4,19). It may be hypothesized that TSPO-negative but <sup>18</sup>F-FET-positive areas represent primarily reactive astrocytosis, predicting a less aggressive development, but biopsy control confirmation is needed.

The second aim was to evaluate the correspondence between the VOIs of increased <sup>18</sup>F-FET uptake, the VOIs of increased <sup>123</sup>I-CLINDE binding, and volumes of MR imaging contrast enhancement at baseline. We found that <sup>18</sup>F-FET VOIs overlapped more (93%, 80%, 79%) with contrast-enhanced VOIs than with <sup>123</sup>I-CLINDE VOIs (15%, 30%, 30%). Contrast enhancement reflects a disturbed blood–brain barrier and could be related to treatment and tissue necrosis and not glioma cell proliferation per se (20).

The binding of  $^{123}$ I-CLINDE to TSPO does not appear to be significantly increased in areas of disrupted blood–brain barrier (10). It is known that glioma cells diffusely infiltrate brain tissue without disrupting the blood–brain barrier (21).

The third aim was to evaluate how <sup>18</sup>F-FET and <sup>123</sup>I-CLINDE VOIs overlap with VOIs representing tumor progression from baseline to follow-up on contrast-enhanced structural imaging. Follow-up VOI analysis was not performed for patient 2 because debulking surgery had taken place between baseline and follow-up scans and extravasation of gadolinium was highly increased in the area around the operation cavity on the follow-up scan. It appears that the VOIs of increased 123I-CLINDE at baseline are a good visual predictor of tumor progression at follow-up (Fig. 1). In line with the visual interpretation, the percentage overlap with VOIs representing tumor progression is higher for <sup>123</sup>I-CLINDE than for <sup>18</sup>F-FET (21% vs. 8% and 72% vs. 55%) in patients 1 and 3. The findings correspond to previous reports demonstrating the contribution of TSPO to the uncontrolled cellular proliferation of glioma cells (22) and the correlation between TSPO density in surgically removed glioma tissue with clinical parameters such as survival time and tumor cell proliferation index (8,9). TSPO is expressed in glioma cells, suggesting that this method may be a more sensitive and specific marker of ongoing tumor cell proliferation and progression than conventional imaging methods. In addition, gliomaassociated macrophages and monocytes contributing to tumor progression through the release of proinflammatory and proangiogenic factors may also add to the TSPO signal (23). Thus, the addition of TSPO imaging to 18F-FET PET has the potential to add information about areas of very active tumor cell proliferation in GBM. This may explain why low-grade gliomas have few or no TSPO binding sites (17).

#### CONCLUSION

These preliminary results from 3 patients with advanced GBM suggest that TSPO imaging is a sensitive and specific marker of GBM and that regional binding predicts areas of active tumor cell proliferation in GBM. Favorable implications can be foreseen for planning surgery and radiotherapy and monitoring the effect of oncologic therapy. <sup>123</sup>I-CLINDE SPECT appears to be less susceptible to blood–brain barrier disruption than <sup>18</sup>F-FET PET. However, to determine the exact role of TSPO imaging compared with imaging of amino acid transport, further studies are warranted.

## DISCLOSURE

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# Study 3

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# The variability of translocator protein signal in brain and blood of genotyped healthy humans using in vivo 123I-CLINDE SPECT imaging – a test-retest study

Ling Feng, Per Jensen, Gerda Thomsen, Agnete Dyssegaard, Claus Svarer, Lars Vestergaard Knudsen, Kirsten Møller, Carsten Thomsen, Jens Damsgaard Mikkelsen, Denis Guilloteau, Gitte Moos Knudsen and Lars H. Pinborg

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The variability of translocator protein signal in brain and blood of genotyped healthy humans

using in vivo <sup>123</sup>I-CLINDE SPECT imaging – a test-retest study

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Running Title: Test-retest variability of <sup>123</sup>I-CLINDE

The aim of the current study was to examine the test-retest reproducibility measured as PD and reliability measured as ICC and COV of <sup>123</sup>I-CLINDE binding in healthy subjects based on the brain SUV and 2TCM with arterial blood as the input function.

#### **MATERIALS AND METHODS**

## Subjects and genotyping

This study was conducted in accordance with the Declaration of Helsinki at the Copenhagen University Hospital, Rigshospitalet, Denmark. The ethical committee of the Copenhagen Capital Region (H-2-2010-086) approved the study protocol. All subjects provided written informed consent.

Due to low TSPO signal in glioblastoma tissue from a patient who was a low affinity binder (LAB) (12), we did not expect to obtain reliable TSPO signals in healthy LABs. Consequently, after screening for the rs6971 polymorphism (12), 16 healthy subjects (MAB:HAB=1:1, 9 females) were included and SPECT scanned twice with an interval of 35±15 days. All healthy subjects had normal physical and neurological examinations, and blood tests. Table 1 shows a summary of age, gender, TSPO genotype and scan information.

## Arterial blood sampling and centrifugation

From a radial arterial cannula, blood samples for measuring radioactivity in plasma and whole blood were drawn manually at 0.25, 0.5, 0.75, 1, 1.5, 2, 2.5, 3, 4, 6, 8, 10, 12, 15, 20, 25, 30, 35, 45, 65 and 85 min post injection of <sup>123</sup>I-CLINDE. Additionally, samples were drawn at 0.5, 4, 10, 20, 30, 45, 65 and 85 min for metabolite analysis by radio-high performance liquid chromatography (radio-HPLC) (*16*). Fractions of HPLC eluent were collected using a fraction collector device (Foxy Jr FC144, Teledyne Isco) and counted off-line in a well-counter (2480 Wizard2 Gamma Counter, Perkin Elmer) for accurate measurements of parent tracer. For the

first eight subjects the blood was stored on ice for an estimated 30-90 min until centrifugation (Eppendorf centrifuge 5800R with 3500 rpm at 4° C in 7 min). During the course of the study, we discovered that <sup>123</sup>I-CLINDE continued to bind to TSPO in formed blood cells, presumably the monocytes, until separating plasma from the sample by centrifugation. For the remaining 8 subjects, all blood samples were stored on ice and centrifuged within max. 5 min. Total radioactivity in plasma and whole-blood was measured in a well-counter (Cobra 5003; Packard Instruments) and data were decay-corrected to the time of injection. Further, the radioligand *purity* was measured for each batch of <sup>123</sup>I-CLINDE, and trapping efficiency of the HPLC column (*plasma control*) was determined for each scan by spiking water and blank plasma with <sup>123</sup>I-CLINDE. All equipment was cross-calibrated to the SPECT scanner. Blood screening including white blood cell differential count was performed at each scan.

# **Image acquisition**

To block thyroidal uptake of free radioiodine, 200 mg of potassium perchlorate was administered intravenously 20 min before injection of <sup>123</sup>I-CLINDE. A headband was applied to minimize head movements during scanning. Dynamic SPECT scanning with a triple-head IRIX camera (Philips Medical) was started simultaneously with a bolus injection of <sup>123</sup>I-CLINDE (MAP Medical Technologies), and lasted for 90 mins. The protocol consisted of 10x2-minute frames, followed by 7x10-minute frames.

T1-weighted magnetic resonance imaging (MRI) was acquired once for each subject using a 3T Siemens Prisma MR scanner  $3 \pm 49$  (mean  $\pm$  standard deviation, median=0) days after the corresponding SPECT scan. All brain scans were normal as interpreted by a neuroradiologist.

## **Image analysis**

Image preprocessing. For each subject the weighted mean SPECT image was co-registered to the T1-weighted MRI by interactive image overlay (17). Automatic delineation of volumes of interest (VOIs) was performed using probability maps and a data processing pipeline (18). Due to the limited spatial resolution of SPECT images, no segmentation of grey matter, white matter and cerebrospinal fluid was attempted. Seven VOIs were investigated in this study: midbrain, thalamus, pons, striatum, hippocampus, cerebellum (without vermis) and neocortex. These VOIs were defined as volume-weighted means of either the left- and the right-hemisphere or of smaller VOIs: striatum as the mean of caudate nucleus and putamen; and neocortex as the mean of frontal cortex, superior temporal gyrus, medial inferior temporal gyrus, parietal cortex, sensory motor cortex and occipital cortex.

Brain and blood uptake. The radioactive concentrations in blood and brain were studied independently to investigate the test and retest variability. Therefore, percentage difference of the mean SUV (mSUVs) was calculated, where SUV (g/ml) was defined as TAC (kBq/ml) in blood or brain VOIs normalized by the injected dose per body weight (in MBq/kg) (19), and mSUV is defined as the area under the SUV curve normalized by time length.

Kinetic modelling. A 2TCM with arterial plasma function as input ( $K_1$  to  $k_4$  with a fixed blood volume in brain tissue of 5%) was used to quantify <sup>123</sup>I-CLINDE binding as distribution volume ( $V_T$ ), as this has previously been demonstrated to be the optimal method (12).

The arterial input function was generated in two ways, depending on the blood sample centrifugation.

Method 1: For studies with immediate centrifugation of blood samples

To account for tracer purity and trapping efficiency the measured parent fraction (PF) from the fraction collector was first fitted by a multi-exponential function and adjusted as below:

$$PF(t) = a_1 \cdot e^{-b_1 t} + a_2 \cdot e^{-b_2 t} + a_3$$
 eq. 1

$$PF \_adj(t) = (PF(t) + purity - plasma std) / purity$$

The difference between <sup>123</sup>I-CLINDE purity and plasma control indicates the part failed to be trapped by the extraction column of the radio HPLC.

Then the metabolite-corrected plasma input function was calculated as the product of the plasma TAC Cp(t) and the adjusted parent fraction  $PF\_adj(t)$ :

$$C_{mcp}(t) = C_p(t) \cdot PF \_adj(t)$$
 eq. 2

Method 2: For studies with delayed centrifugation of blood samples

Plasma to whole-blood ratio (PoB) and adjusted parent fraction as a function of time were generated for all scans that were accompanied by blood samples undergoing immediate centrifugation. The mean of these functions, respectively, were generated as population-based PoB (PB PoB) and population-based parent fraction (PB PF):

$$PoB_i(t) = a_i \cdot (1 - e^{-b_i t}) + c_i$$

$$PB PoB(t) = \sum_{i=1}^{N} PoB_i(t) N$$
 eq. 3

$$PB PF(t) = \sum_{i=1}^{N} PF adj_i(t) N$$

where N is the number of scans. To generate metabolite-corrected plasma input function ( $C_{mcp}$ ) these two functions were then applied on the measured whole-blood TAC ( $C_w$ ) of the studies with delayed centrifugation as below:

$$C_{mcp}(t) = C_w(t) \cdot PB PoB(t) \cdot PB PF(t)$$
 eq. 4

Image and data pre-processing were performed using MATLAB 8.1 (R2013a, Mathworks Inc., MA) and kinetic modelling was done using PMOD (version 3.0; PMOD Technologies Inc., Switzerland).

# Statistical analysis

To evaluate the test-retest reproducibility and variability of <sup>123</sup>I-CLINDE the following four matrices were used: PD, absolute PD, ICC and COV.

PD: test-retest reproducibility was calculated as the difference between test and retest outcome measures divided by their average, outcome measures being mSUV of blood or brain uptake, and  $V_T$ . The absolute PD was also calculated for comparison to other studies.

*ICC*: ICC evaluates the reliability by measuring within-subject variability relative to between-subject variability as below:

$$ICC = \frac{MSS_{btw} - MSS_{within}}{MSS_{btw} + (k-1)MSS_{within}}$$
 eq. 5

where 
$$MSS_{btw} = \frac{1}{N-1} \sum_{i=1}^{N} (\overline{V}_{Ti} - \overline{V}_{T})^2 MSS_{within} = \frac{1}{N} \sum_{i=1}^{N} \sum_{j=1}^{k} (V_{Tij} - \overline{V}_{Ti})^2$$
 eq. 6

 $\overline{V}_{Ti}$  is the mean of test and retest measures of subject i, and  $\overline{V}_{Ti}$  is the mean of all observations (test and retest of all subjects).  $V_{Tij}$  is one of the measures of subject i. N is the number of subjects, and k is the number of repeated measures (k=2). If  $MSS_{btw}$ =0 there is no reliability, and ICC= -1; if  $MSS_{within}$ =0 there is maximum reliability, and ICC=1.

*COV*: The COV of outcome measures was calculated as the standard deviation divided by the mean across all observations for each VOI.

To test effects of centrifugation, genotype or the test-retest, a linear model fitting using generalized least squares (GLS) was performed in R (version 3.2.2). In general, a GLS model performs similar to a standard linear model, but can furthermore account for a complex

covariance structure between observations. In our case the GLS model allows for different variances between residuals in repeated measures and for correlation between them, unless otherwise stated.

Details about each model applied will be given in the results section. For GLS models, mean and standard error (SE) are given. In other cases, mean and standard deviation are given. Paired-sample t-test, Wilcoxon rank sum test and Pearson's linear correlation were performed using MATLAB 8.1 (R2013a, Mathworks Inc.).

## **RESULTS**

The injected radioactivity in test and retest scans was not statistically different; neither was <sup>123</sup>I-CLINDE purity or plasma control, for details see (Table 1).

# Delayed centrifugation and discrepancy between blood and brain PD.

In the first eight subjects with delayed centrifugation of blood samples, we found a large test-retest PD of mSUV in the plasma parent compound (1.0±31.3%), but much lower PD in the brain (0.7±8.0% - 2.2±18.8% in 7 VOIs).

## **Effect of genotype**

In subjects with immediately centrifuged samples, the parent fraction mSUV was higher in MAB (0.37 $\pm$ 0.07, mean $\pm$ SE) than in HAB (0.22 $\pm$ 0.05, mean $\pm$ SE; p=0.0006, GLS model, age and gender adjusted), with a MAB:HAB ratio of 1.65. A genotype-related difference was also observed in PoB (Supplemental Fig. 1). The mean PoB TAC of HAB (N=2x3) and that of MAB (N=2x5) had different kinetics, especially during the first 20 min, however the difference was not significant. Mean SUV<sub>0-20min</sub> of MABs correlated negatively with monocyte counts (p=0.0341, R<sup>2</sup>=0.4486). A negative trend was also observed for HABs (p=0.1938).

# **Arterial input function**

Immediately centrifuged blood samples. After parent fraction adjustment using eq. 1, the parent fraction between MAB and HAB was still significantly different (*p*=0.0002, GLS model, age and gender adjusted). Furthermore, the parent fraction curves had different starting points: 94% intact <sup>123</sup>I-CLINDE left for MABs, and 77% for HABs; and at 20 min post injection, 53% for MABs, and 36% for HABs; and at 90 min 38% left for MABs and 27% for HABs.

The arterial input function of a MAB and a HAB (Figs. 1C and 1D) showed faster washout in the HAB. Due to the observed differences between genotypes, population-based PoB functions and population-based PF functions were generated separately for these two genetic groups. Before applying the population-based functions to adjust data with delayed centrifugation, method validation was done on the data with immediate centrifugation using a leave-one-out procedure, where population-based functions were generated excluding the scan that the functions were applied on.

These  $PB\_PoB$  functions were applied to the whole-blood TAC Cw first to estimate the plasma TAC Cp. The differences between estimated Cp and the measured data were  $0.2\pm4.3\%$  for MABs (N=2x5), and  $0.6\pm10.8\%$  for HABs (N=2x3), none were significant. Thereafter,  $PB\_PF$  was applied to Cp to estimate the metabolite-corrected plasma input function Cmcp. Differences between population-adjusted estimates and measured Cmcp were  $0.6\pm9.1\%$  for MABs and  $0.5\pm8.0\%$  for HABs, none were significant.

Delayed centrifuged blood samples. Immediate centrifugation increased the mSUV of PoB from 0.95 to 1.07 (p=0.0003, GLS model, genotype adjusted), which was 12.4% more than the mSUV of PoB from subjects whose samples underwent delayed centrifugation; see descriptive data of PoB in (Fig. 2)). Thus, the arterial input functions were underestimated in scans with

delayed centrifugation of blood samples (Figs. 1A and 1B). Therefore, population-adjusted input functions were calculated in these cases.

After adjustment using  $PB\_PoB$  and  $PB\_PF$  on Cw, test-retest PD of Cp decreased from - 22.7±27.6% to -9.4±15.2% (absolute PD of Cp decreased from 27.7% to 14.4%), and PD of Cmcp changed from -2.5±34.6% to -6.4±15.5% (absolute PD of Cmcp decreased from 28.0% to 14.1%) (Supplemental Fig. 2).

#### **Test-retest variability**

*Brain uptake.* Brain mSUV of 7 VOIs were studied. The mSUV of a male HAB and a male MAB at age 49 years (mean age) were 1.03±0.14 g/ml and 0.88±0.15 g/ml (mean±SE) respectively, and the difference was borderline significant (*p*=0.0535, GLS with different residual variances). The higher binding in HABs than in MABs was also shown in (Fig. 1) illustrated as SUVs of neocortex with slower washout in HABs. Mean SUV of HABs was 16.8% higher than MABs, see descriptive data in (Fig. 3A). Ten of 16 subjects had lower brain mSUV in test than retest scans.

Since the beginning of a brain TAC can be influenced by blood flow, mSUV<sub>60-90min</sub> was also studied. The mSUV<sub>60-90min</sub> of a male HAB and a male MAB at age 49 years were  $0.55\pm0.09$  g/ml and  $0.43\pm0.09$  g/ml (mean±SE) respectively (p=0.0105, GLS with different residual variances), and the mSUV<sub>60-90min</sub> of HABs was 27.6% higher than MABs, see descriptive data in (Fig. 3B). The PDs of brain mSUVs across all subjects in 7 VOIs are shown in (Table 2). Test and retest mSUV of the same subject correlated to a high degree, with correlation coefficients of  $0.86\pm0.07$  (p<0.05, N=16). The PD of MABs was on average -3.6±8.1% while that of HABs was -7.0±7.8%, however, the difference was not significant (GLS model, age and gender adjusted), and the absolute PD was neither sensitive to genotype. ICC values for MABs and HABs and the total

samples are given in (Table 2). COV was  $20.0\pm2.3\%$  in MABs and  $9.2\pm2.0\%$  in HABs. Furthermore, ICC and COV differed between MABs and HABs (p=2.29e-05, p=1.38e-05, respectively), with a MAB:HAB ratio of 1.8 for ICC and 2.2 for COV.

Distribution volumes. The V<sub>TS</sub> of 7 VOIs derived from 2TCM modelling of scans with immediately and delayed centrifuged blood samples, with and without population-based adjustment of the input functions, are given in (Supplemental Table 1, 2 and 3). Figure 1 illustrates the 2TCM fitting using the measured or population-based adjusted input functions. The V<sub>T</sub>s of scans with delayed centrifuged blood samples (16 scans, 7 VOIs) before and after population-based adjustment was significantly different (p=0.0176, GLS with different residual variances) with a V<sub>T</sub> value of 5.3±1.1 mL/cm<sup>3</sup> before the adjustment and 4.7±1.1 mL/cm<sup>3</sup> (mean±SE) after the adjustment. Delayed centrifugation of samples led to an underestimation of input functions and therefore caused an overestimation of the V<sub>T</sub>s by 12.4% on average. Using population-based adjusted input functions in scans with delayed blood centrifugation and measured input functions in scans with immediate blood centrifugation, the V<sub>T</sub> (32 scans, 7 VOIs) of a male HAB and a male MAB at age 49 years (mean age) was 7.5±1.4 mL/cm<sup>3</sup> and 4.6±1.4 mL/cm<sup>3</sup> (mean±SE) respectively (p=0.0001, GLS with different residual variances). V<sub>T</sub> of HABs was on average 65% higher than MABs, and there was no test-retest difference, see descriptive data in (Fig. 3C). Test-retest variability measures were calculated for the immediately centrifuged scans and the population-adjusted delayed centrifuged scans both separately and combined (Table 2). For subject 16 the measured purity and plasma standard were not available, thus no adjustment was done.

For the immediately centrifuged scans, test  $V_T$  was 0.28 mL/cm<sup>3</sup> lower than retest  $V_T$  across all 7 VOIs (p=0.0122, GLS model, age and gender adjusted). This GLS model assumes a

homogeneous test-retest difference across VOIs, since there were no interactions between the 7 VOIs and test-retest scans.

The PD between test-retest of all subjects (N=16) across all 7 VOIs evaluated as repeated measures was on average -11.7±8.6% for MABs and -12.1±8.3% for HABs, and the difference was not significant (GLS, age and gender adjusted). COV of MABs was 16.7±3.4%, and of HABs was 26.5±7.7%. Opposite to the brain mSUV, ICC and COV estimated by the V<sub>TS</sub> were borderline significantly higher for HABs than MABs (p=0.0533, p=0.0492, respectively), with a HAB:MAB ratio of 1.6 for ICC and 1.7 for COV.

#### **DISCUSSION**

In this study we investigated the test-retest variability of the second-generation TSPO SPECT ligand, <sup>123</sup>I-CLINDE, in a group of 16 healthy volunteers.

## Population-based approach

We used a population-based approach to adjust data from subjects with delayed blood centrifugation. The feasibility of the approach was validated on the immediately centrifuged data in a leave-one-out manner to avoid bias. After adjustment, the test-retest difference decreased in blood components. If time-dependent binding to blood cells is also observed in other TSPO ligands, this approach could be applied to data if centrifugation is delayed. Furthermore, this approach has the potential to reduce the complexity and time consumption associated with blood handling and determination of metabolites, since only whole blood counts are needed.

## <sup>123</sup>I-CLINDE SPECT variability

ICC values of V<sub>T</sub>s were higher for <sup>123</sup>I-CLINDE SPECT than previously reported in healthy controls for the first-generation TSPO tracer <sup>11</sup>C-PK11195 PET (7) and comparable to second-generation TSPO tracers like <sup>11</sup>C-PBR28 (9) and <sup>11</sup>C-DPA-713 (8). ICC is a statistical analysis to

compare the between-subject variation with the within-subject variation. If HAB and MAB subjects are pooled and not measured separately for each genetic group, ICC values will be higher and not only reflect the variability of the method but also a known biological difference in affinity. In contrast to previous studies in second-generation TSPO tracers we also calculated ICC and COV separately for each genetic group, revealing higher ICC and COV values for HABs than MABs. A plausible explanation for a generally modest reliability when subdividing into genotypes is that the TSPO signal, due to its non-prominent presence under healthy conditions, has a relatively low signal-to-background ratio compared to a disease situation with an amplified TSPO signal driven by disease-related activated glial cells. For MABs the signal-to-background ratio is lower than for HABs, consequently leading to a comparably lower reliability. Percentage difference of brain mSUV and V<sub>T</sub> and the absolute PD for 123I-CLINDE SPECT were slightly smaller than previously reported for <sup>11</sup>C-PBR28 (9).

#### SUV as a semi-quantitative measure

In a clinical setting, SUV is often used as a semi-quantitative surrogate measure to V<sub>T</sub>. This is due to several complications related to the complexity of the blood sampling and analysis. In this paper we studied the genotype difference and test-retest variability using both SUV and V<sub>T</sub>, to investigate the feasibility of using SUV as an outcome measure for a TSPO ligand. From the brain mSUV it is possible to differentiate MAB and HAB, (Fig. 4), with HABs having ~17% higher binding than MABs when quantified using mSUV of the whole TAC, and ~28% higher when quantified using mSUV<sub>60-90min</sub>. However, HABs had on average 65% higher binding than MABs using V<sub>T</sub> estimated by 2TCM modelling. This can be explained by higher affinity of radioligand to TSPO in HABs and the binding of radiotracer to the abundant TSPO in both blood

and peripheral tissues. Thus, less free radioligand would be available in plasma of HABs compared to MABs which would remain undetected using brain SUV as an outcome. In a clinical setting, patients can have a longitudinally varying disease related peripheral immune response and thereby varying peripheral TSPO binding sites. This may cause changing concentrations of parent TSPO tracer in the plasma, which as a consequence influences the brain uptake. In (20) SUV was compared to V<sub>T</sub> in quantifying TSPO changes in baboons' brain caused by lipopolysaccharide using <sup>11</sup>C-PBR28, and SUV failed to detect neuroinflammation as indicated by V<sub>T</sub> changes. It might be ascribed to the violation of assumptions: 1) the arterial input TAC has a consistent pattern from study to study, and 2) the area under the curve of TAC is proportional to the injected dose/body weight. Our study clearly indicated the violation of these assumptions in the case of TSPO imaging. Therefore, only using brain SUV to quantify TSPO binding has its limitations.

## **Effect of blood centrifugation**

Switching to immediate centrifugation improved the test-retest reliability in blood significantly. Since TSPO is also expressed in blood e.g. in monocytes (2), we hypothesize that <sup>123</sup>I-CLINDE continues to bind to TSPO in blood cells and intact <sup>123</sup>I-CLINDE in a blood sample will gradually bind to blood cells causing an underestimation of plasma radioactivity. Our hypothesis is supported by the increase in PoB ratio by immediate centrifugation. The negative correlation between PoB in MABs and monocyte counts also indicate that CLINDE binding is related to TSPO expression in blood. Additionally, the lower starting point of PoB TAC of HABs than TAC of MABs (Supplemental Fig. 1) indicates that binding of CLINDE to blood cells is dependent on the affinity/TSPO polymorphism (21). Previously, in (22) an extra compartment was proposed to describe TSPO ligand binding to the vascular walls. Based on our observations,

the "endothelial compartment" may additionally include binding TSPO in blood cells. However, fast and systematic centrifugation of blood samples will still be required to produce consistent results.

#### **CONCLUSION**

The variability of <sup>123</sup>I-CLINDE binding was investigated in a test-retest setting with 16 healthy subjects. The test-retest variability estimated by ICC and percentage difference was very favourable compared to the first-generation TSPO ligand <sup>11</sup>C-PK11195, and comparable to or slightly better than second-generation TSPO ligands <sup>11</sup>C-DPA-713 and <sup>11</sup>C-PBR28. SUV as a surrogate of the output measure V<sub>T</sub> from 'gold standard'-2TCM has its limitations. Immediate centrifugation of blood samples is essential and can prevent underestimation of the plasma input function. A population-based method could efficiently recover data with delayed centrifugation, and has the potential to be applied to studies using other TSPO ligands.

#### **DISCLOSURE**

There is no conflict of interest.

## **ACKNOWLEDGMENTS**

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Figure 1 The effect of population-based (PB) approach to adjust input functions (IF) with delayed centrifugation (A, B) and the standard uptake values (SUV) of IF and a brain region of mixed affinity binders (MAB) (A, C) and high affinity binders (HAB) (B, D). (A) and (B) show the SUVs of a MAB (Subj 6) and a HAB (Subj 3), separately, both with delayed centrifugation of blood samples. Two tissue compartment model (2TCM) fitting to neocortex using the input functions (red curves) were also given. (C) and (D) show the SUVs of a MAB (Subject 10) and a HAB (Subject 14), both with immediate centrifugation of blood samples.

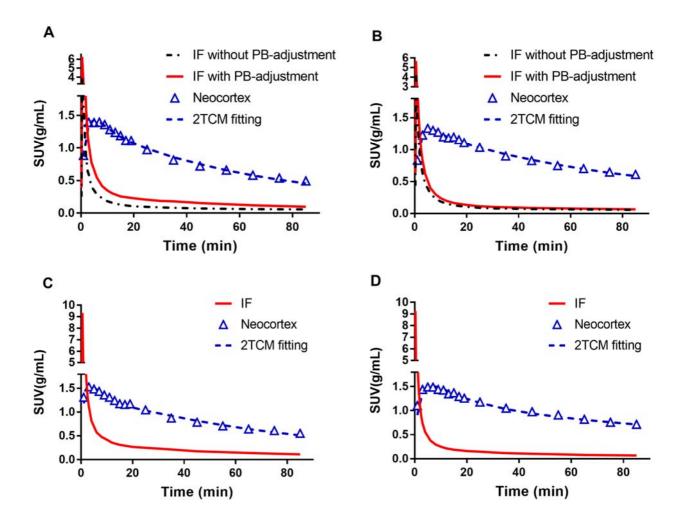


Figure 2 Effect of immediate centrifugation of blood samples on plasma to whole-blood ratio (PoB)

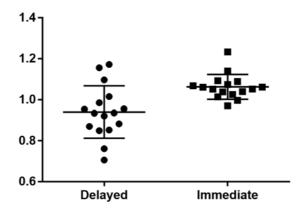


Figure 3 Cerebellar brain mean standard uptake values (mSUV) (A), mSUV from 60 to 90 min (B) and distribution volume V<sub>T</sub>(C) categorized by genotype: mixed affinity binder (MAB) and high affinity binder (HAB)

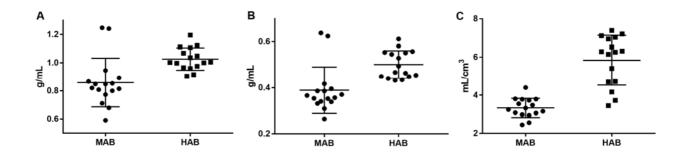


Figure 4 Brain mean standard uptake values (mSUVs) of a typical mixed affinity binder (MAB): subject 12 and a high affinity binder (HAB): subject 11.

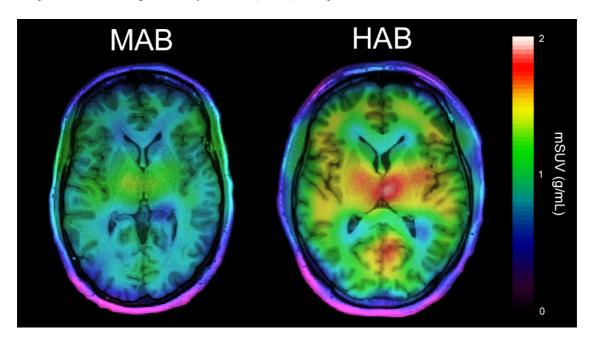


Table 1 Demographic data and methodological information specified by subject

| Subject | Gender | Age<br>(yrs) | Body<br>weight<br>(kg) | TSPO<br>genotype | Injected activity (MBq) | Scan<br>interval<br>(days) | Centrifugation | Purity<br>(%) | Plasma<br>control<br>(%) |
|---------|--------|--------------|------------------------|------------------|-------------------------|----------------------------|----------------|---------------|--------------------------|
| 1       | M      | 61           | 80                     | MAB              | 120.9                   | 21                         | delayed        | 96.1          | 95.4                     |
|         |        |              |                        |                  | 117.8                   |                            |                | 93.5          | 91.4                     |
| 2       | M      | 70           | 92                     | MAB              | 112.2                   | 42                         | delayed        | 95.4          | 95.4                     |
|         |        |              |                        |                  | 123.6                   |                            |                | 94.9          | 93.8                     |
| 3       | M      | 59           | 76.5                   | HAB              | 118.1                   | 35                         | delayed        | 96.4          | 94.7                     |
|         |        |              |                        |                  | 112.0                   |                            |                | 94.9          | 93.2                     |
| 4       | F      | 38           | 110                    | HAB              | 116.3                   | 28                         | delayed        | 96.0          | 95.0                     |
|         |        |              |                        |                  | 114.3                   |                            |                | 90.7          | 88.4                     |
| 5       | F      | 62           | 70                     | HAB              | 119.0                   | 42                         | delayed        | 96.1          | 95.1                     |
|         |        |              |                        |                  | 119.4                   |                            |                | 96.6          | 94.0                     |
| 6       | F      | 58           | 50                     | MAB              | 116.0                   | 28                         | delayed        | 96.1          | 96.2                     |
|         |        |              |                        |                  | 119.7                   |                            |                | 95.3          | 91.4                     |
| 7       | F      | 68           | 87                     | HAB              | 124.8                   | 56                         | delayed        | 92.3          | 90.1                     |
|         |        |              |                        |                  | 115.1                   |                            |                | 95.2          | 90.5                     |
| 8       | M      | 52           | 94                     | HAB              | 118.5                   | 56                         | delayed        | 98.1          | 93.8                     |
|         |        |              |                        |                  | 124.1                   |                            |                | 89.8          | 88.5                     |
| 9       | M      | 42           | 80                     | MAB              | 118.8                   | 28                         | immediate      | 95.7          | 94.6                     |
|         |        |              |                        |                  | 115.8                   |                            |                | 93.4          | 90.3                     |
| 10      | F      | 45           | 86.2                   | MAB              | 117.4                   | 56                         | immediate      | 95.4          | 94.6                     |
|         |        |              |                        |                  | 114.1                   |                            |                | 94.9          | 94.7                     |
| 11      | M      | 35           | 100                    | HAB              | 119.9                   | 56                         | immediate      | 93.0          | 93.2                     |
|         |        |              |                        |                  | 114.4                   |                            |                | 90.7          | 89.9                     |
| 12      | F      | 40           | 73                     | MAB              | 112.0                   | 7                          | immediate      | 92.8          | 91.8                     |
|         |        |              |                        |                  | 130.7                   |                            |                | 94.7          | 94.1                     |
| 13      | M      | 24           | 95.6                   | MAB              | 115.4                   | 14                         | immediate      | 94.4          | 92.2                     |
|         |        |              |                        |                  | 116.0                   |                            |                | 92.0          | 90.2                     |
| 14      | F      | 69           | 83                     | MAB              | 123.0                   | 28                         | immediate      | 94.5          | 92.7                     |
|         | _      |              |                        |                  | 127.1                   |                            |                | 94.4          | 90.4                     |
| 15      | F      | 30           | 68                     | HAB              | 105.8                   | 42                         | immediate      | 91.0          | 82.9                     |
|         |        |              |                        |                  | 116.0                   |                            |                | 91.1          | 88.1                     |
| 16      | F      | 34           | 74.2                   | HAB              | 118.4                   | 28                         | immediate      | NA            | NA                       |
| 3.6     |        | 40 :         | 02.5:                  |                  | 116.3                   | 25.                        |                | NA            | NA                       |
| Mean    |        | 49±          | 82.5±                  |                  | 117.9±                  | 35±                        |                | 93.4±         | 91.4±                    |
| ±STD    |        | 15           | 14.4                   |                  | 4.9                     | 15                         |                | 1.7           | 3.2                      |

Note: HAB: high affinity binder; MAB: mixed affinity binder; STD: standard deviation; NA: not available.

Table 2 Test-retest variability

|                 | V <sub>T</sub> using por<br>input funct | r using population-adjuste input function of delayed | justed | $V_{T} \ using \ population-adjusted \\ input \ function \ of \ delayed \\ N=8$ | iate centrifug<br>N=8 | gation |                          | $V_T$ of all | V <sub>T</sub> of all subjects N=16 | =16    |        |                 | Brain upt | Brain uptake N=16 |        |        |
|-----------------|---|--|--------|---|-----------------------|--------|--------------------------|--------------|-------------------------------------|--------|--------|-----------------|-----------|-------------------|--------|--------|
|                 | centrifu                                | centrifugation N=8                                   | ×      |   |                       |        |                          |              |                                     |        |        |                 |           |                   |        |        |
| Regions         | PD (%) Absolute ICC                     | Absolute   | ICC    | PD (%)  | Absolute              | ICC    | PD (%)                   | Absolute     |                                     | ICC of | ICC of | PD (%)          | Absolute  | ICC of            | ICC of | ICC    |
|                 |   | PD (%)   |        |   | PD (%)                |        |                          | PD (%)       | MABs                                | HABs   | all    |                 | PD (%)    | MAB               | HAB    | of all |
| Midbrain        | $2.8\pm14.5$                            |  | 96.0   | 12.4 0.96 $-1.5 \pm 15.6$   | 11.8                  | 98.0   | $0.6\pm14.8$             | 12.1         | 0.41                                | 0.91   | 0.93   | $-2.5 \pm 11.5$ | 10.1      | 0.72              | 0.39   | 0.71   |
| Thalamus        | $11.0\pm10.5$                           | 12.8   |        | 0.94 $-12.3 \pm 13.3$   | 12.7                  | 98.0   | $\textbf{-0.7} \pm 16.7$ | 12.8         | 89.0                                | 0.84   | 06.0   | $-4.7 \pm 14.6$ | 11.6      | 0.74              | 0.53   | 0.71   |
| Pons            | $-2.3 \pm 14.2$                         | 9.6  | 0.90   | 0.90 $-8.2 \pm 23.8$  | 18.9                  | 0.72   | $-5.3 \pm 19.2$          | 14.2         | 0.54                                | 09.0   | 0.82   | $-2.7 \pm 12.4$ | 11.0      | 0.81              | 0.52   | 0.77   |
| Pallidostriatum | $5.8\pm21.9$                            | 17.6   | 0.85   | $-7.0 \pm 19.0$   | 14.1                  | 0.87   | $-0.6 \pm 20.9$          | 15.8         | 0.22                                | 0.79   | 0.85   | $-2.2 \pm 13.7$ | 11.1      | 0.79              | 0.50   | 0.74   |
| Hippocampus     | $14.9\pm17.3$                           | 15.7   | 0.75   | $-5.9 \pm 11.8$   | 9.7                   | 0.91   | $4.5\pm17.9$             | 12.7         | 0.58                                | 0.78   | 0.83   | $-2.0 \pm 15.2$ | 12.3      | 0.57              | 0.23   | 0.56   |
| Cerebellum      | $9.6 \pm 9.4$                           | 10.2   | 0.94   | 0.94 $-4.9 \pm 10.0$  | 8.2                   | 96.0   | $2.3\pm12.0$             | 9.2          | 92.0                                | 0.88   | 0.95   | $-1.5 \pm 9.4$  | 7.8       | 06.0              | 0.45   | 0.87   |
| Neocortex       | $5.8\pm12.6$                            | 10.2   | 0.92   | 0.92 $-3.2 \pm 12.8$  | 10.5                  | 0.92   | $1.3\pm13.2$             | 10.4         | 0.88                                | 0.79   | 0.92   | $-2.8 \pm 9.6$  | 8.5       | 0.91              | 0.57   | 0.85   |
| 4               |   | 1.00   |        | (   |                       | ١      | •                        |              |                                     |        |        |                 |           |                   |        |        |

Note: PD: percentage difference; ICC: intraclass correlation coefficient; VT: distribution volume.

# Supplemental Material

Figure 1 Plasma to whole-blood ratio of HAB (N=2x3) and MAB (N=2x5) with immediate centrifugation

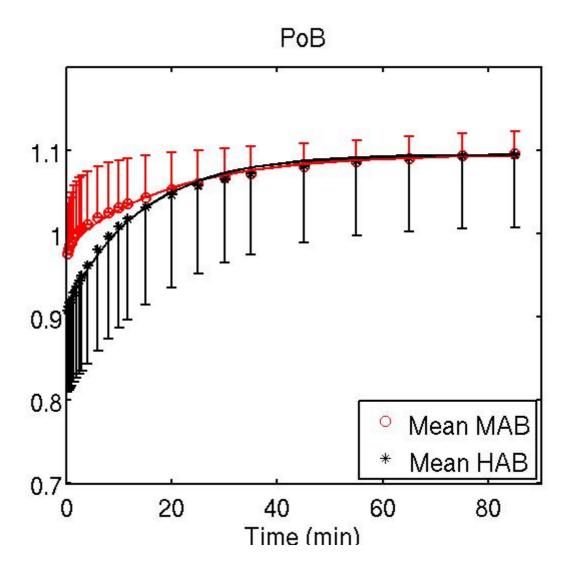


Figure 2 Test-retest absolute percentage difference of radioligand concentration in plasma and metabolite-corrected plasma input function before applying the population-adjusted method: Plasmsa Cp and Plasma input function Cmcp; and after: Population-based Cp (PB-Cp) and PB-Cmcp

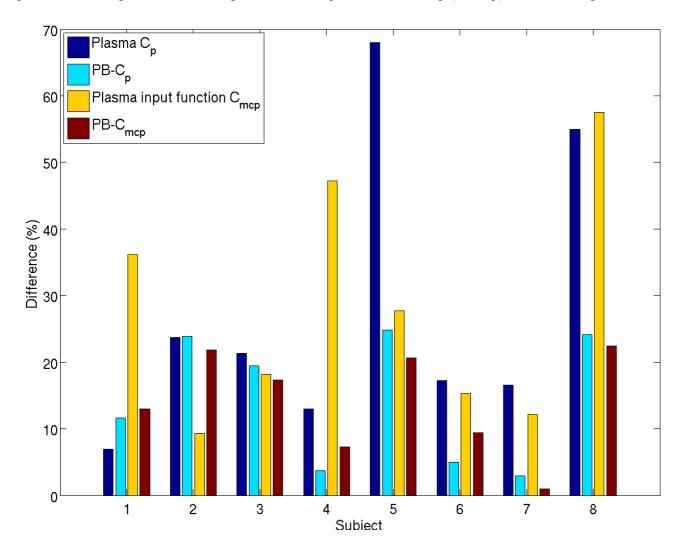


Table 1 Regional distribution volume  $V_T$  (mL/cm<sup>3</sup>) from 2TCM of the immediate centrifuged scans for high affinity binders (HAB) and mixed affinity binders (MAB)

| ID              | HA   | AΒ     | HA   | AΒ     | HA   | AΒ     | M    | AΒ     | M    | AB     | M    | AB     | M    | AΒ     | M    | AB     |
|-----------------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|
| 1               | Test | Retest |
| Regions         | 11   | 11     | 15   | 15     | 16   | 16     | 9    | 9      | 10   | 10     | 12   | 12     | 13   | 13     | 14   | 14     |
| Midbrain        | 7.5  | 9.1    | 7.1  | 6.5    | 10.2 | 13.4   | 6.3  | 5.2    | 5.1  | 5.1    | 6.1  | 5.3    | 5.0  | 5.0    | 4.1  | 4.3    |
| Thalamus        | 8.3  | 9.5    | 7.4  | 7.3    | 8.2  | 11.7   | 3.1  | 4.2    | 3.5  | 3.5    | 4.4  | 4.7    | 4.6  | 4.9    | 5.0  | 5.5    |
| Pons            | 8.5  | 6.4    | 8.8  | 8.1    | 8.2  | 12.0   | 3.3  | 4.1    | 5.3  | 5.8    | 4.7  | 4.4    | 4.5  | 6.8    | 4.6  | 4.6    |
| Pallidostriatum | 6.5  | 8.9    | 7.3  | 7.3    | 8.8  | 8.0    | 3.2  | 3.4    | 3.5  | 2.9    | 3.5  | 3.6    | 3.9  | 5.6    | 3.8  | 4.1    |
| Hippocampus     | 5.6  | 7.4    | 6.0  | 5.8    | 7.1  | 7.5    | 3.1  | 3.1    | 2.9  | 3.5    | 3.7  | 4.0    | 6.2  | 5.7    | 4.1  | 4.4    |
| Cerebellum      | 5.4  | 6.3    | 6.3  | 6.3    | 6.5  | 7.0    | 2.4  | 3.1    | 3.2  | 3.0    | 3.5  | 3.3    | 3.5  | 3.8    | 3.7  | 3.8    |
| Neocortex       | 5.0  | 6.0    | 6.4  | 6.1    | 6.1  | 7.2    | 2.3  | 2.7    | 3.5  | 3.1    | 3.3  | 3.3    | 4.5  | 4.1    | 3.9  | 4.0    |

Table 2 Regional distribution volume  $V_T$  (mL/cm<sup>3</sup>) from 2TCM of the delayed centrifuged scans without population-based adjustment of the metabolite-corrected input function, but with parent fraction adjustment for high affinity binders (HAB) and mixed affinity binders (MAB)

| ID              | M    | AВ     | M    | AB     | M    | AВ     | HA   | AΒ     | H    | AΒ     | HA   | AΒ     | H    | AΒ     | HA   | AΒ     |
|-----------------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|
| 1               | Test | Retest |
| Regions         | 1    | 1      | 2    | 2      | 6    | 6      | 3    | 3      | 4    | 4      | 5    | 5      | 7    | 7      | 8    | 8      |
| Midbrain        | 5.0  | 5.8    | 4.7  | 4.4    | 10.2 | 10.5   | 7.5  | 12.3   | 8.7  | 7.3    | 12.9 | 9.5    | 7.1  | 6.2    | 7.5  | 5.7    |
| Thalamus        | 5.6  | 6.7    | 4.4  | 3.9    | 9.6  | 10.6   | 7.6  | 11.2   | 8.3  | 6.5    | 12.8 | 7.4    | 5.1  | 4.7    | 9.0  | 5.5    |
| Pons            | 4.4  | 4.7    | 4.4  | 4.5    | 9.3  | 10.8   | 6.4  | 12.5   | 9.0  | 10.4   | 11.4 | 11.2   | 7.1  | 5.3    | 8.1  | 6.8    |
| Pallidostriatum | 3.5  | 4.6    | 4.3  | 3.9    | 7.5  | 8.6    | 6.1  | 11.6   | 7.3  | 5.2    | 9.4  | 8.2    | 4.8  | 4.1    | 5.3  | 4.9    |
| Hippocampus     | 4.5  | 4.7    | 3.1  | 3.5    | 8.2  | 7.5    | 6.0  | 9.9    | 7.3  | 8.6    | 8.9  | 7.0    | 4.5  | 4.8    | 7.4  | 4.4    |
| Cerebellum      | 3.6  | 3.8    | 2.9  | 2.9    | 6.7  | 7.1    | 5.7  | 9.5    | 6.9  | 5.0    | 8.3  | 6.0    | 4.7  | 4.0    | 5.3  | 3.9    |
| Neocortex       | 3.3  | 3.8    | 3.0  | 2.7    | 6.5  | 7.1    | 5.0  | 8.0    | 6.5  | 4.6    | 8.1  | 5.8    | 4.6  | 4.0    | 5.4  | 3.6    |

Table 3 Regional distribution volume  $V_T$  (mL/cm<sup>3</sup>) from 2TCM of the delayed centrifuged scans with population-based adjustment of the metabolite-corrected input function for high affinity binders (HAB) and mixed affinity binders (MAB)

| ID              | M    | AB     | M    | AВ     | M    | AВ     | HA   | AΒ     | HA   | AΒ     | HA   | AB     | HA   | AΒ     | HA   | AB     |
|-----------------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|------|--------|
|                 | Test | Retest |
| Regions         | 1    | 1      | 2    | 2      | 6    | 6      | 3    | 3      | 4    | 4      | 5    | 5      | 7    | 7      | 8    | 8      |
| Midbrain        | 4.7  | 5.4    | 5.4  | 4.9    | 6.9  | 5.4    | 15.4 | 17.6   | 7.8  | 8.8    | 11.1 | 10.7   | 5.7  | 4.8    | 6.3  | 5.8    |
| Thalamus        | 4.5  | 4.7    | 5.1  | 4.6    | 6.6  | 5.3    | 9.5  | 8.7    | 7.7  | 6.0    | 13.9 | 12.5   | 4.1  | 4.2    | 7.2  | 6.0    |
| Pons            | 3.7  | 3.2    | 5.0  | 5.1    | 5.8  | 5.4    | 8.8  | 12.2   | 7.8  | 7.6    | 11.2 | 12.4   | 5.8  | 5.6    | 7.4  | 7.6    |
| Pallidostriatum | 5.0  | 3.1    | 4.4  | 4.4    | 4.9  | 4.4    | 7.4  | 9.4    | 6.8  | 5.5    | 8.6  | 10.0   | 3.9  | 3.3    | 4.6  | 5.1    |
| Hippocampus     | 5.0  | 3.2    | 4.2  | 4.4    | 5.1  | 3.6    | 7.4  | 7.1    | 5.8  | 5.6    | 7.3  | 6.8    | 4.4  | 4.2    | 5.3  | 4.2    |
| Cerebellum      | 2.9  | 2.6    | 3.2  | 3.1    | 4.4  | 3.8    | 7.2  | 7.4    | 6.2  | 4.7    | 7.1  | 7.0    | 3.7  | 3.4    | 4.7  | 4.2    |
| Neocortex       | 2.3  | 2.6    | 3.2  | 3.1    | 4.2  | 3.7    | 6.1  | 6.4    | 5.8  | 4.2    | 7.1  | 6.6    | 3.8  | 3.4    | 4.5  | 4.4    |

# Study 4

Expression, evolution and prognostic value of translocator protein in middle cerebral artery stroke patients - A longitudinal <sup>123</sup>I-CLINDE-SPECT imaging study.

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Running title: TSPO in stroke

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Word count: 5490

# **Abstract**

#### Introduction

Molecular imaging of the translocator protein (TSPO) may provide insight into regional neuroinflammation following ischemic stroke. In this study, we measured the regional cerebral TSPO expression over time after ischemic stroke using <sup>123</sup>I-CLINDE single photon emission computed tomography (SPECT) imaging.

### **Methods**

Twelve patients (ten high affinity (HAB) and two mixed affinity binders (MAB)) who suffered a first-ever ischemic middle cerebral artery stroke and ten healthy volunteers (all HAB) were recruited for the study. Patients underwent concomitant <sup>123</sup>I-CLINDE SPECT, structural magnetic resonance imaging (MRI) and were clinically tested between week 1-2 (scan 1), week 5-6 (scan 2), and week 15-18 weeks (scan 3). Patients were additionally clinically tested at 25-26 weeks. The healthy volunteers were scanned once with <sup>123</sup>I-CLINDE SPECT and structural MRI. TSPO expression was measured in lesional and perilesional regions of interest (ROIs) as well as in ROIs connected to the lesion and ROIs unrelated to the lesion. Furthermore, The necrotized volume was determined from the structural MRI at scan 3.

#### Results

Lesional TSPO expression was high at scan 1 (median  $V_T = 10.5 \text{ mL/cm}^3$ ) and decreased by 35.6% from scan 1 to 3 (p=0.004). In contrast, TSPO expression in brain regions unrelated to the stroke lesion was significantly lower in stroke patients at scan 1 compared to similar ROIs in healthy volunteers (p=0.001), but increased over time to values similar to those of healthy volunteers. No significant correlations between initial <sup>123</sup>I-CLINDE binding and clinical test scores found. However, the initial overlap percentage of the T2-weighted lesion ROI and the <sup>123</sup>I-CLINDE lesion ROI correlated negatively with the volume of lesion necrosis on structural MRI at scan 3 (p=0.030).

#### **Conclusions**

This study shows a remarkable intersubject variation in the spatial and temporal pattern of neuroinflammation in patients after a MCA stroke with cortical involvement. TSPO expression in periinfarct or remote areas to the stroke did not correlate with clinical test scores. This implies

that TSPO expression after stroke is multifaceted and cannot be attributed to either neuroprotective or detrimental effects of neuroinflammation.

Initial TSPO levels were lower in regions unrelated to the stroke when compared to healthy subjects and normalized after 4 months, which may be an effect of anti-inflammatory cytokine release or stroke induced immunomodulation.

# Introduction

Ischaemic stroke remains a leading cause of acquired disability and a source of increasing disease burden worldwide despite thrombolytic therapy and novel treatment strategies, such as acute mechanical thrombectomy of occluded cerebral vessels ('WHO | The Atlas of Heart Disease and Stroke'; Berkhemer *et al.*, 2015). In the weeks and months following an ischemic stroke, rehabilitation treatment focus solely on stroke care, physio- and occupational therapy in order to improve outcome. No medical treatment has proved effective to enhance functional recovery of cerebral stroke patients during the rehabilitation period. Additional research in the cellular and functional changes of the recovering brain is warranted.

Upon brain damage, such as after stroke, microglia in the brain may transform from a 'resting' ramified phenotype into a 'reactive' state and move to the site of the lesion (Davalos et al., 2005). Reactive microglia together with invading blood borne macrophages may have detrimental effects on patient recovery because of excessive collateral damage to healthy tissue (Huang et al., 2013), but also has the potential to augment functional recovery by suppressing inflammation and facilitate neurogenesis (Cherry et al., 2014). Glial reactivity after brain injury may be investigated by molecular imaging of the 18 kDa translocator protein (TSPO), which has been applied in both experimental stroke models and, to a limited extent, in human stroke patients (Boutin and Pinborg, 2015). The largest previous study in human stroke by Thiel et al. (2010) included 16 patients with subcortical stroke scanned longitudinally within the first 3 weeks and at 6 months follow-up with <sup>11</sup>C-PK11195 positron emission tomography (PET) and diffusion tensor MRI (DTI). The study established a correlation between pyramidal tract damage quantified with DTI and increased <sup>11</sup>C-PK11195 uptake in the brainstem at 6 months follow up (Thiel et al., 2010). A study by Gerhard et al. (2005) found increased expression of TSPO in connected ipsilesional areas and the contralesional thalamus at 150 days after stroke onset. Furthermore, when controlling for pyramidal tract damage a negative partial correlation was discovered between initial <sup>11</sup>C-PK11195 uptake in the brainstem and follow-up motor function 12

as evaluated by the Rivermead Motor Function Test. It has been hypothesised that TSPO upregulation in areas connected to the lesion can show effects of Wallerian degeneration but also in reactive microglia with beneficial properties for stroke rehabilitation (Boutin and Pinborg, 2015).

<sup>123</sup>I-CLINDE is a second generation TSPO tracer for single photon computed tomography (SPECT) imaging and has previously been used in human studies with stroke, glioblastoma multiforme and anti N-methyl-D-aspartate receptor encephalitis (Feng *et al.*, 2014; Jensen *et al.*, 2015; 2015b). Recently, the <sup>123</sup>I-CLINDE SPECT modality has been validated in a test-retest study showing similar test-retest variability as the two second-generation TSPO PET tracers <sup>11</sup>C-PBR28 and <sup>11</sup>C-DPA713 (Feng *et al.*, 2016). <sup>123</sup>I-CLINDE is a second generation TSPO tracer and therefore susceptible to the rs6971 polymorphism which affects the affinity of tracer to TSPO (Owen *et al.*, 2012).

The aim of the current study was to accurately describe the temporal and spatial development in TSPO expression from 2 to 18 weeks in MCA stroke patients with cortical involvement and relate finding to clinical test scores.

## We hypothesize:

- Increased perilesional binding of <sup>123</sup>I-CLINDE to TSPO at 1-2 weeks is a biomarker of poor recovery at 25-26 weeks.
- Increased binding of <sup>123</sup>I-CLINDE to TSPO in the ipsilesional thalamus and pons at 1-2 weeks is a biomarker of poor recovery at 25-26 weeks.
- Increased binding of <sup>123</sup>I-CLINDE to TSPO in the contralesional thalamus at 5-6 weeks is a biomarker of good recovery at 25-26 weeks.

# **Materials and methods**

## Patients, healthy volunteers and genotyping

This study was conducted in accordance with the Declaration of Helsinki at the Copenhagen University Hospital, Rigshospitalet, Denmark. The ethical committee of the Copenhagen Capital Region (H-2-2010-086 amendment 39319) approved the study protocol. All subjects provided written informed consent. Patients were recruited from Bispebjerg hospital and Rigshospitalet, Copenhagen University hospital, Denmark. Twelve stroke patients with first-ever infarction in the MCA territory, a lesion involving cortical brain tissue and impairment of the contralesional

upper extremity were included within the first week after onset. Patients were scanned longitudinally three times at 1-2, 5-6 and 15-18 weeks after stroke onset with concomitant <sup>123</sup>I-CLINDE SPECT and structural MRI. Table 1 shows the patient data and Figure 1 shows the study design. All subjects were genotyped for the rs6971 polymorphism to determine TSPO binder-status as described previously (Feng *et al.*, 2014) revealing 10 HABs and 2 mixed MAB. Accordingly, a cohort of healthy volunteers was genotyped and 10 healthy volunteers with the HAB genotype were included in the study and <sup>123</sup>I-CLINDE SPECT and structural MRI scanned once. All healthy volunteers had normal physical and neurological examinations, and blood tests. Table 2 shows the healthy volunteer data.

## Image acquisition and arterial blood sampling

## [123 I]CLINDE-SPECT scanning

To block thyroidal uptake of free radioiodine, 200 mg of potassium perchlorate was administered intravenously 20 min before injection of <sup>123</sup>I-CLINDE. A headband was applied to minimize head movements during scanning. Dynamic SPECT scanning with a triple-head IRIX camera (Philips Medical) was started simultaneously with a bolus injection of <sup>123</sup>I-CLINDE (MAP Medical Technologies), and lasted for 90 minutes. The acquisition protocol consisted of 10 x 2-minute frames, followed by 7 x 10-minute frames.

#### **Blood sampling**

Blood samples were drawn manually from a cannula inserted into the radial artery arterial at 0.25, 0.5, 0.75, 1, 1.5, 2, 2.5, 3, 4, 6, 8, 10, 12, 15, 20, 25, 30, 35, 45, 65 and 85 min for measuring radioactivity in plasma and whole-blood. Additionally, 8 blood samples were drawn at 0.5, 4, 10, 20, 30, 45, 65 and 85 min for metabolite analysis by radio-high performance liquid chromatography (radio-HPLC). Fractions of HPLC eluent were collected using a fraction collector device (Foxy Jr FC144, Teledyne Isco) and counted off-line in a well-counter (2480 Wizard2 Gamma Counter, Perkin Elmer) for accurate measurements of parent tracer. During the study we discovered that the handling of blood-samples affects the test-retest variance (Feng *et al.*, 2016) and due to this, the time between sampling and centrifugation was changed from within 90 minutes to 5 minutes. Subject specific details on the centrifugation can be found in Table 1 for patients, all blood samples from healthy volunteers were centrifuged immediately. Total radioactivity in plasma and whole-blood was measured in a well-counter (Cobra 5003; Packard Instruments) and data were decay-corrected to the time of injection. Furthermore, the radioligand *purity* was measured for each batch of <sup>123</sup>I-CLINDE, and trapping efficiency of the

HPLC column (*plasma control*) was determined twice for each blood sample batch by spiking water and blank plasma with <sup>123</sup>I-CLINDE. All equipment was cross-calibrated to the SPECT scanner. Blood screening including white blood cell differential count was performed at each scan session.

#### Structural MRI

Three-dimensional T1- and T2-weighted magnetic resonance images (MRI) were acquired within 24 hours of the corresponding [<sup>123</sup>I]CLINDE-SPECT for patients and healthy volunteers. Voxel-size was 1x1x1 mm for both T1 and T2 images. MRI was performed using a 3-T Prisma scanner (Siemens, Erlangen). A trained neurologist interpreted the structural MRI scans from all healthy volunteers to be normal.

## Image analysis

### Image pre-processing and regions of interest

For each subject, the T1- and T2-weighted MRIs were co-registered to the time-weighted mean SPECT image by interactive image overlay (Willendrup et al., 2004). Regions of interest (ROIs) were delineated both automatically and manually. Automatic ROI delineation was performed on both patients and HC's using probability maps using a data processing pipeline as described previously (Svarer et al., 2005). Automatically delineated ROIs were defined according to the side of the lesion for the individual patients and labelled as either ipsi- or peri-lesional. Five automatically delineated ROIs were investigated in this study: Ipsilesional cerebellum, contralesional occipital cortex, ipsilesional thalamus, contralesional thalamus and pons. Regions connected to the lesion was defined as: Ipsi- and contralesional thalamus and pons. Regions unrelated to the lesion was defined as: Ipsilesional cerebellum and contralesional occipital cortex. Stroke lesion ROIs were manually delineated on the MRI and SPECT images acquired at the first time point. Structural T2-weighted lesion ROIs were delineated around the area of the stroke, shown as the high intensity voxels on T2-weighted structural MRIs. Further, a SPECT lesion ROI was delineated on the SPECT image by automatically selecting voxels in the vicinity of the stroke with a value above 1.5 times the mean cerebellar <sup>123</sup>I-CLINDE uptake. Finally, a perilesional SPECT ROI was defined by subtracting the manually delineated T2-weighted lesion ROIs from the SPECT lesion ROI. Regions of lesion necrosis were manually delineated at the boundary of the region which shows loss of T1-weigted MRI signal at scan 3. Volume sizes of manually delineated ROIs and the overlap between SPECT and MRI regions were estimated.

The ipsilesional cerebellum ROI for patient no. 10 and the cerebellum ROI for healthy volunteer no. 3 were excluded due to the SPECT recording not encompassing total cerebellum. See Figure 2 for examples of the T2-weighted and SPECT lesion ROIs. Due to the limited spatial resolution of SPECT images, no segmentation of grey matter, white matter and cerebrospinal fluid was attempted. All pre-processing was performed using Matlab R2013a (Mathworks Inc.).

### Kinetic modelling

Parent fraction (PF) curves were adjusted to account for the tracer trapped in the HPLC as:  $PF_{adj} = PF \times (1 + purity - plasma\ control)$ .

PF measurements were lost due to failure of the HPLC apparatus at scan 3 of patient 11 and scan 2 of patient 12. In these two occasions, The  $PF_{adj\_mean}$  was estimates as the mean of the two remaining  $PF_{adj}$  curves.

<sup>123</sup>I-CLINDE binding was quantified as distribution volume ( $V_T$ ) by a two-tissue compartment model (rate constants:  $K_1$ ,  $k_2$ .  $k_3$  and  $k_4$ , with a fixed blood volume in brain tissue of 5%) as described in (Feng *et al.*, 2016). Kinetic modelling was done using PMOD 3.0 (PMOD Technologies Inc., Switzerland).

### Clinical evaluation

At each scan and clinical follow-up, patients were evaluated under supervision by a trained neurologist performing a battery of rehabilitation measures (Figure 1). Stroke severity and functional status were assessed by National Institute of Health Stroke Scale (NIHSS), Scandinavian Stroke Scale (SSS), Fugl-Meyer Assessment of the Upper Extremity (FMA-UE), Motricity Index (MI), Action Research Arm Test (ARAT), Nine-Hole Peg Test (NHPT), Grip strength (GS), Modified Ranking Scale (MRS) and Barthel Index (BI). NHPT and GP was measured for both affected and non-affected extremity three times and the mean fraction of the affected vs. non-affected extremity was calculated. Recovery was calculated for each measure as the PD between scan 1 and 25 week follow-up.

### Statistical analysis

Data are presented as medians [1<sup>st</sup> quartile; 3<sup>rd</sup> quartile] for continuous variables and percentage for categorical variables.

To investigate whether TSPO expression in stroke patients changed over time, we compared regional  $V_T$  at the three time points. However, due to the disparity in affinity of  $^{123}$ I-CLINDE to TSPO between HAB and MAB, two separate analyses were performed. In the first analysis, we disregarded the two MAB patients and performed repeated measures ANOVAs on the group of the remaining ten HAB patients, with separate models for each ROI. The magnitude of the change over time was additionally quantified by estimating the relative percentage differences (PD) between scan 1 and scan 3, computed as:

$$PD_{V_T} = \frac{mean \, V_{T \, scan \, 3} - mean \, V_{T \, scan \, 1}}{mean \, V_{T \, scan \, 1}} \times 100\%$$

Where the mean  $V_T$  is mean value for all subjects.

In the second analysis, to include the total patient cohort of HABs and MABs, we computed the PD between scan 1 and 3 as above, for all twelve patients, and performed a two-sided one-sample t-test.

To explore the association between TSPO and other experimental variables, we performed Pearson correlations for the following experimental variables:

- To compare TSPO expression to clinical outcome, the regional V<sub>T</sub> at scan 1 (lesion, perilesional, ipsi- and contralesional thalamus, and pons) was correlated to PD of rehabilitation measures between scan 1 and the 25-26 week follow-up.
- The same correlation was done for the regional  $V_T$  at scan 2 and the PD of rehabilitation measures between scan 2 and the 25-26 week follow-up.
- To explore the finding from experimental studies that TSPO is expressed predominantly perilesionally in permanent ischemia in our data, the percentage overlap between the <sup>123</sup>I-CLINDE lesion and the T2-lesion at scan 1 was correlated to the percentage atrophy of the T2-lesion at scan 3 and also correlated to the PD of rehabilitation measures between scan 1 and follow-up.

To investigate the impact of stroke on brain regions that was unrelated to the stroke lesion,  $V_T$  of ipsilesional cerebellum and contralesional occipital cortex for stroke patients at scan 1,2 and 3 were compared to the  $V_T$ s of cerebellum and occipital cortex in healthy volunteers using Mann-Whitney U tests.

Repeated measure ANOVA, t-test, pearson correlation, and Mann-Whitney U tests were performed using Prism 6.0c (Graphpad Software Inc.). Percentage difference was calculated using Matlab (Mathworks inc).

# **RESULTS**

The clinical characteristics of the stroke patients and healthy volunteers are summarized in Table 1 and 2. Descriptive statistics are given in table 3.

In general, the TSPO expression showed massive variance. This was evident for the initial scans where the size and intensity of the CLINDE uptake was very different for the patients.

Furthermore, the heterogeneity in change in expression between scan 1, 2 and 3 was generally sizeable. See figure 3 for a visual comparison of all patients.

# Lesional and perilesional TSPO expression in stroke patients

The repeated measures ANOVA showed a significant decrease in lesional  $V_T$  for the HAB patients over the three scans (p=0.004) and a trend towards a perilesional decrease (p=0.07). Distribution volumes for HAB stroke patients at scan 1,2 and 3 were  $10.5 \pm 2.3$ ,  $8.16 \pm 2.0$  and  $6.9 \pm 1.2$  mL/cm<sup>3</sup>, respectively. This corresponds to a relative decrease between scans 1 and 3 of 34.7%. Performing a two tailed t-test on the PD from scan 1 to 3 of lesional and perilesional  $V_T$  on all 12 patients revealed a significant mean decrease of 22.6 % lesionally (p=0.04) and 21.3 % perilesionally (p=0.001). The temporal evolutional of lesional and perilesional  $^{123}$ I-CLINDE uptake is visualised on  $^{123}$ I-CLINDE SPECT images in Figure 2 and 3.

## Correlations between TSPO expression, atrophy and clinical outcome measures

We did not find any correlations between  $V_T$  of  $^{123}$ I-CLINDE lesionally, perilesionally, pons or ipsi- and contralesional thalamus at scan 1 or 2 and the recovery in rehabilitation measures. See figure 5 for a Pearson correlation plot between  $V_Ts$  of  $^{123}$ I-CLINDE and recovery.

A negative correlation was found between the percentage overlap of SPECT and T2-weighted structural lesion at scan 1 and the volume of lesion necrosis at scan 3 (p=0.001).

## TSPO expression in ROIs unrelated to the lesion

Comparing  $^{123}$ I-CLINDE binding in ipsilesional cerebellum and the perilesional occipital cortex of 10 HAB stroke patients to the occipital cortex of 10 HAB healthy volunteers by Mann-Whitney U test revealed that stroke patients had significantly lower TSPO expression at scan 1 (p=0.001 and p=0.002). However, no significant differences were found for scan 2 (p=0.11 and p=0.52) and scan 3 (p=0.45 and p=0.31). Between scan 1 and 3, the mean  $V_T$  in the ipsilesional cerebellum and the perilesional occipital cortex significantly increased by 72.3% and 51.4% (p=0.01 and p=0.002) for the HAB patients, respectively. Results are presented graphically in Figure 4 C and D.

# **DISCUSSION**

# TSPO evolution after stroke and TSPO as a biomarker of recovery

This study shows that the TSPO expression after stroke is a particularly dynamical and heterogeneous process. In human stroke, eleven previous publications have reported on TSPO expression after stroke (Ramsay *et al.*, 1992; Gerhard *et al.*, 2000, 2005; Pappata *et al.*, 2000; Price, 2006; Radlinska *et al.*, 2009; Thiel *et al.*, 2010; Gulyas *et al.*, 2012; Gulyás *et al.*, 2012; Feng *et al.*, 2014; Ribeiro *et al.*, 2014). Five of these studies have included longitudinal TSPO imaging in a total of 25 stroke patients (Boutin and Pinborg, 2015), reporting negligible TSPO upregulation in the first 72 hours after stroke in the lesional and perilesional areas, followed by a significant increase at day 5 (Price, 2006). A further increase was observed in the lesioned hemisphere at day 13-20 and at later time points, between 3-4 weeks and 6-8 months, TSPO levels have been demonstrated to decrease at the lesion site (Ramsay *et al.*, 1992). The current is in agreements with previous findings as it shows initial high TSPO expression lesionally and perilesionally between 1-2 weeks after stroke and a gradual decrease at 3-4 weeks and 15-18 weeks. However, it is also very apparent that the initial increase in TSPO expression is very heterogeneous in both intensity and size and that the heterogeneity of the TSPO expression prevails throughout the entire study period (Figure 2 and 3).

Prior to the initiation of this study, we hypothesized that the initial perilesional TSPO expression would be a biomarker of poor recovery and that the initial TSPO expression in ROIs connected to the lesion would be a biomarker of good recovery. In our study we found no correlation between the binding of <sup>123</sup>I-CLINDE to TSPO in the examined ROIs at scan 1 and recovery in

the follow-up clinical test scales. An explanation for this negative finding may be that the TSPO expression after human stroke is complex and cannot be directly associated to functional recovery because that it reflects a combination of pro- and anti-inflammatory properties in the glia and macrophages with upregulated TSPO. A tracer with selectivity to the pro-inflammatory or anti-inflammatory properties of glia would be advantageous in this case, however, such tracer does not currently exist.

Another explanation may be ceiling and floor effects of rehabilitation measures which are commonly known issues when studying stroke recovery, especially for minor to moderate strokes (Lindemann *et al.*, 2012). These effects can potentially hamper the measurement of clinical progress and thus bias the results.

## TSPO expression in MCA stroke compared to healthy volunteers

To our knowledge, this is the first study to show low TSPO expression in non-stroke related areas in the subacute phase after stroke in man, followed by a longitudinal TSPO normalization of TSPO expression as compared to healthy volunteers.

Several possible biological hypotheses can be made for this finding.

Reactive microglia and macrophages release pro-inflammatory and anti-inflammatory cytokines like IL-10,  $TGF_{\beta}$  and IGF (Patel *et al.*, 2013), however, the spatial distribution in the brain of these cytokines in relation to stroke are not well known. It may be that anti-inflammatory cytokines are expressed more predominantly than pro-inflammatory cytokines in regions remote to the lesion. Post stroke immunomodulation is a known phenomenon occurring after stroke and is increase susceptibility to infections (Samary *et al.*, 2016).

Another explanation for initial low TSPO expression in ROIs unrelated to the lesion compared to healthy volunteers, is that the microglial cells could have redistributed within the brain and migrated to the site of the lesion (Kettenmann *et al.*, 2011). However, TSPO is not expressed to a large extent in resting microglia cells, and would not contribute markedly to the background TSPO signal in healthy brain (Chen and Guilarte, 2008).

This finding infers that one should be very careful with using a supposedly non-affected brain region (eg. cerebellum or unaffected hemisphere) as a reference for quantification of the TSPO expression. Nearly 30% of stroke patients develop depression in the initial weeks after stroke onset (Paolucci, 2008). An interesting biological mechanism for future study may be the link between decreased TSPO expression after stroke and post stroke depression.

## TSPO expression and lesion necrosis

A negative correlation between percentage overlap of SPECT and structural lesion delineated on T2 weighted MRI at scan 1 and the volume of lesion necrosis at scan 3 was discovered. This corresponds to previous findings in animal studies where TSPO is expressed primarily lesionally in transient ischemia whereas in cases of permanent ischemia, TSPO expression occurs predominantly in the outer infarct margins and peri-lesional areas and to a lesser extent in the infarct core (Boutin and Pinborg, 2015).

### Limitations

The groups of healthy volunteers and stroke patients are not matched by age with a median patient age of 61 and a median healthy volunteer age of 34. Previously, TSPO imaging with <sup>11</sup>C-PK11195 has demonstrated increasing TSPO expression with age in a cohort of 25 healthy subjects including 10 children (Kumar *et al.*, 2012). An age-related increase in TSPO expression would cause a smaller difference between stroke patients and healthy volunteers in the case of the current study where the volunteer group is younger than the patient group. Two of the patients had delayed centrifugation of blood samples. This will possibly cause an overestimation of V<sub>T</sub> due to an under-estimation of the plasma parent compound as previously described (Feng *et al.*, 2016). However, this issue would not accentuate the difference between patients and healthy volunteers as all blood samples for healthy volunteers were centrifuged immediately. Hence, the actual decrease in non-affected brain regions of stroke patients is likely lower than what the current results show.

# CONCLUSION

In conclusion, the current study shows that the TSPO expression is an remarkably temporally and spatially dynamic and complex process but not a useable biomarker for functional recovery after stroke. TSPO expression in regions unrelated to the stroke are reduced two weeks after stroke compared to similar regions in healthy volunteers and normalizes within the first 4 months. This may be a proxy to the peripheral immunomodulation known to occur after ischemic stroke or the release of anti-inflammatory cytokines from glial cells in the lesional region.

# **DISCLOSURES**

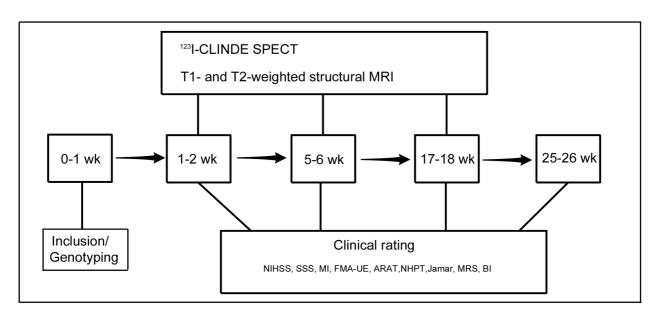
This work was financially supported by the European Union's Seventh Framework Programme (FP7/2007-2013) under grant agreement n° HEALTH-F2-2011-278850 (INMiND), the Danish Council for independent Research, the Reseach Committee of Rigshospitalet, and Savværksejer Jeppe Juhl og hustru Ovita Juhls Mindelegat.

# **ACKNOWLEDGEMENTS**

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# Figures and Tables

Figure 1. Study design



Patients were recruited and genotyped for the rs6971 polymorphism in the first days after stroke. Scanning with <sup>123</sup>I-CLINDE SPECT was performed at 1-2, 5-6, and 17-18 weeks after stroke. Clinical rating was performed at scan days and at follow-up 25-26 weeks after stroke. The scanning of healthy volunteers is not included in this figure. Abbreviations: Wk: Week NIHSS: National Institute of health stroke scale, MI: Motricity index, FMA-UE: Fughl-Meyer assessment

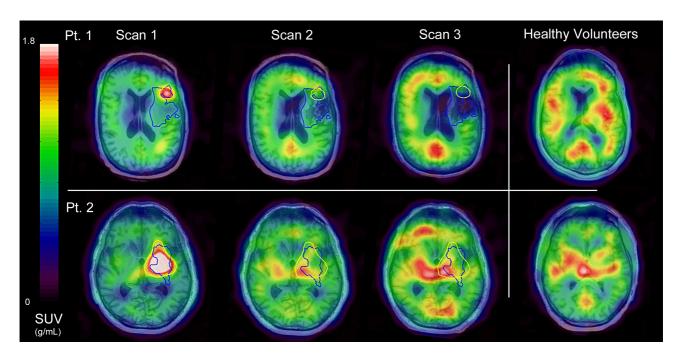


Figure 2. <sup>123</sup>I-CLINDE SPECT and structural MRI in stroke patients

T1-weighted MRI overlaid by <sup>123</sup>I-CLINDE SPECT in patient no. 1 (HAB) and 2 (HAB) scanned longitudinally three times after left MCA stroke and compared to two healthy HAB volunteers. For each row, scan 2, 3 and the healthy volunteer scan were coregistered to scan 1. All SPECT images were normalised by bodyweight and injected <sup>123</sup>I-CLINDE dose. Yellow ROIs depict the lesional SPECT upregulation and blue ROIs depict the structural lesion delineated on T2-weighted MRI (not shown) on scan 1.

Both patients demonstrated low TSPO expression in cortical regions not related to the stroke at scan 1, gradually increasing towards healthy control intensities at scan 3.

Pt. 1 demonstrated a small volume of TSPO upregulation in the infarct margin at scan 1 and the predominant part of the lesion was necrotized at scan 3, probably as a result of permanent ischemia. Pt. 2 demonstrated high lesional and perilesional TSPO expression at scan 1, and only a small part of the lesion was necrotized at scan 3, possibly as a result of better perfusion of the lesional area.

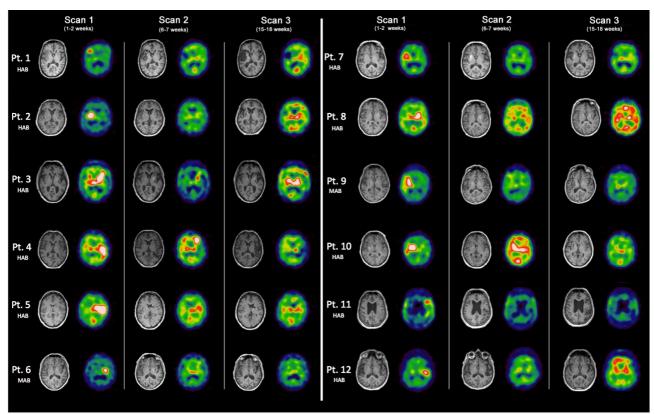
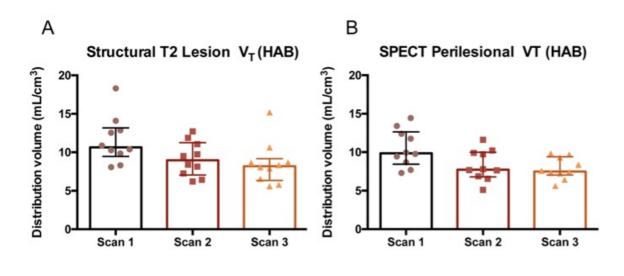
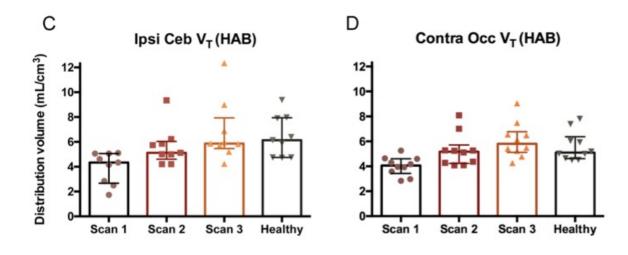


Figure 3. T1-Weighted MRI and <sup>123</sup>I-CLINDE SPECT comparison of all patients. For each patient, the <sup>123</sup>I-CLINDE SPECT is normalised by weight and injected <sup>123</sup>I-CLINDE dose.

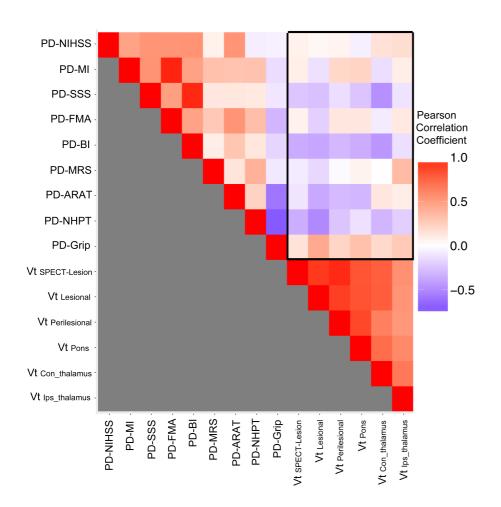
Figure 4. Evolution of TSPO expression and comparison to healthy volunteers





An illustration of  $V_T$ 's in ten HAB patients and ten HAB healthy volunteers. Columns represent median values and brackets span the interquartile range. Longitudinally,  $V_T$ 's decreased in the structural lesion delineated on T2-weighted MRI (A) and in the perilesional SPECT ROI (B). In brain regions unrelated to the stroke  $V_T$ 's in patients were lower than for healthy volunteers at scan 1 and increased towards normal values at scan 3 for the ipsilesional cerebellum (C) and contralesional occipital cortex (D).





Pearson correlation coefficient plot between the ROI  $V_T$ 's at 1-2 weeks after stroke and the PD of rehabilitation measures between 1-2 weeks and 24-25 weeks. No correlation was significant. Abbreviations: Percentage difference (PD), National Institutes of Health Stroke Scale (NIHSS), Scandinavian Stroke Scale (SSS), Fugl-Meyer Assessment of the Upper Extremity (FMA-UE), Motricity Index (MI), Action Research Arm Test (ARAT), Nine-Hole Peg Test (NHPT), Grip strength (Grip), Modified Ranking Scale (MRS) and Barthel Index (BI).

**Table 1. Patients** 

| Patient no. | Days<br>after<br>stroke | Gender | Age at inclusion (Years) | lesion<br>side /<br>Artery | Etiology                  | TSPO<br>Genotype | Injected activity (MBq) | Centrifugation |
|-------------|-------------------------|--------|--------------------------|----------------------------|---------------------------|------------------|-------------------------|----------------|
| Pt. 1       | 10                      |        |                          | T 0                        | TCA 1:                    |                  | 108.9                   | Delayed        |
|             | 38                      | F      | 52                       | Left<br>MCA                | ICA dissection (Trauma)   | HAB              | 113.4                   | Delayed        |
|             | 101                     |        |                          | 141071                     | (Trauma)                  |                  | 103.1                   | Delayed        |
| Pt. 2       | 10                      |        |                          | Ι. Ο                       | TCA 1: 4:                 |                  | 112.3                   | Delayed        |
|             | 46                      | M      | 63                       | Left<br>MCA                | ICA dissection<br>- (HT)  | HAB              | 102.5                   | Delayed        |
|             | 157                     |        |                          | 1,1011                     | (111)                     |                  | 117.5                   | Immediate      |
| Pt. 3       | 12                      |        |                          | 1.0                        | TEL 1                     |                  | 135.7                   | Immediate      |
|             | 40                      | M      | 57                       | Left<br>MCA                | Thrombo-<br>embolism (HC) | HAB              | 114.8                   | Immediate      |
|             | 130                     |        |                          | 1,1011                     | emoonsm (110)             |                  | 126.8                   | Immediate      |
| Pt. 4       | 15                      |        |                          | Ι. Ο                       | Thrombo-                  |                  | 110.9                   | Immediate      |
|             | 43                      | M      | 60                       | Left<br>MCA                | embolism                  | HAB              | 115.9                   | Immediate      |
|             | 139                     |        |                          |                            | (HT+HC)                   |                  | 128.3                   | Immediate      |
| Pt. 5       | 11                      |        |                          | D: 1.4                     | Thrombo-                  |                  | 127.3                   | Immediate      |
|             | 39                      | F      | 42                       | Right<br>MCA               | embolism                  | HAB              | 114.7                   | Immediate      |
|             | 131                     |        |                          |                            | (Contraception)           |                  | 108.9                   | Immediate      |
| Pt. 6       | 9                       |        |                          | D: ~1.4                    | ICA diametica             |                  | 121.5                   | Immediate      |
|             | 35                      | M      | 68                       | Right<br>MCA               | ICA dissection<br>- (HC)  | MAB              | 123.2                   | Immediate      |
|             | 135                     |        |                          |                            | ()                        |                  | 113.0                   | Immediate      |
| Pt. 7       | 7                       |        |                          | Diaht                      | Thrombo-                  |                  | 118.9                   | Immediate      |
|             | 41                      | F      | 72                       | Right<br>MCA               | embolism                  | HAB              | 117.7                   | Immediate      |
|             | 126                     |        |                          |                            | (AF)                      |                  | 108.3                   | Immediate      |
| Pt. 8       | 13                      |        |                          | D: als4                    | Thrombo-                  |                  | 117.8                   | Immediate      |
|             | 41                      | F      | 68                       | Right<br>MCA               | embolism -                | HAB              | 122.1                   | Immediate      |
|             | 125                     |        |                          |                            | (HC+HT)                   |                  | 103.2                   | Immediate      |
| Pt. 9       | 10                      |        |                          | Left                       | Thrombo-                  |                  | 137.0                   | Immediate      |
|             | 37                      | F      | 59                       | MCA                        | embolism (HC)             | MAB              | 117.4                   | Immediate      |
|             | 121                     |        |                          |                            |                           |                  | 122.1                   | Immediate      |
| Pt. 10      | 13                      |        |                          | Diaht                      | ICA dissection            |                  | 123.7                   | Immediate      |
|             | 41                      | F      | 55                       | Right<br>MCA               | - (HT)                    | HAB              | 130.5                   | Immediate      |
|             | 132                     |        |                          |                            |                           |                  | 111.3                   | Immediate      |
| Pt. 11      | 10                      |        |                          | ΙcΦ                        | Thrombo-                  |                  | 113.6                   | Immediate      |
|             | 38                      | M      | 61                       | Left<br>MCA                | embolism                  | HAB              | 112.8                   | Immediate      |
|             | 122                     |        |                          |                            | (HT)                      |                  | 114.4                   | Immediate      |
| Pt. 12      | 10                      |        |                          | D: 1 (                     | Thrombo-                  |                  | 117.6                   | Immediate      |
|             | 44                      | M      | 62                       | Right<br>MCA               | embolism                  | HAB              | 124.9                   | Immediate      |
|             | 122                     |        |                          |                            | (AF)                      |                  | 109.9                   | Immediate      |

Demographic data for the stroke patients. Abbreviations Pt: Patient, MCA: Middle cerebral artery, HT: Hypertension, HC: Hypercholesterolemia, AF: Atrial fibrillation, HAB: High affinity binder, MAB: Mixed affinity binder.

**Table 2. Healthy volunteers** 

| Volunteer no. | Gender | Age at inclusion (Years) | Injected<br>activity<br>(MBq) |
|---------------|--------|--------------------------|-------------------------------|
| HV 1          | M      | 49                       | 124.9                         |
| HV 2          | M      | 36                       | 119.9                         |
| HV 3          | F      | 48                       | 104.5                         |
| HV 4          | M      | 25                       | 112.5                         |
| HV 5          | F      | 34                       | 115.3                         |
| HV 6          | F      | 44                       | 116.2                         |
| HV 7          | F      | 30                       | 105.8                         |
| HV 8          | F      | 34                       | 118.4                         |
| HV 9          | F      | 24                       | 128.2                         |
| HV 10         | F      | 21                       | 120.9                         |

Healthy volunteer data: Gender, age and injected activity for the healthy volunteers.

Table 3. Descriptive data

| Descriptive statistics                               | Unit               | Median | Quartiles<br>[1st;3rd] | Range       |
|--|--------------------|--------|------------------------|-------------|
| Patient age  | Years              | 60.7   | [55.9;66.7]            | [41.7;71.7] |
| Healthy volunteer age                                | Years              | 33.6   | [24.3;45.5]            | [21.4;49.0] |
| NIHSS at stroke onset                                | Points             | 17.5   | [13.3;19.8]            | [8;22]      |
| NIHSS at scan 1                                      | Points             | 6.5    | [4.3;10.0]             | [1;12]      |
| NIHSS at follow-up                                   | Points             | 1.5    | [1.0;4.5]              | [0;5]       |
| T2 lesion volume at scan 1                           | mL                 | 40.5   | [25.1;63.8]            | [17.5;76.1] |
| T1 necrosis volume at scan 3                         | mL                 | 4.7    | [3.1;15.9]             | [2.0;50.8]  |
| Overlap: SPECT Lesion/ T2 Lesion                     | %                  | 68.8   | [51.6;80.0]            | [6.4;90.4]  |
| V <sub>T</sub> lesional scan 1 (HAB)                 | mL/cm <sup>3</sup> | 10.2   | [8.6;12.6]             | [7.5;15.0]  |
| V <sub>T</sub> lesional scan 2 (HAB)                 | mL/cm <sup>3</sup> | 8.0    | [6.3;10.5]             | [5.4;11.0]  |
| V <sub>T</sub> Lesional scan 3 (HAB)                 | mL/cm <sup>3</sup> | 7.1    | [6.2;7.8]              | [4.6;8.5]   |
| V <sub>T</sub> Con Occ cortex scan 1 (HAB)           | mL/cm <sup>3</sup> | 4.1    | [3.4;4.6]              | [2.8;5.3]   |
| V <sub>T</sub> Con Occ cortex scan 2 (HAB)           | mL/cm <sup>3</sup> | 5.2    | [4.2;5.7]              | [4.1;8.1]   |
| V <sub>T</sub> Con Occ cortex scan 3 (HAB)           | mL/cm <sup>3</sup> | 5.8    | [5.1;6.8]              | [4.3;9.1]   |
| V <sub>T</sub> Occ cortex (healthy volunteers) (HAB) | mL/cm <sup>3</sup> | 5.1    | [4.6;6.4]              | [4.5;7.8]   |

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Expression, evolution and prognostic value of Translocator protein in middle cerebral artery stroke patients - A longitudinal [123I]CLINDE-SPECT imaging study (15 authors)

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